

Biological and Water Quality Monitoring

Prepared for City of Ryde

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Cover Image
Shrimptons Creek by Nathan Harrison

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Executive Summary

The Autumn 2011 report completes the seventh and final year of the City of Ryde Council's Biological and Chemical Water Quality Monitoring Strategy. During this period, Sydney Water (and other contractors) collected macroinvertebrate and water chemistry samples from five creek systems of the Ryde Local Government Area (LGA). These were Archers, Shrimptons, Buffalo, Porters and Terrys creeks.

Water quality results for Autumn 2011 indicated Archers, Shrimptons, Buffalo, Porters and Terrys creeks did not meet, on all or virtually all sampling occasions, ANZECC (2000) guidelines for the protection of aquatic ecosystems for total oxidised nitrogen, total nitrogen, ammonium (NH₄) and dissolved oxygen. However, concentrations varied between sites on each creek and between the five creeks. The ANZECC (2000) recommended concentration for faecal coliforms was exceeded at one site in Buffalo Creek and one site in Porters Creek.

The impaired macroinvertebrate communities recorded in each of the five study streams reflect the poor water quality highlighted in the comparison of results to ANZECC (2000) guidelines and, probably, other unmeasured parameters. Water quality results of the monitoring program suggest that while some similarity exists between the five creeks, influences on water chemistry are not the same across the City of Ryde LGA.

A total of 2,044 macroinvertebrates were collected from the edge habitat of these creek systems in Autumn 2011. 46 different taxa were recorded from a total of 78 taxa that have been collected throughout the monitoring program.

Macroinvertebrate results from Autumn 2011 indicate that Archers, Shrimptons, Buffalo, Porters and Terrys creeks had impaired macroinvertebrate communities. These results are reflective of what has been observed previously during the monitoring program. Of the five creeks, Archers Creek appeared to have the richest stream health and Shrimptons Creek the poorest. Stream health is, however, similar across the five creeks. EPT taxa were found in low numbers and abundances with only five families collected throughout the monitoring program.

Multivariate analysis indicated a trend of slight changes in macroinvertebrate community composition between sampled seasons for each creek. Shrimptons Creek has provided the most variability in community structure during the period of the monitoring program, while Terrys Creek has provided the most stable.

BIOENV results returned only weak to mild correlations. The strongest correlation for all five creeks consisted of total oxidised nitrogen, conductivity, dissolved oxygen, ratio of total length of pipe/catchment area, and ratio of number of outlets/catchment area.

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1 Introduction

Sydney Water has developed this report in response to engagement under the City of Ryde Council Tender Number COR-RFQ-29/09. This report for the Autumn 2011 period forms part of the City of Ryde Council's Biological and Chemical Water Quality Monitoring Strategy.

Under the strategy, Sydney Water carried out macroinvertebrate and water quality sample collection, analysis and reporting for the five creek systems of the Ryde LGA. This strategy is a seven-year program in which all five creeks would be monitored for the first two years. For the remaining five years the intention was to target two of the five creeks each year on a rotational basis. In Spring 2006, Ryde Council agreed to continue regular monitoring of all five sites for the remainder of the program after discussions during the Spring 2006 presentation. This would more accurately measure natural variations in stream health during drier and wetter hydrological conditions and set a better baseline for management decisions across all creek catchments.

This Autumn 2011 report ends the seventh year of the program. Macroinvertebrates and water chemistry were each sampled in March and April 2011 at all five sites. Further water quality monitoring was conducted at an additional eight sites.

Monitoring macroinvertebrates and water chemistry enables the City of Ryde Council to:

- Evaluate chemical and biological water quality monitoring for short and long term interpretation and temporal evaluation of creek health over the duration of the strategy
- Detail where, when and how often samples should be taken from creeks within the Ryde Local Government Area based on existing site data, catchment position, accessibility and trends identified
- Prescribe how to sample for macroinvertebrates at each site, building on the standard protocols designed by AUSRIVAS
- Provide a series of options for identification of key indicator taxa to family and/or morphospecies
- Identify suitable indices such as SIGNAL SF to assess water quality, including calculation of the Observed/Expected (OE50 and OE0 SIGNAL2) ratios from the respective AUSRIVAS predictive models for autumn, spring, and combined seasons
- Provide the basis for an appraisal of a standard monitoring strategy to be integrated into a community monitoring program such as Streamwatch
- Provide the foundation to augment the Streamwatch capacity within the City of Ryde, including options for improved education awareness of water quality issues within schools and community groups.
- Provide information and direction on potential infrastructural works to complement water quality monitoring and improve overall creek health.

2 Study Area

The five designated sites (Figure 1) of the City of Ryde Council’s Biological and Chemical Water Quality Monitoring Strategy are:

- Site 1: Terrys Creek near M2 motorway at the end of Somerset Road, North Epping
- Site 2: Shrimptons Creek at Wilga Park
- Site 3: Porters Creek, sampled after the creek exits the Ryde depot
- Site 4: Buffalo Creek, accessed through private property (52 Higginbotham Rd)
- Site 5: Archers Creek at Maze Park

Additional water quality sites for Shrimptons, Porters and Buffalo creeks were sampled for various analytes in Autumn 2011. Refer to Table 8 for these locations.

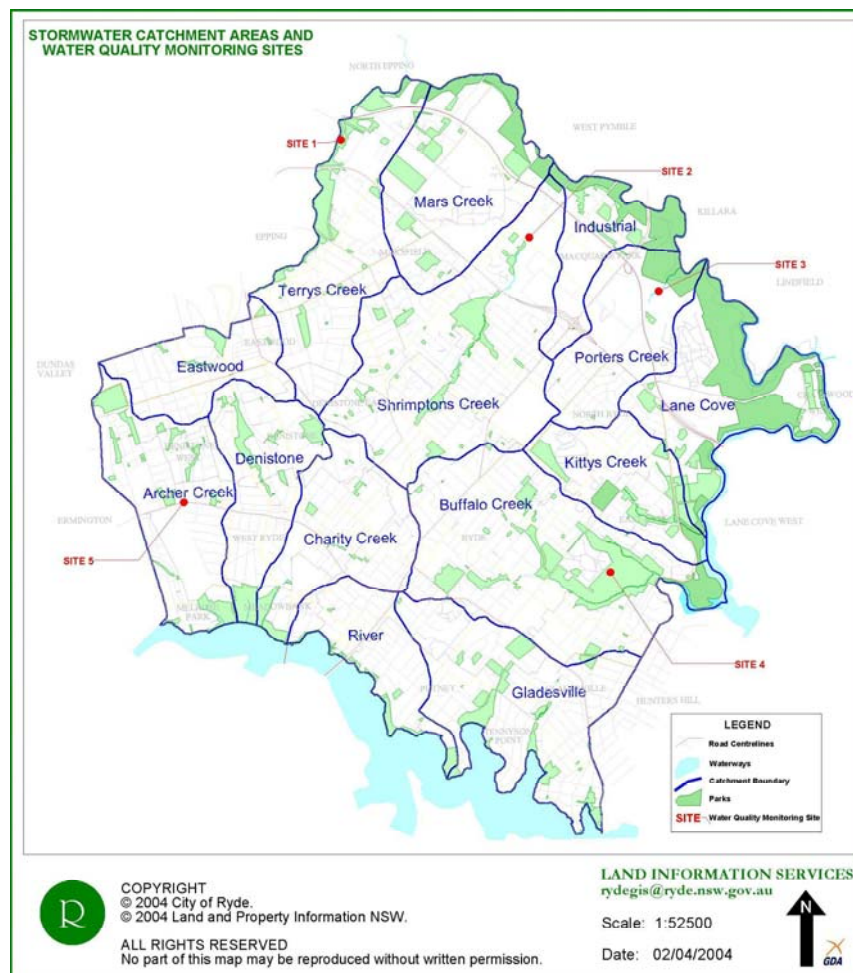


Figure 1 Site locations in the City of Ryde’s Biological and Chemical Water Quality Monitoring Strategy

2.1 Autumn 2011 sampling events

Two sampling events were conducted in Autumn 2011 for the City of Ryde Biological and Chemical Water Quality Monitoring Strategy. Sampling was conducted at all five creeks in each of the following periods:

- March 2nd, 3rd and 4th 2011
- April 8th and 12th 2011



Figure 2 Archers Creek in Autumn 2008 showing completed rehabilitation work

3 Methodology

3.1 Macroinvertebrate sampling

Rapid assessment macroinvertebrate sampling was conducted in accordance with AUSRIVAS protocols for NSW (Turak et al., 2004). Sydney Water staff that conducted field sampling have met criteria of the in-house test method for macroinvertebrate identification and enumeration. Use of experienced staff addresses issues identified by Metzeling et al. (2003).

Three edge habitat samples were collected from each site within a pre-selected area in March and April during the Autumn 2011 season, as specified in the City of Ryde Biological and Chemical Water Quality Monitoring tender document COR-RFQ-29/09. The 'edge' habitat is defined as areas with little or no current. These areas were sampled with a hand-held dip net with 320 mm by 250 mm opening and 0.25 mm (250 µm) mesh that conformed to ISO 7828-1985 (E). The net was swept from open water towards the shore, working over a bank length of about 10 m moving in an upstream direction. In the process, deposits of silt and detritus on the stream bottom were stirred up so that benthic animals were suspended and then caught in the net.

The net contents were then emptied into a large white sorting tray with a small amount of water to allow live macroinvertebrate specimens to be picked out with fine forceps and pipettes for a period of 40 minutes. If new taxa are collected between 30 and 40 minutes, sorting will continue for a further 10 minutes. If no new taxa are found after 10 minutes the picking ceases. If new taxa are found, the 10-minute processing cycle is continued up to a maximum total sorting time of 1 hour. There is no set minimum number of animals collected using the NSW protocols (Turak et al., 2004).

All specimens collected were preserved in small glass jars containing 70% ethanol with a clear label indicating site code, creek name, date, habitat and name of Sydney Water staff sampler. Sampling equipment was washed thoroughly between samples to prevent the cross contamination of animals.

3.2 Macroinvertebrate sample processing

Macroinvertebrates were identified and enumerated to the family taxonomic level, except for: non-biting midges (Chironomids) to sub-family; aquatic worms to Class Oligochaeta; and aquatic mites to Order Acarina. The method used, SSWI433 *In-house test method macroinvertebrate cataloguing, identification and counting*, is in compliance with the requirements of AS ISO/IEC 17025 *General Requirements for the Competence of Testing and Calibration Laboratories* under technical accreditation number 610 issued by National Association of Testing Authorities (NATA) and has been employed since 1997. In particular, macroinvertebrate identification was performed using appropriate published keys listed in Hawking (2000), internal keys to the Sydney Water macroinvertebrate reference collection, unpublished descriptions and voucher specimens.

Quality assurance was conducted as per SSWI434 *In-house test method quality control of macroinvertebrate identification, counting and archiving of collections* in compliance with the requirements of AS ISO/IEC 17025 *General Requirements*

for the Competence of Testing and Calibration Laboratories under technical accreditation number 610. Quality assurance was conducted on 5% of samples collected for this study. Quality assurance is further described in Appendix 1.

3.3 Water quality sampling

Water chemistry was sampled on one occasion in March and April for Autumn 2011 at a similar time to the macroinvertebrate sampling.

Samples were taken by filling the sample bottles directly from the surface of the stream. Temperature, pH and dissolved oxygen were measured on site as per methods summarised in Table 1.

Table 1 Water chemistry parameters, method of analysis in field

ANALYTE	METHOD
pH, Dissolved Oxygen	WTW meter
Temperature	Thermometer

Samples for the analysis of turbidity, conductivity, total dissolved solids (TDS), faecal coliforms, total phosphorus, total nitrogen (as a measure of total oxidised nitrogen and total kjeldahl nitrogen), total alkalinity and ammonia were returned to the laboratory and analysed by the methods summarised in Table 2 within 12 hrs of sampling.

Table 2 Water chemistry parameters, method of analysis in laboratory

ANALYTE	DETECTION LIMIT	METHOD
Turbidity	0.10 NTU	APHA 2130B
Total Dissolved Solids	10 mg/L	APHA 2450 C
Faecal Coliforms	1 cfu/100mL	APHA 9222-D
Total Phosphorus	0.002 mg/L	APHA4500P- H
Alkalinity (CaCO ₃ /L)	0.5 mg/L	APHA 2320 B
Oxidised Nitrogen	0.01 mg/L	APHA 4500-NU43
Total Kjeldahl Nitrogen	0.1 mg/L	Calculation
Ammoniacal Nitrogen	0.01 mg/L	APHA 4500-NU40
Total Nitrogen	0.1 mg/L	APHA 4500-NU57
Conductivity	0.1 mS/m	APHA 2510 B

Additional water quality sample collection and measurements in Autumn 2011 on Shrimptons, Buffalo and Porters creeks allowed spatial comparisons of collected

variables on each creek in an attempt to investigate potential dry weather point sources.

While not sampled at the frequency suggested by ANZECC (2000), the water quality results did allow characterisation of each study creek against ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia) and Recreational Water Quality & Aesthetics (Secondary).

3.4 Rainfall Data

Daily rainfall data from the Marsfield Bureau of Meteorology Station number 066156 are presented where records existed. For the few missing records, data was substituted from West Pymble 66189 and/or Turramurra 66158 Bureau of Meteorology Stations. This was done on the recommendation of the Bureau of Meteorology.

3.5 Comparison with historical data

The City of Ryde Council Tender Number COR-EOC-05/07 requested compilation and analysis of all available and comparable historic raw data back to 2001. This allows a temporal evaluation of ecological health of the five creeks under study. Ecowise supplied raw macroinvertebrate and water chemistry data (Spring 2004 to Autumn 2006). This data, together with seasonal data collected by Sydney Water from Spring 2006 to Autumn 2011, allowed the compilation of data points as summarised in Table 3. Earlier data were unavailable in a suitable format for this purpose or had comparability issues, such as the location sampled on Porters and Buffalo creeks in Spring 2004.

3.6 Data analyses

After identification and enumeration of macroinvertebrates, the data were analysed with univariate and multivariate analysis techniques.

Univariate methods

Data analyses were performed using a number of biological indices and predictive models. These included:

- Diversity index EPT (mayfly, stonefly, caddis fly) richness
- Biotic index SIGNAL-SF
- Output from AUSRIVAS predictive models (Eastern Edge Autumn; Eastern Edge Spring; Combined Edge)
 - AUSRIVAS OE50
 - AUSRIVAS OE0 SIGNAL2

The range of each measure has been plotted in this report with +/-1 standard deviation of the mean for basing ecological decisions (ANZECC, 2000). Presenting data in this way attempts to take account of variation at study sites and provide a basis in future years to enable management tracking and or as a basis for making management decisions.

EPT richness

The diversity index EPT (Ephemeroptera - mayfly, Plecoptera - stonefly and Trichoptera - caddisfly families) richness is based upon the sensitivity of these taxa to respond to changes in water quality condition (Lenat 1988). The number of these taxa found at a site can generally be used as an indicator of stream biological health, although some EPT taxa are more tolerant.

Some caution must be applied when interpreting patterns based on EPT taxa. Many of these macroinvertebrates are sensitive to natural changes in streams, such as altitude. In general, EPT taxa tend to favour higher altitude streams. However, Sydney Water has observed a diverse range of these taxa at altitudes as low as ten metres in undisturbed waterways of the greater Sydney region and in the Clyde River. The absence of these taxa in streams may be attributable to human disturbances within urban catchments and/or a decline in flow over recent years.

SIGNAL-SF

The original version of the Stream Invertebrate Grade Number Average Level (SIGNAL) biotic index (Chessman, 1995-Sydney Water data) was refined to include the response of SIGNAL to natural and human influenced (anthropogenic) environmental factors (Growth et al. 1995), variations in sampling and sample processing methods (Growth et al. 1997; Metzeling et al. 2003) and setting sensitivity grades of the taxa objectively (Chessman et al. 1997; Chessman et al. 2002). "S" indicates Sydney region version and "F" indicates taxonomy is at the family level. SIGNAL-SF has been derived from macroinvertebrate data of the greater Sydney region (Chessman et al. 2007). Water quality status of clean water has been established in the index using data from near pristine reference sites in the bushland fringes of Sydney by using the 10th percentile of the average score

of these reference sites. SIGNAL-SF allows a direct measure of test site condition and incorporates abundance information from the rapid assessment sampling.

The first step in calculating a SIGNAL-SF score is applying predetermined sensitivity grade numbers (from 1, tolerant to 10, highly sensitive) to family counts that occur within a location habitat sample. Then multiply the square root transformed count of each family by the sensitivity grade number for that family, summing the products, and dividing by the total square root transformed number of individuals in all graded families. Families that are present in the samples but with no grade numbers available are removed from the calculation of the SIGNAL-SF score for the sample (very few animals). This procedure was repeated for each sample. Calculation then occurs of a location specific average and a measure of variation (plus and minus one standard deviation of the average score) through time as recommended by Australian and New Zealand Water Quality Guidelines for Fresh and Marine Waters (2000) was made to allow stream health comparisons between sampling occasions for each creek and between creeks. Comparisons in this manner allow ranking of stream health as a guide to management decisions.

As aquatic mites (Order Acarina) and aquatic worms (Class Oligochaeta) are left at higher taxonomic levels in the AUSRIVAS protocol, the respective SIGNAL-SF grades of the families of aquatic mites and worms were averaged and used in the calculation of SIGNAL-SF scores for this report.

Arbitrary pollution categories can be assigned (Table 4). Sydney Water has successfully demonstrated the application of this index in stream monitoring of management changes to the sewerage system and subsequent organic pollution responses in creeks from these changes (Besley and Chessman, 2008).

Table 4 Interpretation of SIGNAL-SF scores (Chessman et al., 2007)

SIGNAL-F score	Water quality status
> 6.5	Clean water
5.2-6.5	Possible mild organic pollution
3.8-5.2	Probable moderate organic pollution
< 3.8	Probable severe organic pollution

AUSRIVAS predictive models OE50 output

AUSRIVAS (AUStralian RIVers Assessment System) predictive model is based on the British bioassessment system RIVPACS (River Invertebrate Prediction and Classification System; Wright 1995). The RIVPACS model was modified to suit the environmental conditions present only in Australia (Turak et al. 2004). The AUSRIVAS model is an interactive software package, which uses the macroinvertebrate and environmental data collected from numerous reference river sites across the state of NSW. It is a tool that can quickly assess the ecological health of any river or creek site. Collected macroinvertebrate data are transformed into presence/absence (1 or 0) form, which is also referred to as binary data. The predictor environmental variables required to run for each model

vary as outlined in Tables 5 and 6 but generally include altitude, location (latitude and longitude), stream size characteristics, substratum composition, river alkalinity and rainfall (Turak 2001). These environmental variables allow the software to compare test sites, in this case City of Ryde creek samples, to comparable reference site groups with similar environmental characteristics.

AUSRIVAS models can incorporate data taken from pool edge or riffle habitats. The paucity of riffle habitats at the City of Ryde study sites precludes use of the riffle models. Ecowise collected only four riffle samples between Spring 2004 and Autumn 2006, so only the respective edge models have been employed in comparison of Autumn 2011 data with historical data.

The applicable AUSRIVAS models for comparison of the City of Ryde test creek sites are: the eastern edge Autumn model; eastern edge Spring model; and Combined Season eastern edge model. However, Ecowise (Spring 2005) suggested the later model does not allow changes in condition between seasonal sampling events for the City of Ryde strategy. The later model has been included here for completeness. Ransom et al. (2004) describes this model as preferable, as it maximizes the family list for the test site being examined.

The respective model uses the test site information and comparable reference site group information to calculate a score called the "OE50 ratio" (observed/expected number of macroinvertebrate families with greater than 50% probability of occurring at a test site) (Coysh et al. 2000). The OE50 ratio provides a measure of impairment at a test site (Ransom et al., 2004). The OE50 ratio of each test site sample also corresponds to a band that assists in interpretation and aids management decisions (Coysh et al. 2000). That is, the band helps to categorise each test site showing how it compares with reference sites from rivers of the same type. Interpretation of the five possible bands of river condition is detailed in Table 5 (Coysh et al. 2000). Thresholds that correspond to these bands of each respective model are detailed in Table 6.

Table 5 Interpretation of bands associated with AUSRIVAS OE50 model output (Coysh et al., 2000)

Band	Description	O/E taxa	O/E taxa interpretations
X	More biologically diverse than reference	<ul style="list-style-type: none"> O/E greater than 90th percentile of reference sites used to create the model 	<ul style="list-style-type: none"> More families found than expected Potential biodiversity 'hot spot' or mild organic enrichment Continuous irrigation flow in a normally intermittent stream
A	Similar to reference	<ul style="list-style-type: none"> O/E within range of central 80% of reference sites used to create the model 	<ul style="list-style-type: none"> Expected number of families within the range found at 80% of the reference sites
B	Significantly impaired	<ul style="list-style-type: none"> O/E below 10th percentile of reference sites used to create the model Same width as band A 	<ul style="list-style-type: none"> Fewer families than expected Potential impact either on water and/or habitat quality resulting in a loss of families
C	Severely impaired	<ul style="list-style-type: none"> O/E below band B Same width as band A 	<ul style="list-style-type: none"> Many fewer families than expected Loss of families from substantial impairment of expected biota caused by water and/or habitat quality
D	Extremely impaired	<ul style="list-style-type: none"> O/E below band C down to zero 	<ul style="list-style-type: none"> Few of the expected families and only the hardy, pollution tolerant families remain Severe impairment

Table 6 Upper thresholds for bands of impairment (OE50 taxa) for AUSRIVAS models developed for NSW (Turak and Waddell, 2001)

Model	Threshold			
	A	B	C	D
Combined edge (East)	1.17	0.82	0.48	0.14
Autumn edge	1.17	0.81	0.46	0.11
Spring edge	1.16	0.83	0.51	0.19

Indicator taxa from the AUSRIVAS predictive models output

AUSRIVAS output identifies taxa that were expected from the respective reference site group to which a test site is being compared. As part of this output missing taxa are listed with greater than 50% probability of occurrence. To provide consistency in this report the definition used by Ecowise (2004, 2005a, 2005b, 2006) has been used in this report. That is, indicator taxa are defined as taxa within the EPT (Ephemeroptera - mayfly, Plecoptera - stonefly and Trichoptera – caddisfly) orders with SIGNAL2 scores of greater than 6.

AUSRIVAS predictive models OE0 SIGNAL-2 output

Together with OE50 output, each AUSRIVAS model generates AUSRIVAS OE50-SIGNAL2 values and AUSRIVAS OE0-SIGNAL2 values. This output incorporates SIGNAL2 (Chessman 2003a) tolerance grades derived from reference sites across NSW sampled to create the AUSRIVAS models in NSW. Please note SIGNAL2 tolerance grades are different to the greater Sydney region tolerance grades of SIGNAL-SF, as the later has been derived from sites of the Sydney region and not from broadly across NSW.

An example calculation of AUSRIVAS OE50-SIGNAL2 values was provided in the previous Ecowise reports, this example was sourced from Chessman (2003a). In the Spring 2006 report AUSRIVAS OE50-SIGNAL2 values were found to be quite variable and so it was not recommended for use in future temporal comparisons. The large variation recorded in this measure provided little ability to detect future changes in community structure from future management decisions.

In contrast, AUSRIVAS OE0-SIGNAL2 values were found to have less variance and the recommendation was that it be calculated in Autumn 2007 and beyond. The lesser variation of AUSRIVAS OE0-SIGNAL2 is attributed to the inclusion of taxa with 50% probability of occurrence or more used to calculate AUSRIVAS OE50-SIGNAL2 and additional taxa with less than 50% probability of occurrence.

No bands have been developed for SIGNAL2 (Coysh et al. 2000). However, values of around 1 would be similar to reference condition (Chessman pers comm.). Using AUSRIVAS calculated values are recommended by Chessman (2003a) as a way to overcome natural variation, which is an issue for calculation of SIGNAL2 as described in Chessman (2003b).

Multivariate methods

Data analyses were performed using the PRIMER software package (Clark and Warwick 2001). Analysis techniques included:

- Classification and ordination, SIMPROF test
- SIMPER
- BIOENV

These analysis techniques complement univariate analyses by exploring patterns of macroinvertebrate community structure. Macroinvertebrate community structure at a site can also be referred to as the biological signature. Prior to analysis, the data from the field survey was square root transformed and rare taxa observed in only one sample were removed.

Samples from Autumn 2005 to the final Autumn 2011 season were compared in an ordination for all creeks of the monitoring program to look at context of community composition. Spring 2004 data were not included in these comparisons as comparable sites in Buffalo and Porters creeks were not sampled in Spring 2004, nor were all water quality variables at Archers, Shrimptons and Terrys creeks (Table 3).

Macroinvertebrate data of each creek were also explored by a comparison of data from each sampled season to look at community composition change through time.

Classification, Ordination and SIMPROF test

The group average classification technique was used to place the sampling sites into groups, each of which had a characteristic macroinvertebrate community based on relative similarity of their attributes. Similarities (distances) between the fauna of each pair of sites were calculated using the Bray-Curtis measure, which is not sensitive to rough approximations in the estimation of taxa abundances (Faith et al. 1987), as is the case with rapid assessment sampling. The group average classification technique initially forms pairs of samples with the most similar taxa and gradually fuses the pairs into larger and larger groups (clusters) with increasing internal variability.

Classification techniques will form groups even if the data set actually forms a continuum. The SIMPROF test provides a way to view community structure differences and similarities between samples and overcome the limitation of classification analysis. SIMPROF results have been overlaid onto classification result output (dendrogram), with red lines indicating no difference between samples and the black line indicating a difference in community structure. SIMPROF test groups can be checked against ordination results. Samples were ordinated using the non-metric multidimensional scaling (MDS) technique. Ordination produces a plot of sites on two or three axes such that sites with similar taxa lie close together and sites with a differing taxon composition lie farther apart. When Ordination and SIMPROF test results produce similar overall patterns the analysis can be considered reliable.

Any ordination procedure inevitably introduces distortion when trying to simultaneously represent the similarities between large numbers of samples in only two or three dimensions. The success of the procedure is measured by a stress value, which indicates the degree of distortion imposed. In the PRIMER software package a stress value of below 0.2 indicates an acceptable representation of the original data, although lower values are desirable.

To achieve suitable multivariate representations of data in 2 or 3 dimensions with greater data collection, an analysis strategy to minimize stress (and achieve a better measure of fit) is to pool up macroinvertebrate data of the same season for each creek to produce one data point per season per creek, as demonstrated in the Spring 2007 report for all five creeks. This analysis strategy has been adopted for the ordination plot of all creeks in this report.

SIMPER

The SIMPER routine was employed to investigate community structure between and within groups of sites as detailed above. This routine employs Bray Curtis similarities to examine the contribution of individual taxa to the average similarity between groups and also within groups.

BIOENV

The extrinsic physical and chemical characteristics of the creeks were compared to the intrinsic macroinvertebrate community structure using the BIOENV routine. The underlying similarity matrix was constructed with the normalised Euclidean Distance association measure option. This option enabled a comparison of water quality variables without undue weight being assigned by differing unit scales. Log10 transformations were applied to faecal coliforms, ammonia, oxidised

nitrogen, total phosphorus, total kjeldahl nitrogen, total nitrogen, turbidity, conductivity, and total dissolved solids. All other physical and chemical variables listed in Table 2 were untransformed in the BIOENV analysis.

4 Results

4.1 Water quality and site observations

Water quality results are presented separately for the five creeks with reference to ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia) and Recreational Water Quality and Aesthetics (Secondary). While not to the sampling frequency suggested by ANZECC (2000), the water quality results did allow characterisation of each creek against these guidelines. Historical average refers to data collected at the core site between Spring 2004 and Spring 2010.

Archers Creek

Water quality results for Archers Creek in Autumn 2011 are presented in Table 7. Overall water quality results for the period were similar to previous report periods. Results for faecal coliforms, turbidity and conductivity were within guideline limits on both sampling occasions in Autumn 2011.

Ammonium, total phosphorus and total nitrogen concentrations exceeded their respective guideline values in March. The total phosphorus concentration of 65 µg/L also exceeded the historical average for this site (49 µg/L). The oxidised nitrogen concentration of 190 µg/L exceeded the guideline value (40 µg/L) in April but remained below the historical average of 270 µg/L.

Dissolved oxygen saturation was low on both sampling occasions with results of 75.6% and 52.6% falling below the lower guideline of 85% saturation. The results are consistent with the historical average of 63.9% saturation.

Table 7 Water quality results for Archers Creek Autumn 2011

	Guideline	Core site Maze Park		Historical average
		March	April	
Faecal Coliforms CFU/100mL	1,000 ¹	340	310	790
Ammonia µg/L	20 ²	50	<10	80
Oxidised Nitrogen µg/L	40 ²	10	190	270
Total Phosphorus µg/L	50 ²	65	42	49
Total Kjeldahl Nitrogen µg/L	NA	600	300	400
Total Nitrogen µg/L	500 ²	610	490	669
Alkalinity mg CaCO ₃ /L	NA	82.8	68.7	72
Turbidity NTU	50 ²	2.02	1.14	4
Conductivity µS/cm	125-2,200 ²	404	376	480
Total Dissolved Solids mg/L	NA	240	211	286
pH units	6.8-8.0 ²	7.15	6.98	7.1
Dissolved Oxygen DO % saturation	85-110 ²	75.6	52.6	63.9
Temperature °C	NA	24.8	16.5	17.4

¹ ANZECC (2000) guidelines for Recreational Water Quality & Aesthetics (Secondary)

² ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia)

Shrimptons Creek

Water quality results for Shrimptons Creek in Autumn 2011 are presented in Table 8. Turbidity, pH and conductivity results were within the respective guideline values for all sites in March and April.

The overall water quality of the Bridge Street site was the best of the four sites on Shrimptons Creek. The only exception to meeting the guidelines was for dissolved oxygen saturation, which fell below the lower limit of 85% saturation in both March and April.

The water quality of the core site at Wilga Park was also good only producing exceedences for oxidised nitrogen and dissolved oxygen saturation in both March and April. The oxidised nitrogen concentration in April (190 µg/L) exceeded the historical average (174 µg/L).

The Kent Road site exceeded the guideline values for total phosphorus (186 µg/L) and total nitrogen (1,210 µg/L) in March, oxidised nitrogen (120 µg/L) in April, and dissolved oxygen saturation in March and April (18.7% and 60.8%, respectively).

The Quarry Road site had the poorest water quality of the Shrimptons Creek sites, exceeding the respective guideline values for ammonia (40 and 30 µg/L), oxidised nitrogen (1,110 and 1,070 µg/L), total phosphorus (102 and 168 µg/L) and total nitrogen (1,740 and 1,460 µg/L) in March and April and dissolved oxygen saturation in April (75.8%).

Table 8 Water quality results for Shrimptons Creek Autumn 2011

	Guideline	Core site Wilga Park		Kent Road		Bridge Street (d/s Santa Rosa Park)		Quarry Road (u/s Santa Rosa Park)		Historical average
		March	April	March	April	March	April	March	April	
Faecal Coliforms CFU/100mL	1,000 ¹	400	360	1,000	470	54	270	300	460	1,788
Ammonia µg/L	20 ²	10	<10	20	<10	<10	<10	40	30	37
Oxidised Nitrogen µg/L	40 ²	60	190	10	120	<10	20	1,110	1,070	174
Total Phosphorus µg/L	50 ²	35	40	186	45	14	25	102	168	66
Total Kjeldahl Nitrogen µg/L	NA	340	240	1200	280	380	250	630	390	511
Total Nitrogen µg/L	500 ²	400	430	1,210	400	380	270	1,740	1,460	676
Alkalinity mg CaCO ₃ /L	NA	35.4	38.9	74.7	34	110	59.9	60.1	75.5	65
Turbidity NTU	50 ²	0.75	2.00	1.14	2.05	1.15	1.94	2.08	1.45	8
Conductivity µS/cm	125-2,200 ²	186	196	400	212	1,035	392	939	796	367
Total Dissolved Solids mg/L	NA	113	111	222	122	616	211	546	444	219
pH units	6.8-8.0 ²	7.28	6.98	7.01	6.95	7.14	6.85	7.27	7.06	7.0
Dissolved Oxygen DO % saturation	85-110 ²	68.9	65.4	18.7	60.8	42.1	45.6	86.5	75.8	41.5
Temperature °C	NA	20.6	17.4	20.1	17.6	20.3	17.4	19.3	16.7	17.0

¹ ANZECC (2000) guidelines for Recreational Water Quality & Aesthetics (Secondary)

² ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia)

Buffalo Creek

Water quality results for Buffalo Creek in Autumn 2011 are presented in Table 9. Turbidity, conductivity and pH results were within the respective guidelines at all Buffalo Creek sites in March and April.

Faecal coliform concentrations in Buffalo Creek were generally below the guideline of 1,000 CFU/100mL. The exception to this was Buffalo Creek upstream of Burrows Park in April with a concentration of 1,700 CFU/100mL.

Ammonium concentrations were elevated at the core site in March (40 µg/L). Total nitrogen and oxidised nitrogen concentrations were elevated above the respective guideline values in all samples for April and for oxidised nitrogen at the core site at Higginbotham Road in March.

Dissolved oxygen saturation levels were consistently lower than recommended levels (85 – 110% saturation) in samples from the core site at Higginbotham Road in March and April, at the upstream Burrows Park site in March and the downstream Burrows Creek site in April.

Table 9 Water quality results for Buffalo Creek Autumn 2011

	Guideline	Core site Higginbotham Rd		d/s Burrows Park		u/s Burrows Park		Historical average
		March	April	March	April	March	April	
Faecal Coliforms CFU/100mL	1,000 ¹	40	350	590	950	270	1,700	632
Ammonia µg/L	20 ²	40	20	<10	<10	<10	<10	59
Oxidised Nitrogen µg/L	40 ²	70	210	<10	510	<10	620	310
Total Phosphorus µg/L	50 ²	39	32	44	43	61	50	38
Total Kjeldahl Nitrogen µg/L	NA	410	310	490	300	490	440	381
Total Nitrogen µg/L	500 ²	480	520	490	810	490	1,060	670
Alkalinity mg CaCO3/L	NA	65.8	52.6	105	79.5	102	88.1	79
Turbidity NTU	50 ²	2.18	2.57	0.75	3.08	1.74	0.71	9
Conductivity µS/cm	125-2,200 ²	792	369	1,806	796	1,038	702	687
Total Dissolved Solids mg/L	NA	478	209	1,076	454	628	392	399
pH units	6.8-8.0 ²	7.22	7.12	7.03	7.00	7.36	7.36	7.3
Dissolved Oxygen % saturation	85-110 ²	65.0	70.4	85.8	78.9	71.1	98.5	66.9
Temperature °C	NA	20.8	17.1	22.4	17.9	20.9	17.7	17.3

¹ ANZECC (2000) guidelines for Recreational Water Quality & Aesthetics (Secondary)

² ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia)

Porters Creek

Water quality results for Porters Creek in Autumn 2011 are presented in Table 10. Turbidity, pH and conductivity results were within the respective guidelines at all Porters Creek sites in March and April.

Faecal coliform concentrations in Buffalo Creek were generally below the guideline of 1,000 CFU/100mL. The exception to this was the Main Branch at Wicks Road in April with a concentration of over 10,000 CFU/100mL.

Most total nitrogen, oxidised nitrogen and ammonia concentrations from Porters Creek exceeded the respective guideline levels. This is consistent with the respective historical averages of the core site for Porters Creek. Total phosphorous concentrations exceeded the guideline in April at the Main Branch site (220 µg/L).

Table 10 Water quality results for Porters Creek Autumn 2011

	Guideline	Core Site d/s of Depot		Spur Branch		Main Branch Channel (COR staff site)		Main Branch Wicks Road		Historical average
		March	April	March	April	March	April	March	April	
Faecal Coliforms CFU/100mL	1,000 ¹	240	590	6	38	44	89	390	>10,000	5,612
Ammonia µg/L	20 ²	210	330	<10	270	30	80	<10	<10	795
Oxidised Nitrogen µg/L	40 ²	1,270	930	<10	20	200	280	430	770	1,122
Total Phosphorus µg/L	50 ²	19	34	37	38	18	20	27	220	27
Total Kjeldahl Nitrogen µg/L	NA	610	620	580	1000	260	260	240	610	1,238
Total Nitrogen µg/L	500 ²	1,880	1,550	580	1,020	460	540	670	1,380	2,464
Alkalinity mg CaCO3/L	NA	58.8	77.1	177	2.0	46.6	47.4	80.1	88.7	75
Turbidity NTU	50 ²	1.93	1.08	1.51	3.3	1.11	1.05	1.23	1.64	5
Conductivity µS/cm	125- 2,200 ²	299	389	584	800	239	248	389	513	1,831
Total Dissolved Solids mg/L	NA	184	228	335	492	144	141	221	263	1,110
pH units	6.8-8.0 ²	7.56	7.16	7.46	6.94	7.7	7.14	7.67	7.11	7.6
Dissolved Oxygen % saturation	85-110 ²	90.0	105.0	113.0	36.8	99.0	101.0	82.1	97.6	88.8
Temperature °C	NA	20.2	19.0	21.8	18.1	20.9	19.8	20.2	18.5	17.9

¹ ANZECC (2000) guidelines for Recreational Water Quality & Aesthetics (Secondary)

² ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia)

Terrys Creek

Water quality results for Terrys Creek in Autumn 2011 are presented in Table 11. Results for faecal coliforms, turbidity, conductivity and pH were within guideline limits on both sampling occasions in Autumn 2011.

The ammonium (1,030 µg/L), total nitrogen (1,680 µg/L), total Kjeldahl nitrogen (1,650 µg/L) and total phosphorus (243 µg/L) concentrations for March were extremely high, all exceeding the historical averages for this site. While the concentrations of these analytes had returned to more typical levels in April, the oxidised nitrogen (170 µg/L) concentration increased to exceed the guideline.

Dissolved oxygen saturation levels were below the guideline range on both sampling occasions, although the April result of 84.6% was only slightly below the lower guideline level of 85% saturation.

Table 11 Water quality results for Terrys Creek Autumn 2011

	Guideline	Core Site Sommerset Park		Historical average
		March	April	
Faecal Coliforms CFU/100mL	1,000 ¹	120	210	2,432
Ammonia µg/L	20 ²	1,030	<10	109
Oxidised Nitrogen µg/L	40 ²	30	170	180
Total Phosphorus µg/L	50 ²	243	33	43
Total Kjeldahl Nitrogen µg/L	NA	1,650	260	414
Total Nitrogen µg/L	500 ²	1,680	430	590
Alkalinity mg CaCO ₃ /L	NA	94.5	42.9	59
Turbidity NTU	50 ²	2.72	1.58	6
Conductivity µS/cm	125-2,200 ²	552	243	405
Total Dissolved Solids mg/L	NA	309	138	240
pH units	6.8-8.0 ²	7.25	7.19	7.2
Dissolved Oxygen % saturation	85-110 ²	61	84.6	63.4
Temperature °C	NA	20	16.8	15.5

¹ ANZECC (2000) guidelines for Recreational Water Quality & Aesthetics (Secondary)

² ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia)

4.2 Rainfall data

Daily rainfall data from the Marsfield Bureau of Meteorology Station number 066156 are presented below in Figure 3 including the Autumn 2011 sampling period and preceding five months. In the five months preceding the April 2011 sampling event 387 mm of rainfall occurred within a range of 15 – 134 mm per month. The total annual rainfall recorded for each year of the Water Quality Monitoring Program for the City of Ryde is listed in Table 12. Close to average rainfall conditions were experienced in 2003, 2007, 2008 and 2009 and 2010 and less than average conditions were recorded in 2004, 2005 and 2006.

Table 12 Total rainfall by year

Year	Rainfall (mm)
2003	1,262
2004	905
2005	788
2006	730
2007	1,430
2008	1,203
2009	992
2010	1,249
2011	329 (Until May)

The rainfall in late 2010 was characterised by periods of consistent light to medium falls. This pattern changed in early 2011 when very little rain fell between January and mid March. Consistent light to medium falls from mid March was punctuated by several events of daily rainfall greater than 30 mm.

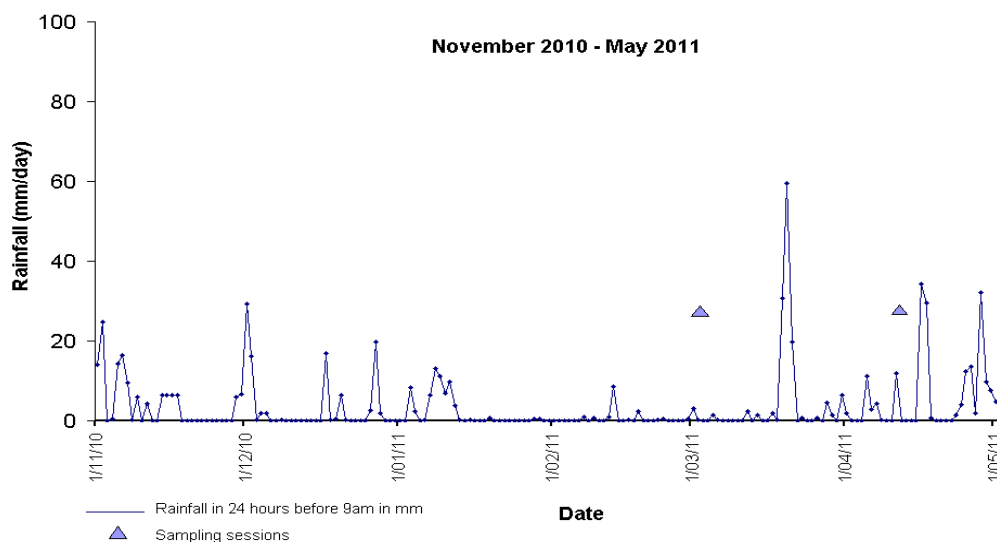


Figure 3 Daily rainfall data 1st November 2010 to 1st May 2011 with sampling occasions indicated

4.3 Macroinvertebrate Results

General Characteristics of Aquatic Macroinvertebrate Assemblages

- A total of 2,044 macroinvertebrates were collected from all five sites in Autumn 2011
- From that, 46 different taxa were recorded
- A total of 78 taxa families have been collected from the edge habitat of all five creeks from Spring 2004 to Autumn 2011
- This compares with 157 taxa of the SIGNAL-SF index of the greater Sydney region, although this total includes taxa from the edge habitat as well as all other stream habitats

The collection of Autumn 2011 samples has seen an increase in the total taxa collected in three of the five creeks, including four at Archers Creek, two at Porters Creek and one at Porters Creek. Terry's Creek has the highest total taxa collected and Shrimptons Creek the lowest during the monitoring program. However, the total taxa are similar for all five creeks (Table 13).

Table 13 Number of taxa recorded in each creek in specified sample periods

Sampling Seasons	Archers	Shrimptons	Buffalo	Porters	Terrys
Spring 04 - Spring 10	55	52	52	56	60
Spring 04 - Autumn 11	59	52	54	57	60

Macroinvertebrate results for comparable samples (Table 3) are consolidated in Appendix 4.

The larvae of the Sydney Hawk dragonfly, *Austrocordulia leonardi* (listed as endangered under the FM Act 1994), and the Adams Emerald dragonfly, *Archaeophya adamsi* (listed as endangered under the FM Act 1994), are potentially found in the Sydney basin region. Neither of these macroinvertebrates was observed in Autumn 2011 samples and is not listed in historical data.

EPT Richness

The average EPT taxa richness has been summarised for each of the five creeks over the monitoring period (Figure 4). This summary indicated that EPT taxa are rarely collected from the five sampled creeks.

Of the five creeks sampled in Autumn 2011, Porters Creek recorded the highest number of EPT taxa with an average of one. All the remaining creeks displayed averages of less than one, with Shrimptons and Terrys Creeks recording zero EPT taxa (Figure 5).

Autumn 2011 results for all creeks fell within what has been previously recorded and had little impact on the average presence of EPT taxa over the sampling period (Figure 4). Porters Creek and Archers Creek showed the highest diversity index, however they still did not average a single EPT taxa. Terrys Creek and Shrimptons Creek displayed the lowest occurrence of EPT taxa for both Autumn sampling and the overall sampled seasons (Figure 4).

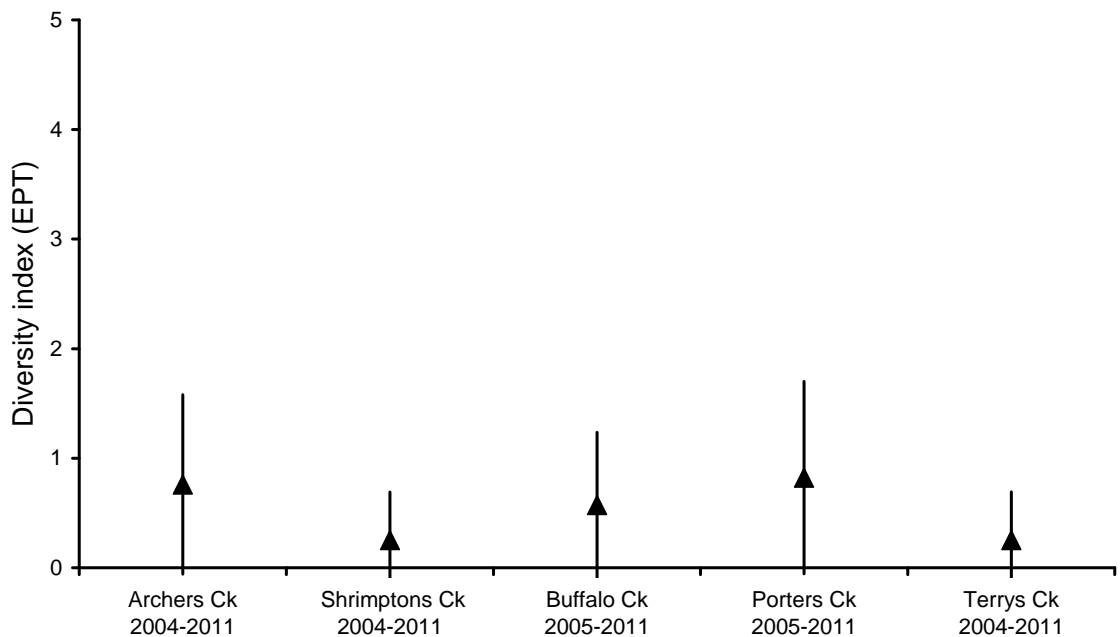


Figure 4 EPT richness of all creeks of monitoring program

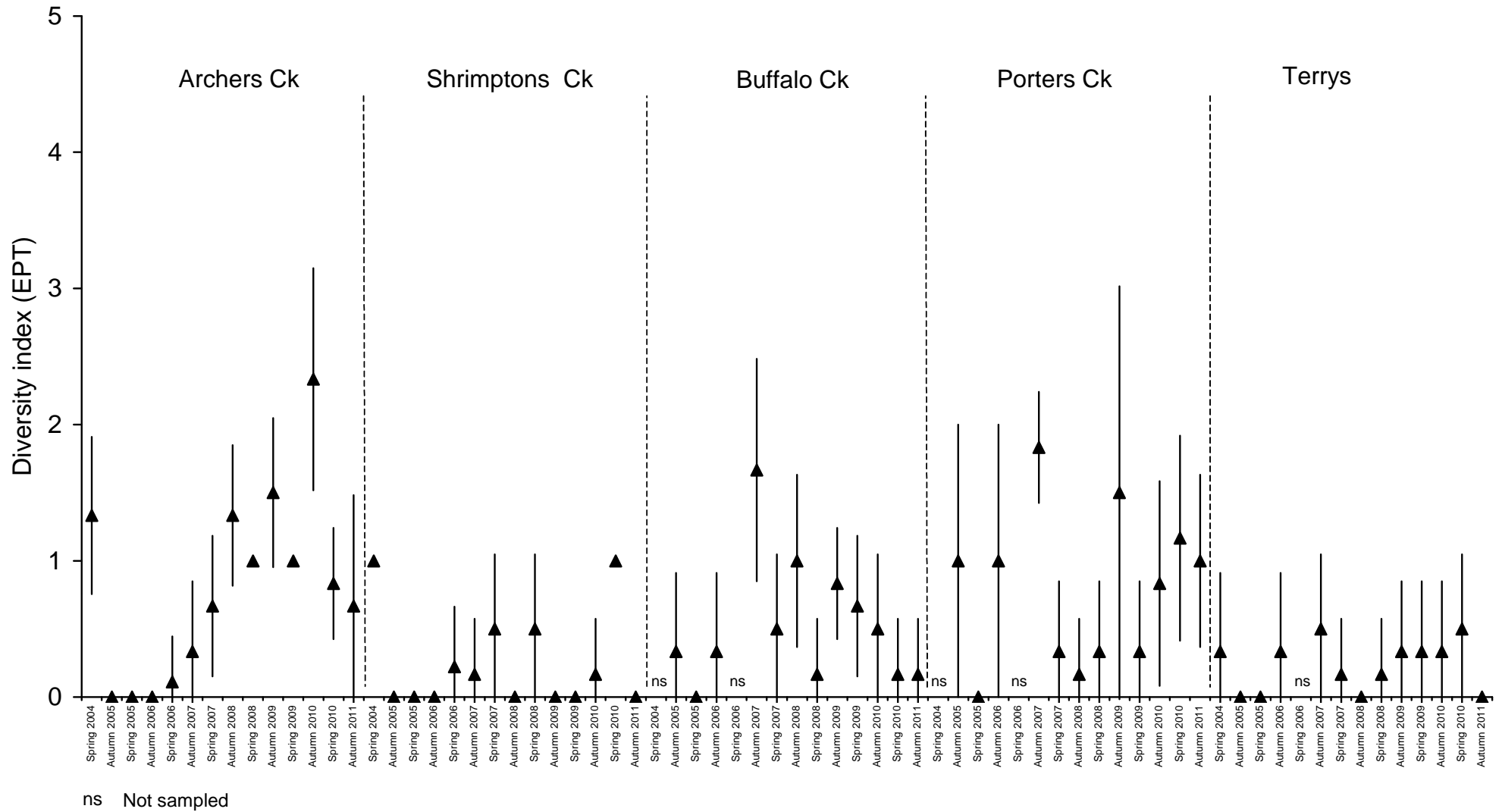


Figure 5 EPT richness by season

SIGNAL-SF

The SIGNAL-SF biotic index indicated impaired macroinvertebrate communities, with average stream health being indicative of probable moderate and probable severe organic pollution for all creeks.

Archers Creek narrowly had the highest average stream health and Shrimptons Creek narrowly the lowest when assessed with SIGNAL-SF for the sampling periods between Spring 2004 and Autumn 2011. When all five creeks are compared in terms of ANZECC (2000) guidelines (± 1 standard deviation of the average), the overlapping ranges of stream health indicate no significant difference exists between the creeks. Shrimptons Creek has the largest range in stream health, which reflects the change in average stream health recorded between Spring 2004 and Autumn 2011 (Figure 6 and Figure 7).

Archers Creek has shown a trend of higher average stream health in Autumn when compared to the respective Spring season. This trend has continued in Autumn 2011 with a slight increase over the previous season. Other than this seasonal variation there has been little change through time in Archers Creek (Figure 7).

Average stream health has shown very little variation in the previous seven sampling seasons for Shrimptons Creek, Autumn 2011 recorded a slight decrease from Spring 2010. Shrimptons Creek average stream health peaked in Autumn 2007 after steadily increasing from Autumn 2005, when it recorded the lowest stream health from all five creeks for all sampling periods (Figure 7).

The average stream health in Buffalo Creek increased slightly in Autumn 2011, which is consistent with the seasonal variations from previous years. The stream health has had little variation through time except for Spring 2008 when average stream health dropped significantly, to its lowest score of the program (Figure 7).

Porters Creek has shown a trend of higher average stream health in autumn when compared to the respective spring season. This trend has continued in Autumn 2011 with a slight increase over the previous season. Stream health has been slightly increasing since Spring 2008. Autumn 2011 has recorded the highest average since first being sampled in Autumn 2005 (Figure 7).

The range of average stream health for Terrys Creek has been very narrow throughout the sampling program and Autumn 2011 falls within what has been previously recorded. There had been a small but consistent decline in average stream health from Spring 2005 to Spring 2008, with the next four seasons having slightly higher average stream health. However, the average stream health has decreased slightly in Autumn 2011, recording the lowest autumn average during the monitoring program (Figure 7).

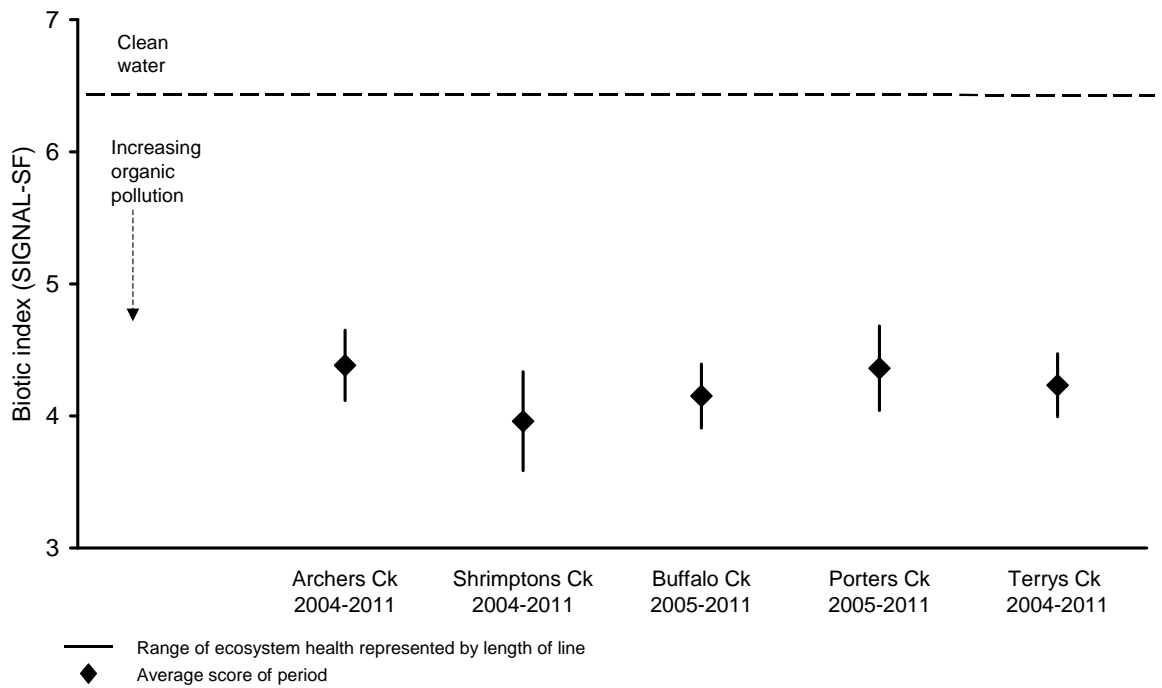


Figure 6 SIGNAL-SF of all creeks of monitoring program

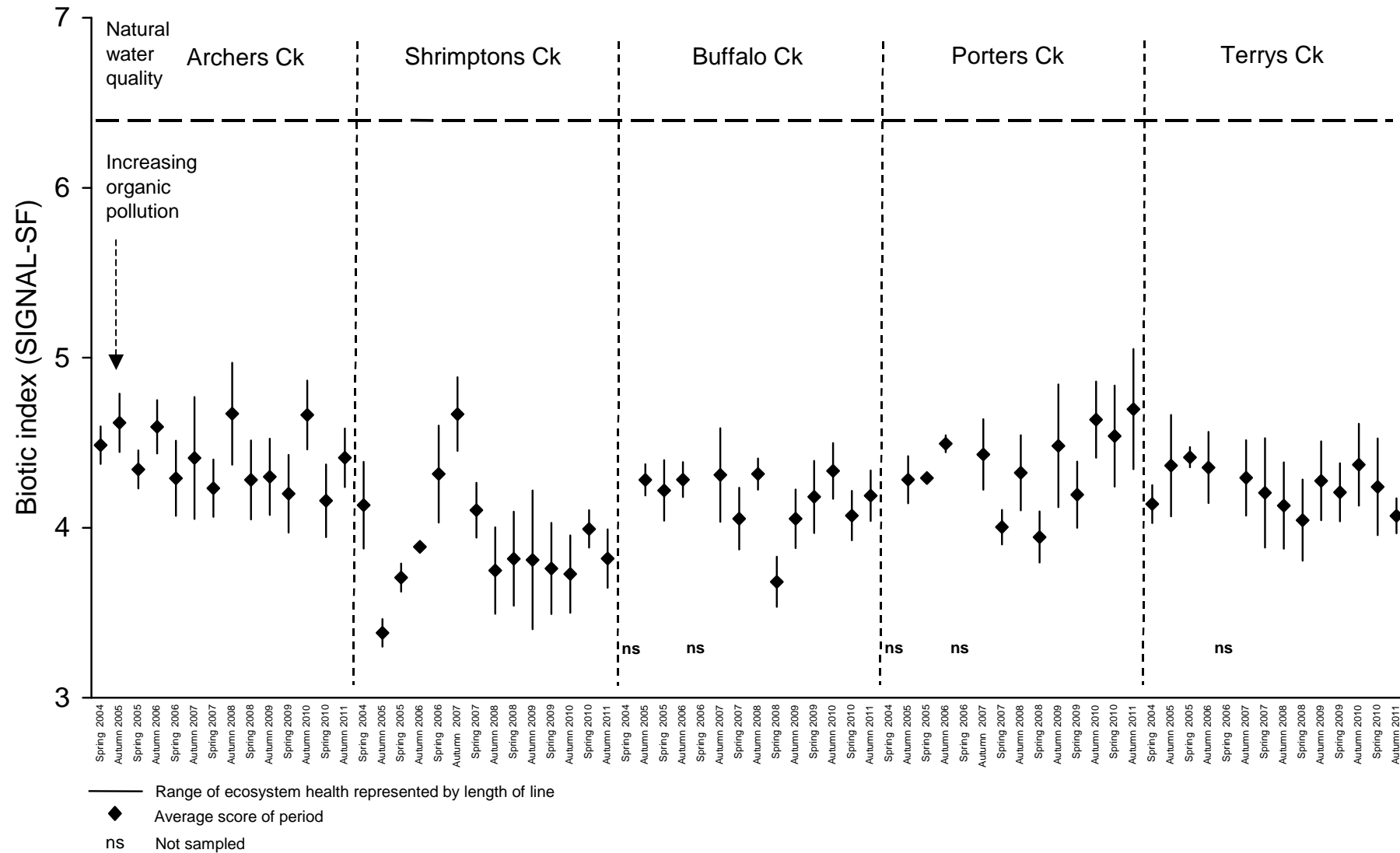


Figure 7 SIGNAL-SF by season

AUSRIVAS OE50

The addition of Autumn 2011 data to the Autumn edge AUSRIVAS OE50 model allowed for the Combined Season edge output to be updated.

The 2011 Autumn edge and 2010/11 combined season model output does not include an OE50 average score for Archers Creek. The model output described it as *outside the experience of the model* (OEM). The same output occurred for both Autumn 2009 and Autumn 2010 edge and combined seasons.

The four creeks that did produce an OE50 average score for the Autumn 2011 sampling period showed only a small variation in average stream health compared to Autumn 2010. Shrimptons and Porters Creeks remained in the severely impaired band. Buffalo Creek had a slight increase in stream health, moving from the severely impaired band to the significantly impaired band. Terrys Creek had a slight decrease in stream health, moving from the significantly impaired band to the severely impaired band (Figure 9).

The Combined Season edge model output for 2010/11 showed no significant variation in average stream health. Shrimptons and Buffalo Creeks remained in the severely impaired band. Porters Creek remained in the significantly impaired band. The only change was Terrys Creek moving from the significantly impaired band to the severely impaired band. Despite the change in bands there was not a significant change in stream health from the previous 2009/10 sampling period (Figure 10).

The Combined Season edge AUSRIVAS OE50 model output summary shows Archers and Porters Creeks fall within the significantly impaired range. However, data points can only be generated for Archers Creek up to 2008 due to the model output describing it as OEM. Shrimptons, Buffalo and Terrys Creeks remain severely impaired (Figure 13).

The Autumn and Spring edge AUSRIVAS OE50 model output indicates similar trends in stream health across all five creeks. Archers and Terrys Creeks have the marginally higher average stream health of the five creeks and Shrimptons Creek is marginally poorer. The Autumn model output is generally indicative of higher stream health when compared to the respective Spring seasons for all creeks (Figure 8, Figure 9, Figure 11 & Figure 12).

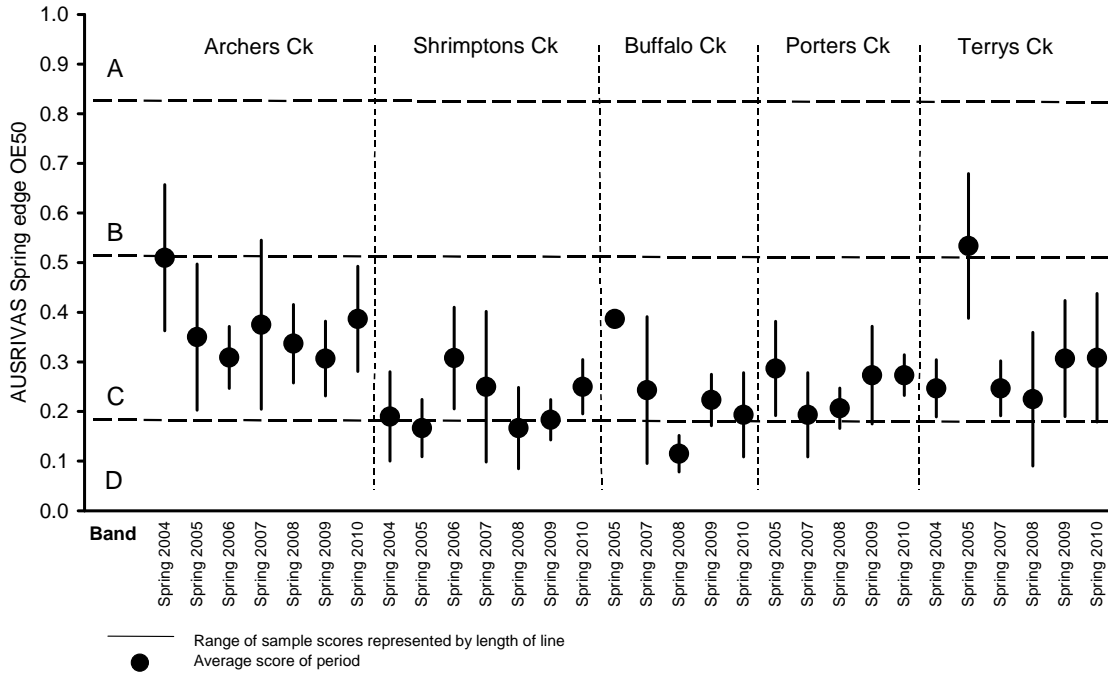


Figure 8 AUSRIVAS OE50 of all creeks from Spring edge model

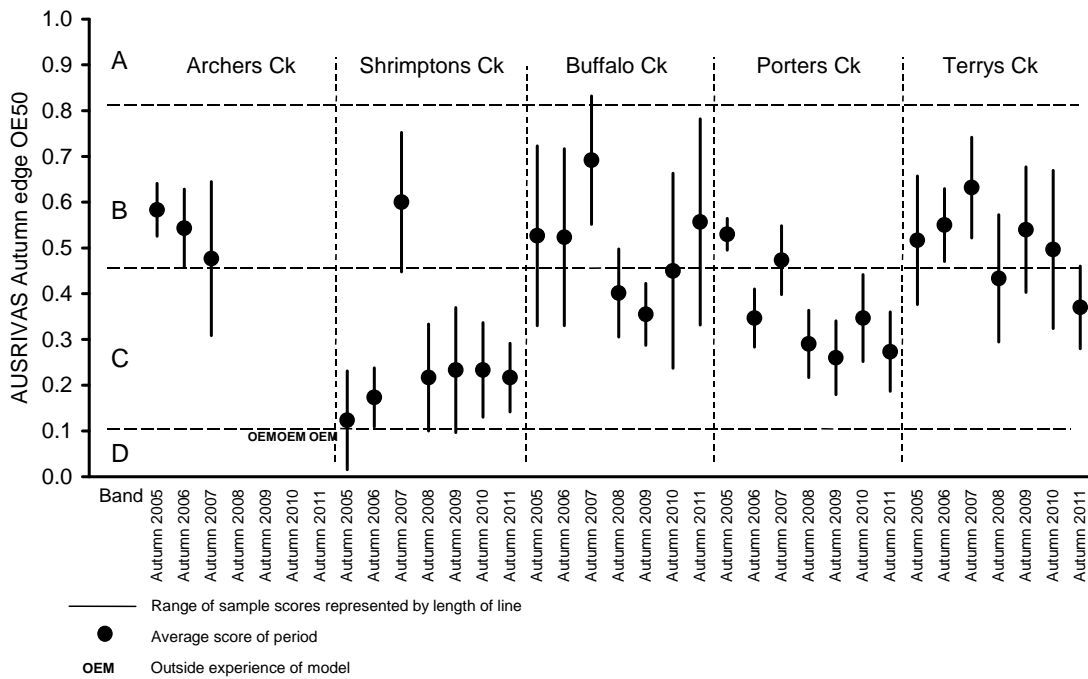


Figure 9 AUSRIVAS OE50 of all creeks from Autumn edge model

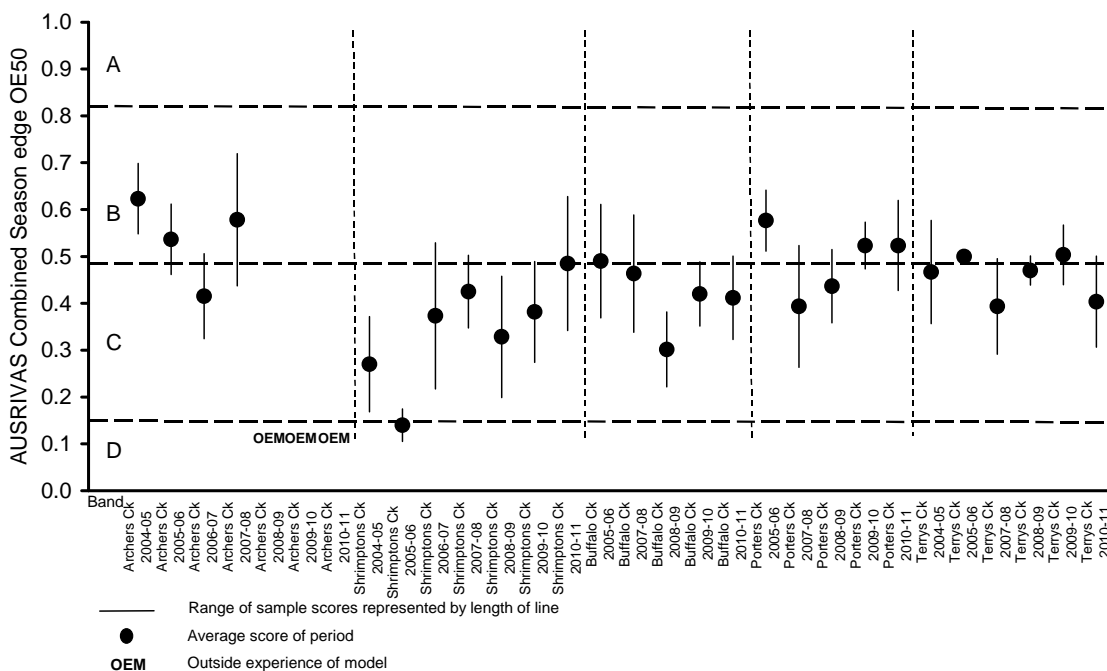


Figure 10 AUSRIVAS OE50 of all creeks from combined season edge model (with financial year data combined)

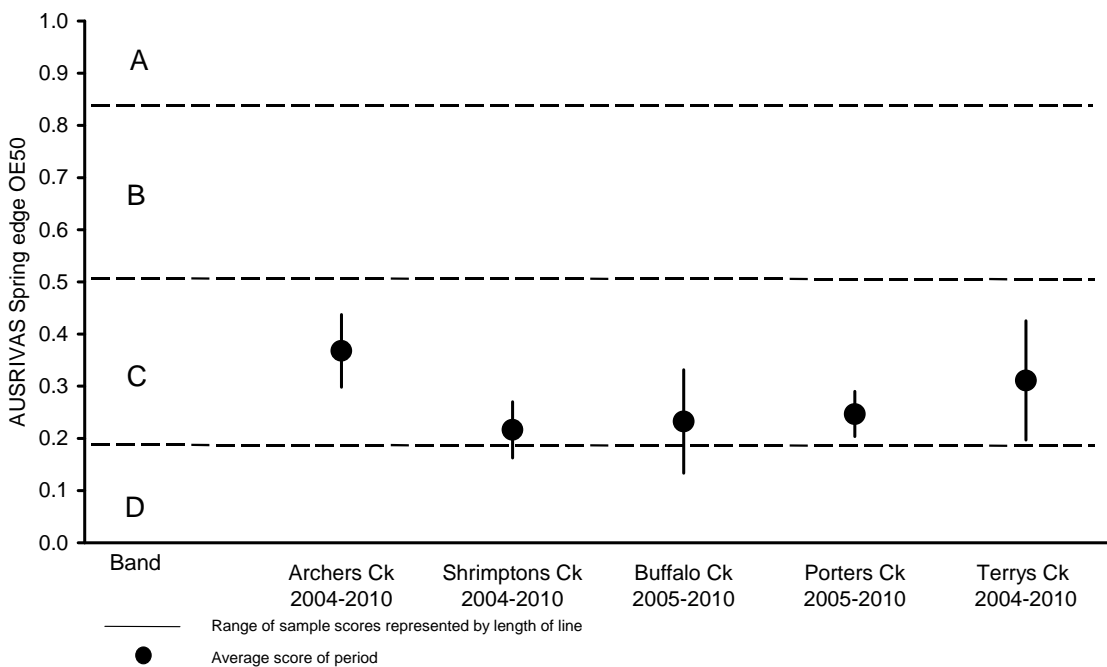


Figure 11 AUSRIVAS OE50 summary of all creeks from Spring edge model

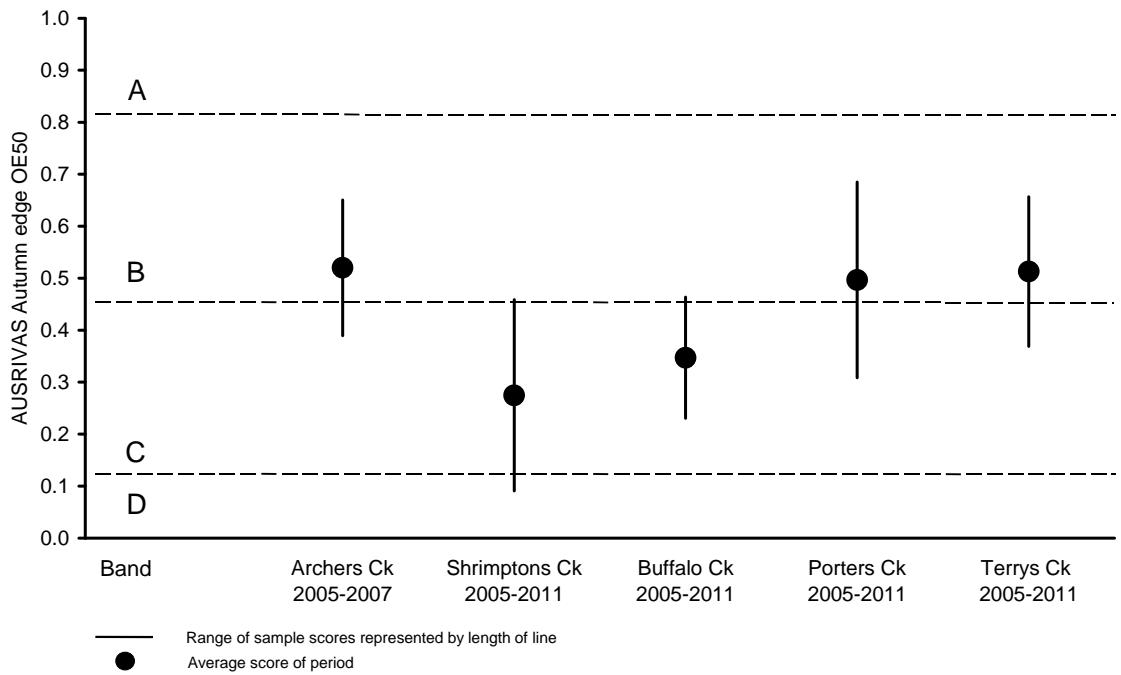


Figure 12 AUSRIVAS OE50 summary of all creeks from Autumn edge model

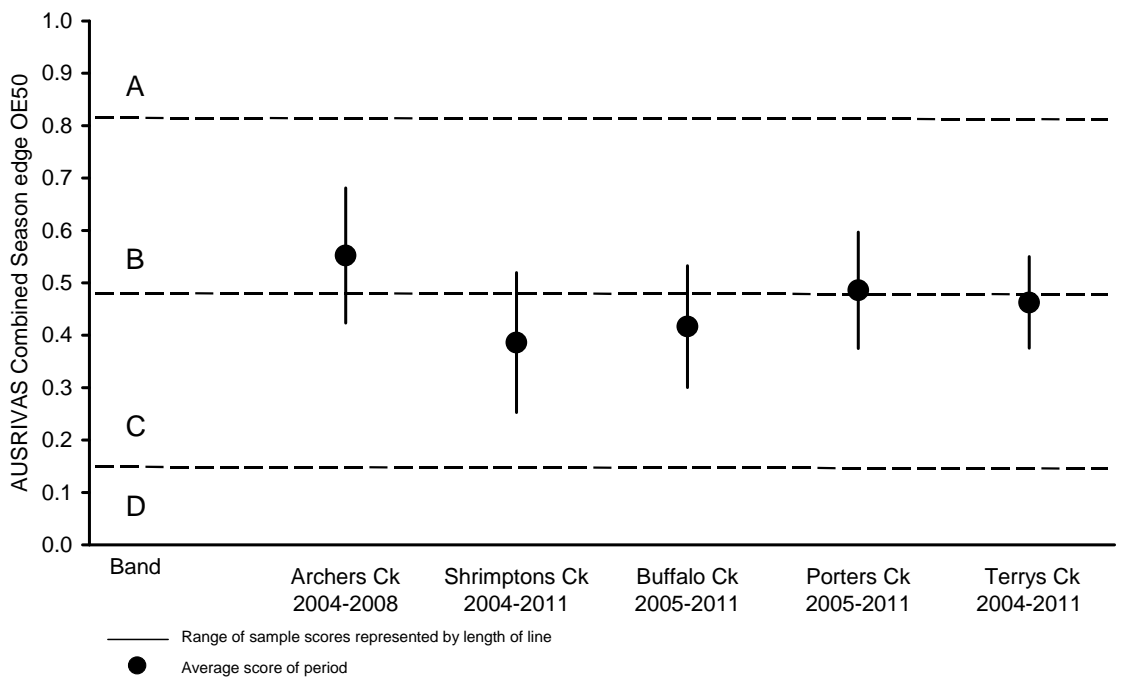


Figure 13 AUSRIVAS OE50 summary of all creeks from combined season edge model (with financial year data combined)

EPT Indicator taxa from AUSRIVAS predictive model output

AUSRIVAS output identifies taxa that were expected to be representative of a sample when compared to the respective reference site group by the AUSRIVAS model. As part of this output, missing taxa are listed with greater than 50% probability of occurrence. Indicator taxa are defined as taxa within the EPT orders (Ephemeroptera - mayfly, Plecoptera - stonefly and Trichoptera – caddisfly) with SIGNAL2 scores of greater than 6 (as per previous reports).

Across the five creeks of the monitoring program, missing EPT indicator taxa identified by AUSRIVAS Autumn edge and Combined season edge model output listed 16 taxa as missing. The taxa identified included three mayfly larvae (Ephemeroptera), two stonefly larvae (Plecoptera) and 11 caddisfly larvae (Trichoptera). The same indicator taxa were identified by both of the aforementioned model outputs.

There were three families of EPT taxa found during the Autumn 2011 sampling period, none of which were considered EPT indicator taxa.

AUSRIVAS OE0 SIGNAL2

The addition of Autumn 2011 data to the Autumn edge AUSRIVAS OE0 SIGNAL2 model allowed for the Combined Season edge output to be updated, as this is presented on a financial year basis.

The 2011 Autumn edge and 2010/11 combined season model does not include an average score for OE0 SIGNAL2 for Archers Creek. The model output described it as *outside the experience of the model* (OEM). The same output occurred for both Autumn 2009 and Autumn 2010 edge and combined season models.

The average scores for the AUSRIVAS OE0 SIGNAL2 Autumn edge for the 2011 sampling period were similar for all creeks, with no significant differences when compared to Autumn 2010 results. Shrimptons Creek and Porters Creek remained similar to the previous sampling period and Buffalo Creek displayed a slight increase in average stream health. While Terrys Creek's score fell below the previous two autumn seasons (Figure 15).

The AUSRIVAS Combined Season edge OE0 SIGNAL2 model output indicated the average scores from the recent 2010/11 sampling period increased slightly for Shrimptons, Buffalo and Porters Creeks in comparison to the 2009/10 sampling period. Terrys Creek displayed a small decline in average stream health when comparing the recent sampling period to the preceding sampling period (Figure 16).

The summary of the Autumn model output through time places the average score for the five creeks within a similar range of stream health. Shrimptons Creek has displayed the lowest average stream health for the Autumn edge sampling periods (Figure 18). The Combined Season model displays a similar trend to the Autumn edge model summary for all sampling periods, with Shrimptons Creek again displaying the lowest average scores (Figure 19).

The AUSRIVAS Spring edge OE0 SIGNAL2 output reflects a similar range of stream health when compared to the aforementioned models for all creeks of the sampling program (Figure 14 & Figure 17).

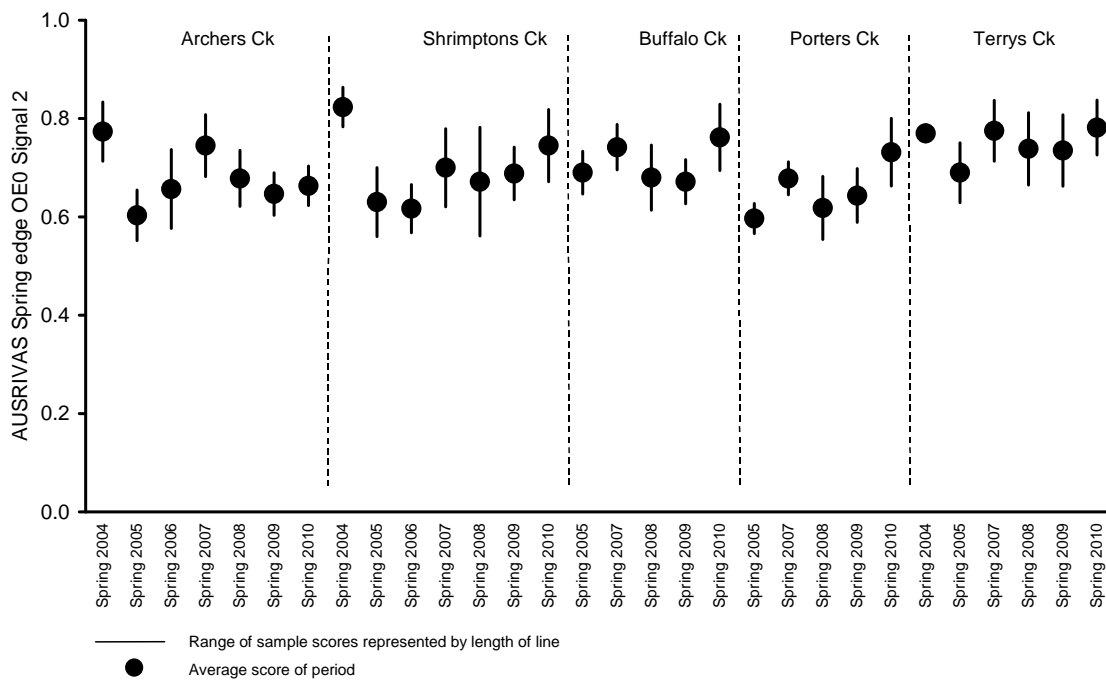


Figure 14 AUSRIVAS OE0 SIGNAL2 of all creeks from Spring edge model

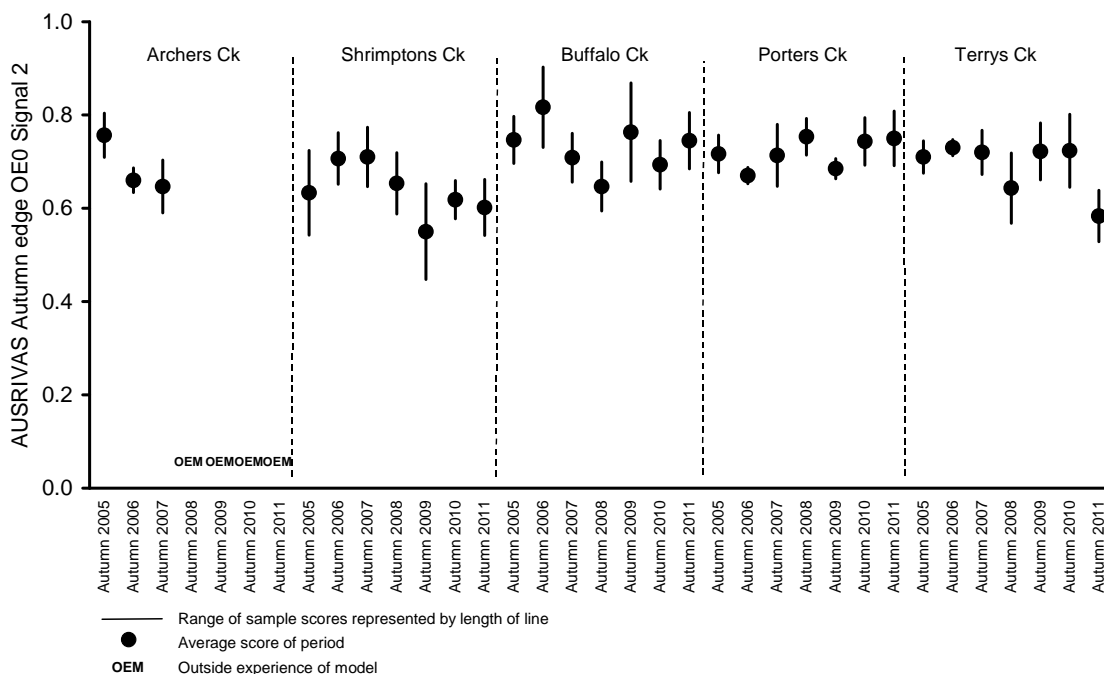


Figure 15 AUSRIVAS OE0 SIGNAL2 of all creeks from Autumn edge model

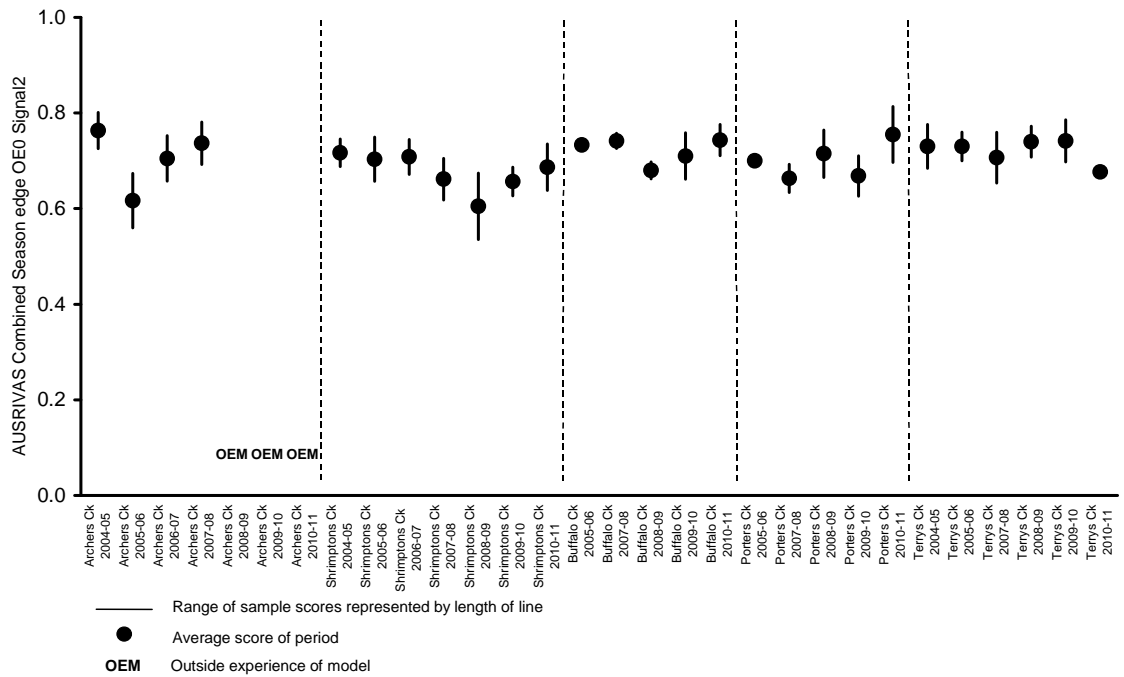


Figure 16 AUSRVAS OE0 SIGNAL2 of all creeks from combined season edge model (financial year data combined)

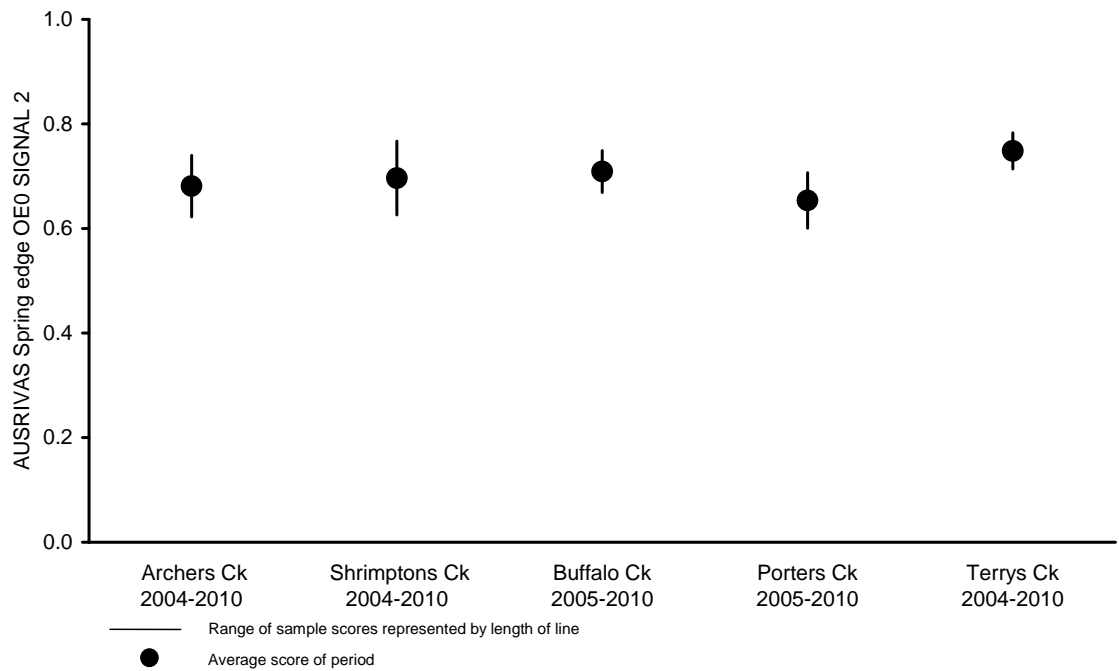


Figure 17 AUSRVAS OE0 SIGNAL2 summary of all creeks from Spring edge model

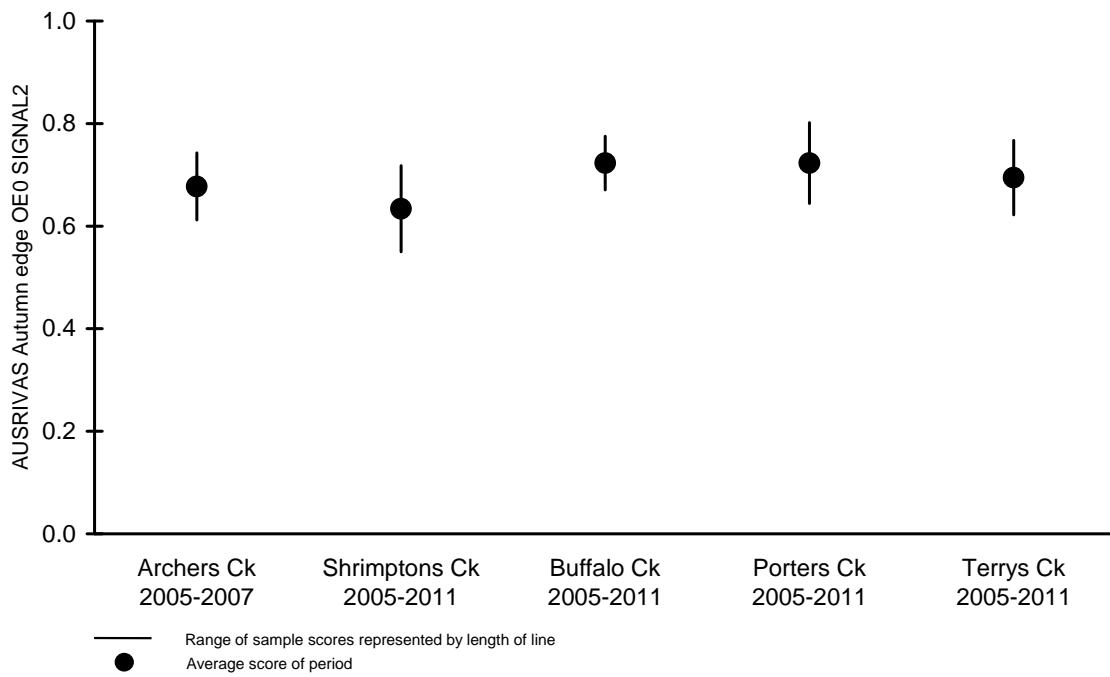


Figure 18 AUSRIVAS OE0 SIGNAL2 summary of all creeks from Autumn edge model

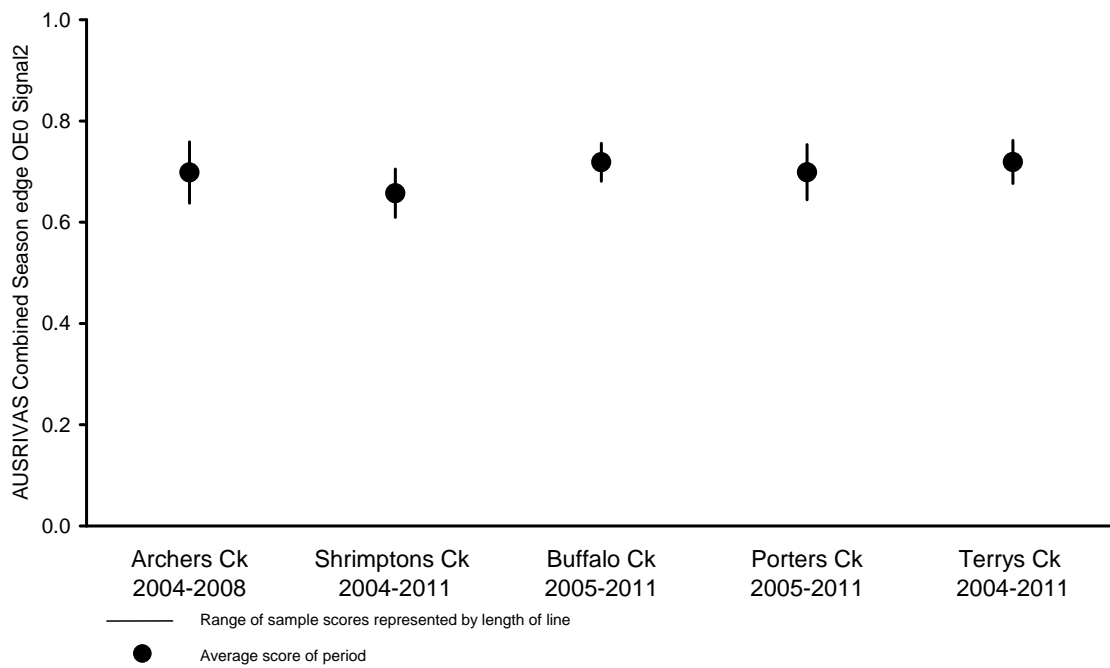


Figure 19 AUSRIVAS OE0 SIGNAL2 summary of all creeks from combined season edge model (with financial year data combined)

Multivariate Analyses

Ordination and SIMPROF test

In order to achieve suitable multivariate representations of data in two or three dimensions, replicates from the same season for each creek were merged. This produces one data point per creek per season, which minimizes the stress and gives a better measure of fit. This strategy has been used for the combined analysis of all five creeks due to the large number of replicates involved. Presenting the data in this way can be seen as '*reducing the noise*' of the replicate data from the somewhat patchy occurrence of macroinvertebrates at a stream site.

The three-dimensional MDS ordination plot for all five creeks and individual creeks are presented in this report, as the stress value is lower in three dimensions than in two. This lower stress value means the differences in community structure between creeks and seasons is better represented in the three-dimensional plot, despite the three dimensions being represented in two dimensions. A two-dimensional presentation of the ordinations is the preferred reporting format, however ordinations with high stress values (>0.2) are considered inappropriate representations of community structure.

The three-dimension ordination plot highlights variability in the community structure of Shrimptons Creek, both between its seasons and when compared to the other creeks through time. Terrys Creek has the least variation in community structure of the five creeks of the program. Archers, Buffalo and Porters creeks indicate a similar variability through time (Figure 20).

The SIMPROF test provides another way to view community structure differences and similarities between samples. SIMPROF results are overlaid onto the classification result output (dendrogram), with black lines indicating a real difference in community structure and red lines no real difference.

All creeks are largely separated into their own test groups as highlighted by the SIMPROF test. Shrimpton Creek samples are in the first and third test groups to be separated, with Autumn/Spring 2005 and Autumn 2006 (43% similarity) being separated from all other Shrimpton Creek samples. Archers Creek (Autumn 2011 & Autumn 2008) and Terrys Creek (Autumn 2011) are separated into their own test groups. All of the 'real' test groups separated by the SIMPROF test apart from the first Shrimptons Creek group split at a relatively high similarity (56% $>$) (Figure 21).

All five creeks replicates merged

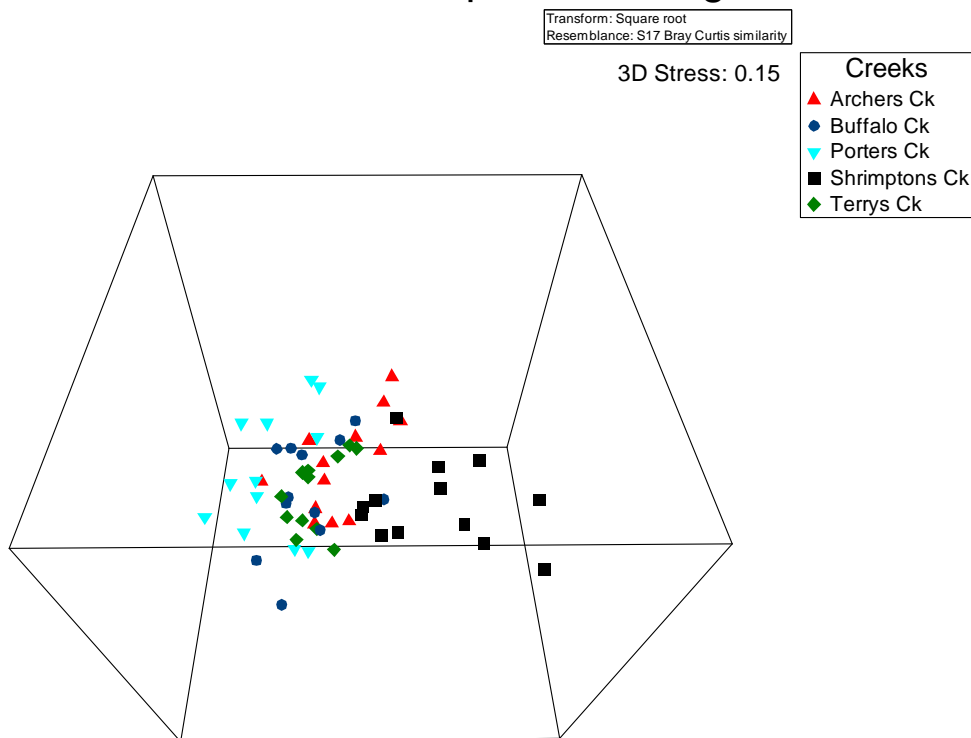


Figure 20 Plot of non-metric multidimensional scaling ordination results of 3-dimension analysis for 2005 to 2011 macroinvertebrate data of all creeks, with each point of the same creek representing a different season

All five creeks replicates merged

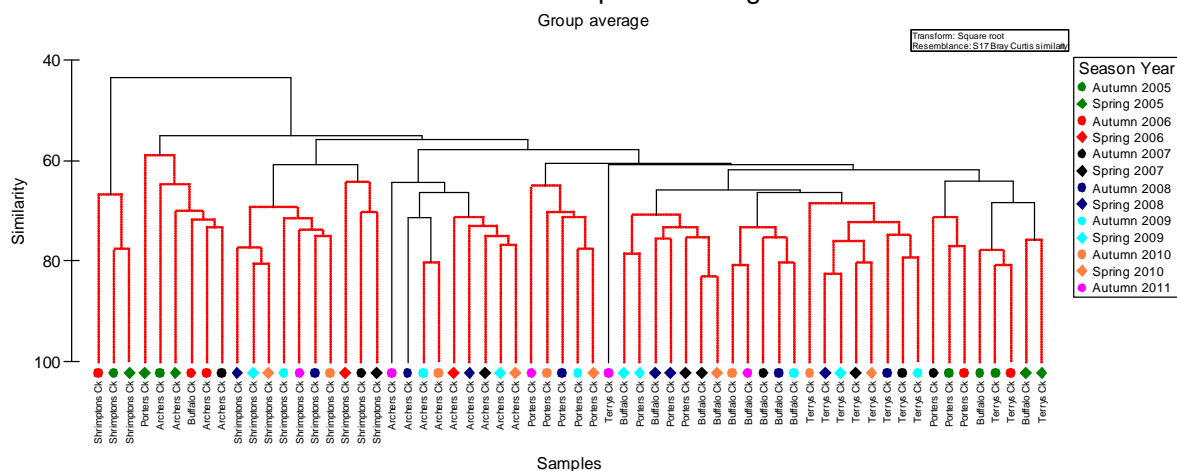


Figure 21 Dendrogram of all five creeks with SIMPROF test sample groups

The Archers Creek ordination plot and SIMPROF test results indicate a general separation of autumn and spring results, with two single samples separated from Autumn 2007 and Spring 2010 (Figure 22 and Figure 23).

Archers Creek

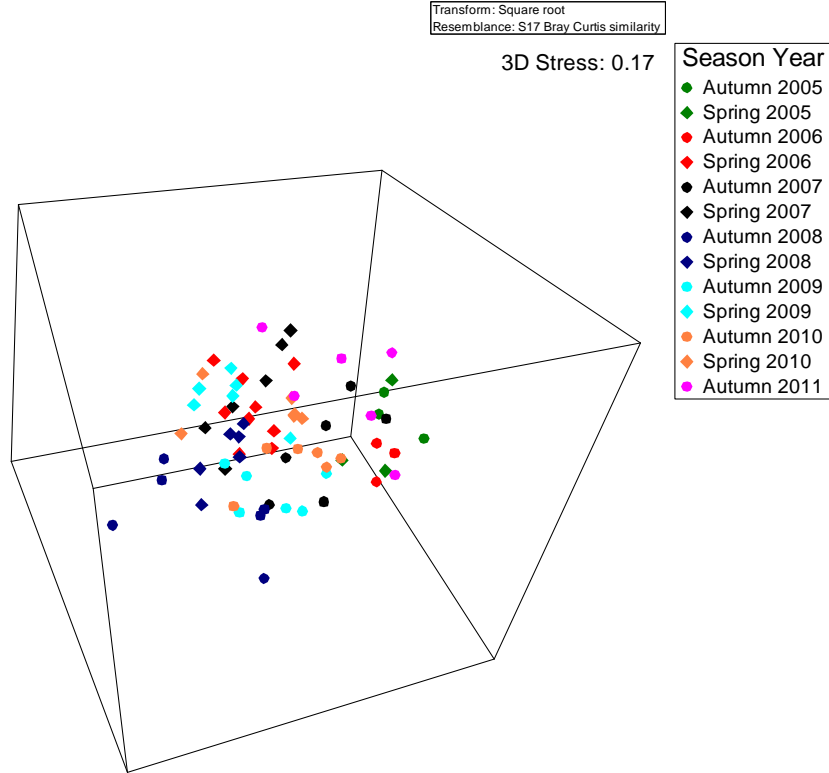


Figure 22 Plot of non-metric multidimensional scaling ordination results of 3-dimension analysis for 2005 to 2011 macroinvertebrate data of Archers Creek

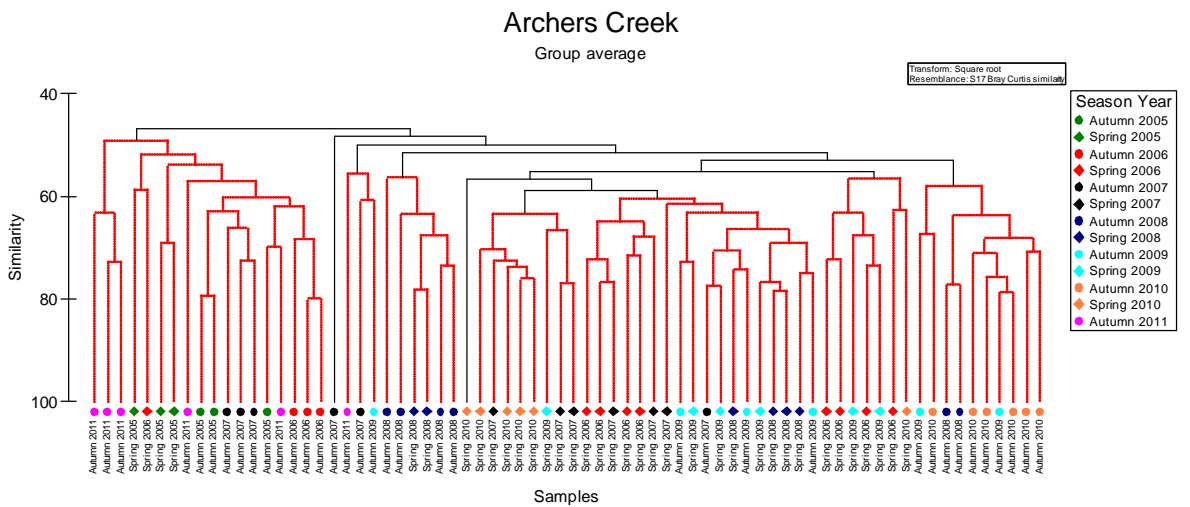


Figure 23 Dendrogram of Archers Creek with SIMPROF test sample groups

The ordination plot of Shrimptons Creek indicates some seasonal separation. The SIMPROF test results show no notable test group separation, despite test group splits at 40 and 45% similarity (Figure 24 and Figure 25).

Shrimptons Creek

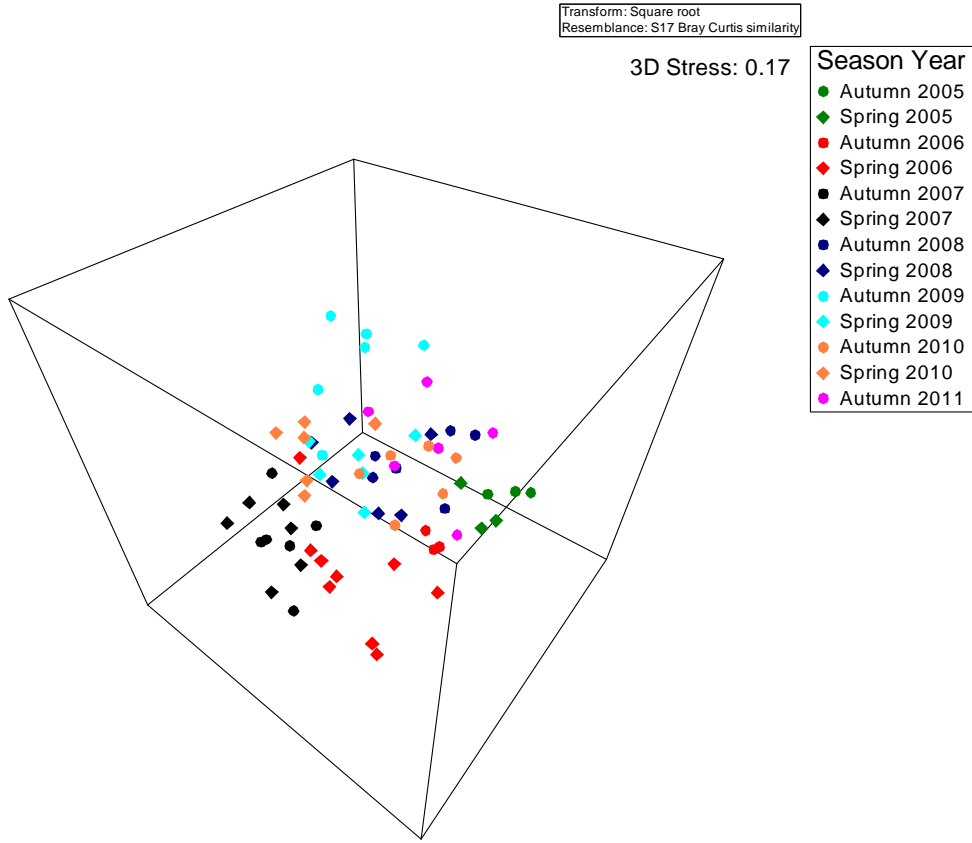


Figure 24 Plot of non-metric multidimensional scaling ordination results of 3-dimension analysis for 2005 to 2011 macroinvertebrate data of Shrimptons Creek

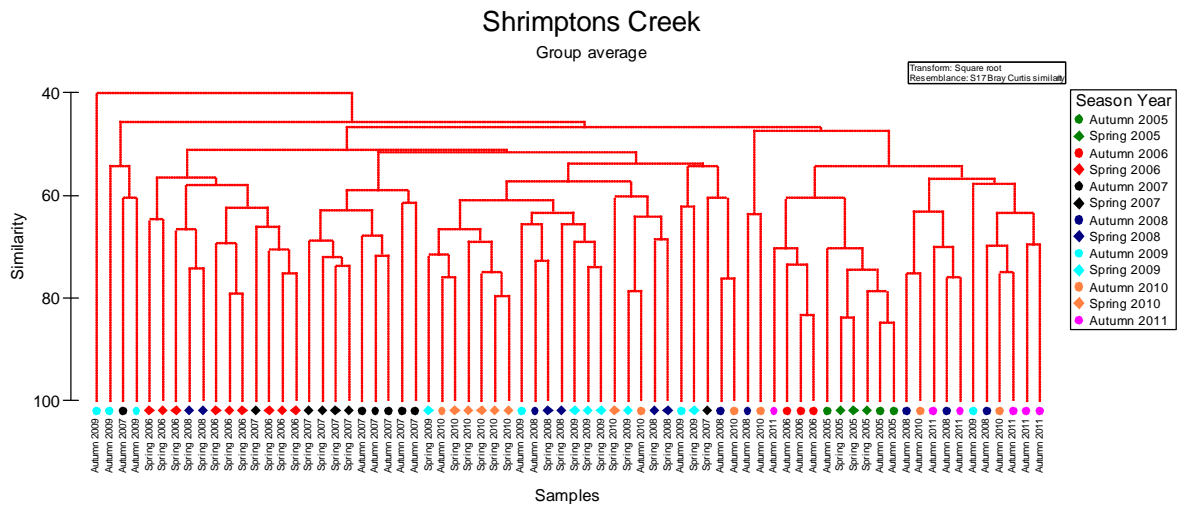


Figure 25 Dendrogram of Shrimptons Creek with SIMPROF test sample groups

The Buffalo Creek ordination plot and SIMPROF test separate Autumn 2005, Spring 2005 and Autumn 2006 from all other samples, the first three seasons sampled in the sampling program. For all other samples there is a general separation of spring and autumn samples in the ordination plot. This separation is particularly evident from the SIMPROF test, where all samples are separated into groups based on season except for Spring 2007 and the first three sampled seasons (Figure 26 and Figure 27).

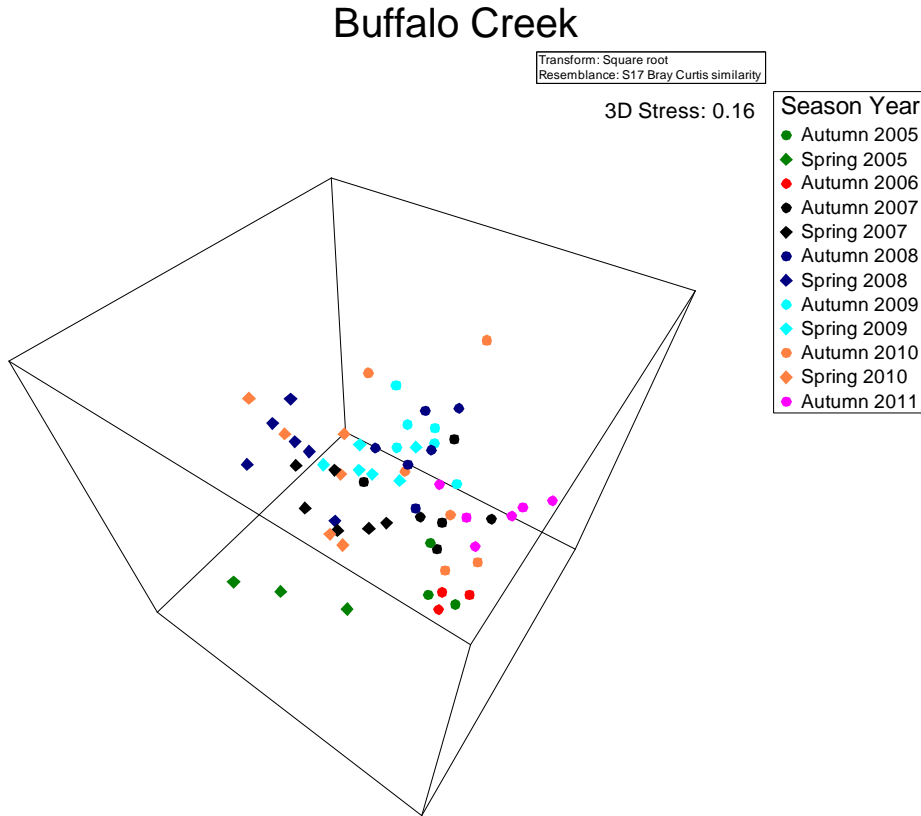


Figure 26 Plot of non-metric multidimensional scaling ordination results of 3 dimension analysis for 2005 to 2011 macroinvertebrate data of Buffalo Creek

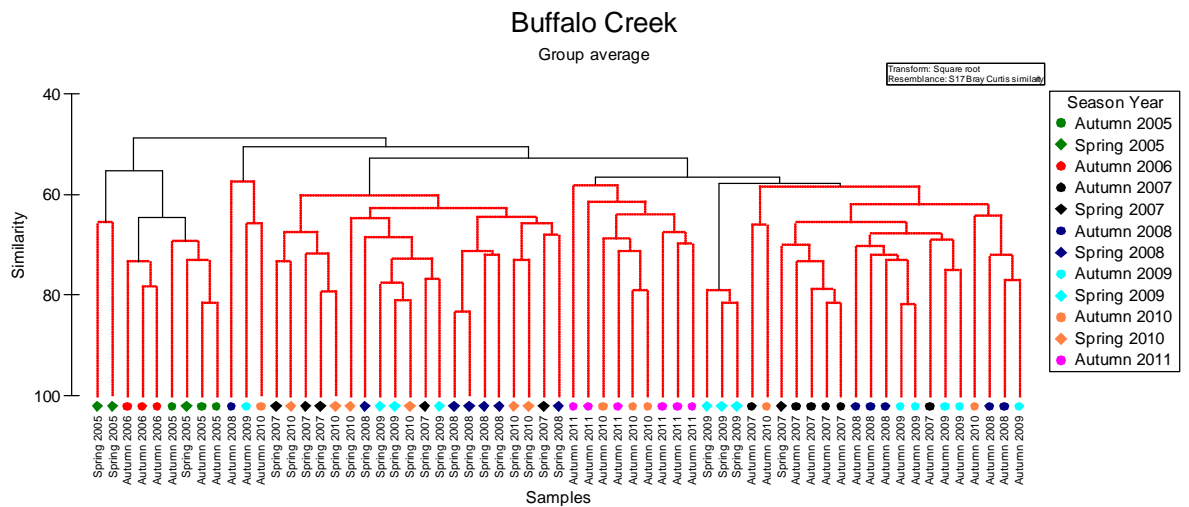


Figure 27 Dendrogram of Buffalo Creek with SIMPROF test sample groups

The Porters Creek ordination plot and SIMPROF test results show a single outlying sample from Autumn 2011 and two groups of samples from Spring 2008. Both multivariate analyses indicate a separation of samples from Autumn 2005 to Autumn 2007 and samples from Spring 2007 to Spring 2010, except one Autumn 2010 sample and Autumn 2011 samples (Figure 28 and Figure 29).

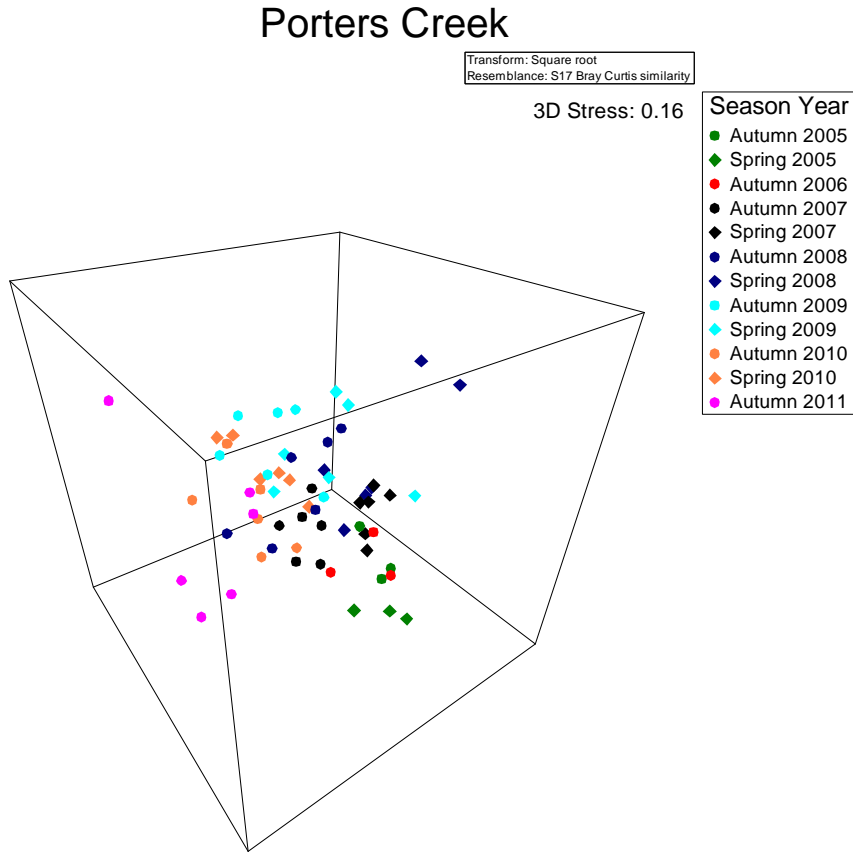


Figure 28 Plot of non-metric multidimensional scaling ordination results of 3 dimension analysis for 2005 to 2011 macroinvertebrate data of Porters Creek

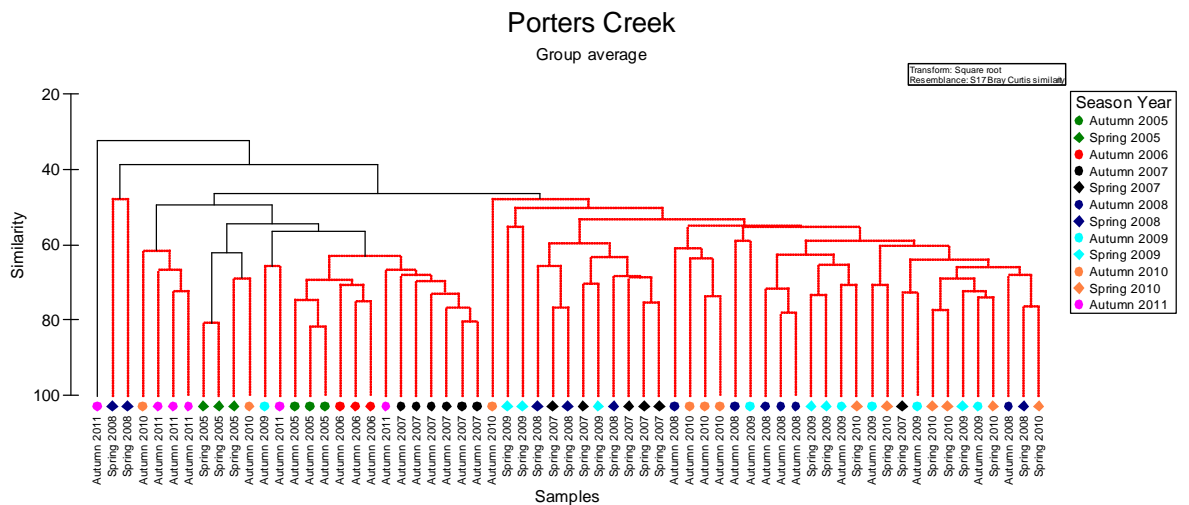


Figure 29 Dendrogram of Porters Creek with SIMPROF test sample groups

The first Terrys Creek test group to be separated by SIMPROF is five of the six Autumn 2010 samples, this separation is evident in the ordination plot. In the remaining samples there is a general separation of spring and autumn samples (Figure 30 and Figure 31).

Terrys Creek

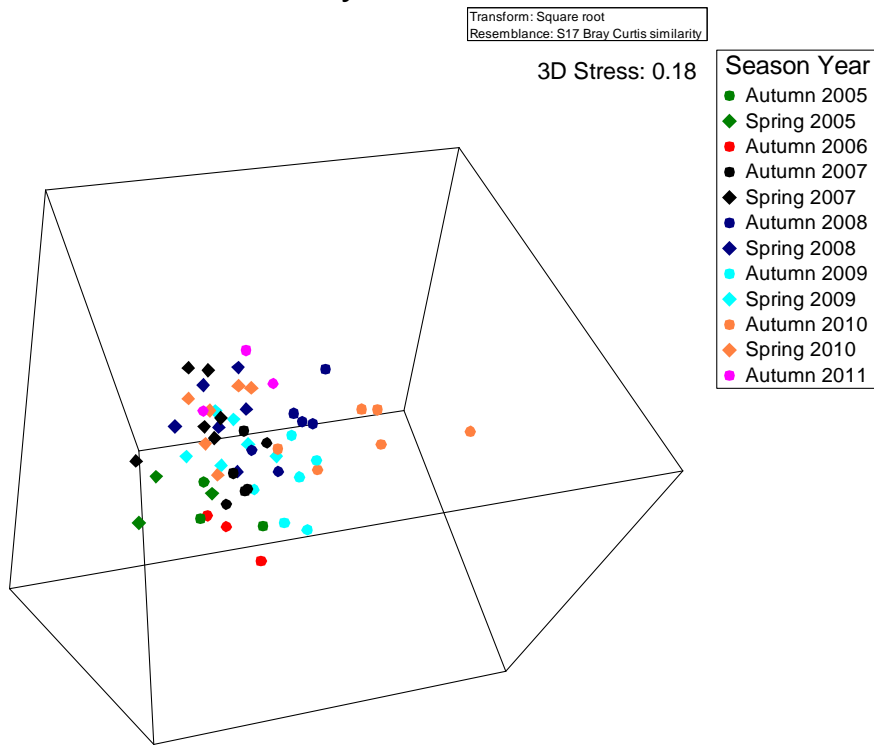


Figure 30 Plot of non-metric multidimensional scaling ordination results of 3-dimension analysis for 2005 to 2011 macroinvertebrate data of Terrys Creek

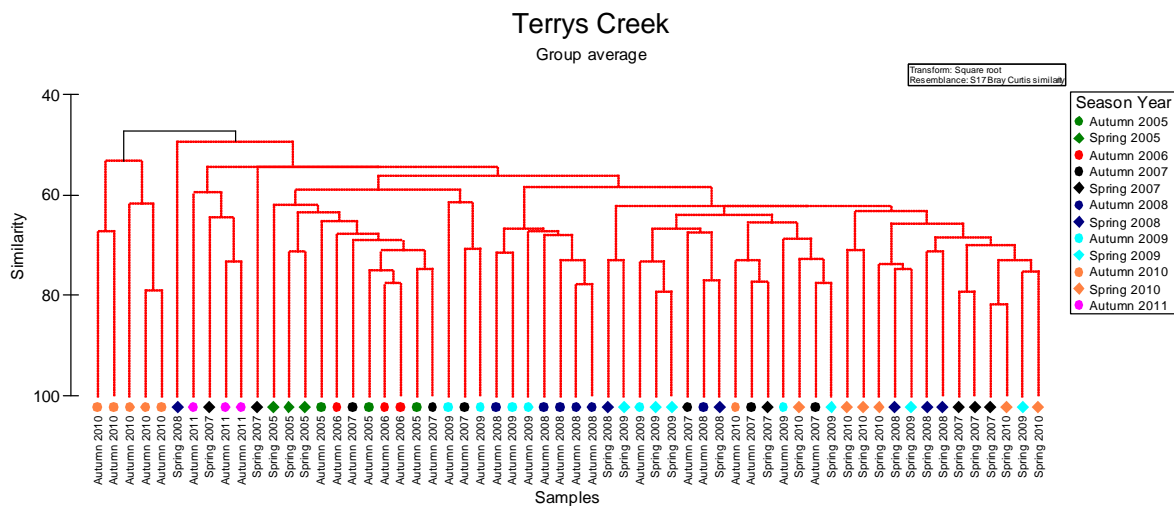


Figure 31 Dendrogram of Terrys Creek with SIMPROF test sample groups

SIMPER

SIMPER results from 2005 to 2011 for each creek indicated that Shrimptons Creek had the lowest overall similarity of 61%. Archers, Porters and Buffalo creeks had slightly higher similarities with 63%, 63% and 66% respectively. Terrys Creek had the highest similarity with 69% (Appendix 5). These similarities reflect the amount of variation (the lower the percentage, the more variation) in macroinvertebrate community structure over time within the individual creeks in the program.

SIMPER compares samples from each creek with those of all other creeks. These results are referred to as average dissimilarity. These values are presented in Table 14 and indicate that samples from Buffalo, Porters and Terrys creeks are most similar. This reflects the closer yet still separate position of those data points in the ordination plot of all five creeks. Shrimptons Creek samples are the most dissimilar to all the other creeks' samples, and this is reflected in the ordination plot of all five creeks (Figure 20).

Table 14 Average dissimilarity between samples of each creek comparisons

	Archers %	Shrimptons %	Buffalo %	Porters %
Shrimptons %	47			
Buffalo %	43	47		
Porters %	45	51	39	
Terrys %	44	46	37	39

SIMPER also looks at the similarity of samples within each of the five creeks of the program, complementing the MDS plots and dendrograms (SIMPROF) in the previous section. The range of average similarity of samples within the five creeks was 48% to 77% (Table 15). The SIMPER output includes individual macroinvertebrate abundances, which are the drivers of the sample similarities.

The largest range in sample similarity (48-77%) is found in Shrimptons Creek, reflective of a significant change in macroinvertebrate community structure. From Autumn 2005 to Autumn 2006, tolerant non-insects dominated community structure, with five to six taxa contributing to 90% of the overall samples. From Spring 2006 to Spring 2007 there was a change, with up to 10 dominant taxa and tolerant insects significantly contributing to the community structure. Since Autumn 2008 to the current Autumn 2011 season, the community structure has returned to being dominated by tolerant non-insects with fewer dominant taxa (Appendix 5).

In the five creeks of the Ryde LGA, common non-insect community members included the introduced snail *Physa acuta* (Physidae), flatworms (DugesIIDae) and worms (Oligochaeta). The tolerant insects found were native non-biting midges (Chironominae), dragonflies (Megapodagrionidae, Coenagrionidae, Isostictidae, Hemicorduliidae) and back-swimmers (Notonectidae). For all five creeks of the monitoring program the single most dominant taxa is the dipteran sub-family, Chironominae.

The SIMPER output for Archers, Buffalo, Porters and Terrys creeks indicated that seasonal variations were the main driver influencing the minor changes in macroinvertebrate community structure.

During the monitoring program Archers Creek spring samples have had a 40% and less contribution from tolerant insects. In contrast the contribution in autumn has been higher at 50% and more. The number of dominant taxa per season has remained relatively stable through time and seasons in Archers Creek.

The community structure in Buffalo Creek has shown a similar seasonal trend to Archers Creek. In Spring 2008 the low taxa dominance of non-insects was particularly evident with an 80% contribution from just three non-insect taxa. This change however, was linked to a catchment disturbance rather than a seasonal shift.

The community structure in spring for Porters and Terrys creeks is dominated by few taxa and higher contributions of non-insects occur. In autumn there is a more diverse range of dominant taxa with a higher contribution by insect taxa.

Table 15 Average similarity of the same season samples for each creek

	Archers %	Shrimptons %	Buffalo %	Porters %	Terrys %
Autumn 2005	68	76	76	77	70
Spring 2005	59	77	66	73	65
Autumn 2006	72	77	75	72	73
Spring 2006	60	62	ns	ns	ns
Autumn 2007	57	60	70	71	66
Spring 2007	61	63	65	68	65
Autumn 2008	61	58	64	60	67
Spring 2008	70	63	66	52	62
Autumn 2009	64	48	69	58	62
Spring 2009	65	62	69	56	67
Autumn 2010	66	58	56	55	59
Spring 2010	64	65	61	68	64
Autumn 2011	52	60	62	51	66

BIOENV

The output of BIOENV routine for all five creeks and for each individual creek is presented in Appendix 6. The strongest correlation of the data for all five creeks for 2005 to 2011 was mild at 0.396. The variables identified in the best correlation consisted of total oxidised nitrogen, conductivity, dissolved oxygen, ratio of total length of pipe/catchment area, and ratio of number of outlets/catchment area. The ratio of number of outlets/catchment area was the only variable that was found in all of the ten best correlations in the BIOENV output. Oxidised nitrogen was found in nine of the ten best correlations.

BIOENV analysis of each individual creek for 2005 to 2011 produced weak to mild correlations Archers (0.237), Shrimptons (0.280), Buffalo (0.243), Porters (0.364) and Terrys (0.260) (Appendix 6).

The strongest correlation for Porters Creek was a mild correlation and included turbidity, conductivity, and total dissolved solids. These variables were in most of the strongest correlations for Porters Creek. The remaining creeks returned only weak correlations and included a variety of variables in their strongest correlation.

Table 16 Catchment storm water delivery characteristics for each creek

Creek	Total Length of Pipe (TLP) (m)	Total Number of pipe Outlets (NO)	Catchment Area (CA)(hectares)	Ratio TLP/CA	Ratio NO/CA
Archers	19,310	65	286	67.5	22.7
Shrimptons	41,797	74	555	75.3	13.3
Buffalo	33,336	62	546	61.1	11.3
Porters	15,797	16	225	70.2	7.1
Terrys	47,952	89	1012	47.4	8.8

5 Discussion

5.1 Water Quality

Water quality results, while not reflecting a sampling frequency suggested by ANZECC (2000), did allow for characterisation of water quality at each study creek against ANZECC (2000) guidelines for Aquatic Ecosystems (Lowland River SE Australia) and Recreational Water Quality and Aesthetics (Secondary).

Results of the Autumn 2011 water quality sampling for Shrimptons, Porters, Buffalo, Terrys and Archers creeks support previous sampling results indicating that urban pollution transport is having an impact on instream water quality. This impact was indicated by low levels of dissolved oxygen and high concentrations of nutrients, especially nitrogen forms.

This trend was observed in 2004 through to 2010 (Ecowise 2004, 2005a 2005b 2006; Sydney Water 2006, 2007a, 2007b, 2008a, 2008b, 2009a, 2009b, 2010a, 2010b). Pollutant concentrations have been spatially variable, indicating that they originate from varying locations over a constantly changing time period.

The rainfall in late 2010 was characterised by consistent light to medium rainfall periods. Very little rain fell between January and mid March. Consistent light to medium rainfall periods from mid March was punctuated by several events of daily rainfall greater than 30 mm.

Porters Creek had a very high faecal coliform density at the Wicks Road site in April 2011 (>10,000 CFU/100mL). Total nitrogen (1,380 µg/L), total kjeldahl nitrogen (610 µg/L) oxidised nitrogen (770 µg/L) and total phosphorus (220 µg/L) concentrations were also elevated. Ammonia concentrations were low at <10 µg/L in both March and April 2011.

The faecal coliform density at Wicks Rd in April 2011 of >10,000 CFU/100mL was later checked and a recording of 32,000 CFU/100mL was found. The sample was greater than 24 hours old by the time of the second result; as such it cannot not be a confirmed result and cannot be used for the report.

The faecal coliform densities at the other Porters Creek sites were all below the 1,000 CFU/100mL guideline in Autumn 2011. The ammonia concentrations of 210 µg/L and 330 µg/L recorded from the core site on Porters Creek downstream of the depot were well below the historical average for this site (795 µg/L) and are the lowest recorded at this site since Autumn 2008.

The indicator species used for faecal coliforms are naturally occurring, harmless inhabitants of the digestive tract of all warm-blooded animals (Boey, 1993). The occurrence of these bacteria in large numbers signifies the presence of faecal pollution and, therefore, the possible presence of pathogenic organisms that occur in faeces. A variety of factors including urban runoff, presence of waterfowl and other wildlife, waste depots, illegal dumping of waste and sewer overflows can influence faecal contamination of urban streams.

The historical averages for faecal coliforms (calculated from the results of the core site in each creek) were above the recommended guideline of 1,000 CFU/100mL (ANZECC, 2000) for three of the five creeks:

- Terrys Creek (2,432 CFU/100mL with 2 individual exceedences)

- Porters Creek (5,612 CFU/100mL with 7 individual exceedences) and
- Shrimptons Creek (1,788 CFU/100mL with 4 individual exceedences).

This indicates that either or both of the individual exceedence results for Terrys Creek were particularly high and contributed to the elevated historical average. A result of 60,000 CFU/100mL was recorded from Terrys Creek in Autumn 2005 - indicating that the majority of faecal coliform results for this site were low with rare instances of very high results.

Archers Creek, while having a faecal coliform historical average falling below the guideline at 790 CFU/100mL, has had a higher number of exceedences; 7 individual results greater than 1,000 CFU/100mL. This indicates a more frequent lower level impact for faecal coliforms at this site.

An unusually high ammonia concentration was recorded from Terrys Creek in March (1,030 $\mu\text{g/L}$. Total nitrogen (1,680 $\mu\text{g/L}$, total Kjeldahl nitrogen (1,50 $\mu\text{g/L}$) and total phosphorus (243 $\mu\text{g/L}$) concentrations were also high. These are the highest recorded for this site. Elevated concentrations for these analytes were also recorded in September 2010.

Ammoniacal nitrogen is often present in sewage effluent because of the decomposition of nitrogen containing compounds in the treated waste. The undissociated form, ammonia (NH_3) is far more toxic to aquatic life than the ionic form, ammonium (NH_4^+). During low pH and temperatures NH_3 dissociates to the less toxic form NH_4^+ . This is then reversed during periods of high pH and temperature.

Laboratory methods for ammonia record the nitrogen content from the ammonium (associated ionic form) ions NH_4^+ . This ion forms compounds with other particles dissolved within the water column. It is harmless to plants and animals within the specified concentration, pH and temperature range. ANZECC (2000) has determined this to be 20 $\mu\text{g/L}$ for the protection of aquatic life in lowland streams with a pH of 8 and temperature of 20⁰C.

Ammonia (NH_3) is a toxic by-product of NH_4^+ that exists as a gas, of which the N content is not measured during the routine laboratory analysis. With increasing temperature and pH, the percentage of NH_3 increases exponentially and it is this compound that is detrimental to aquatic life.

Dissolved oxygen concentrations are an important water quality indicator for the survival of aquatic organisms and control many important physico-chemical processes. The oxygen balance in water is dependant upon physical, chemical and biochemical conditions in the water body. Oxygen input is the result of diffusion from the atmosphere and photosynthesis by algae and other aquatic plants. Dissolved oxygen removal is due to respiration by aquatic organisms, decomposition of organic matter, oxidation of chemically reduced compounds and loss to the atmosphere. The solubility of oxygen in water decreases with increasing temperature, while the respiratory rate of aquatic organisms increases with temperature (Connell, 1993).

Dissolved oxygen concentrations are often subject to large diurnal and seasonal fluctuations as a result of changes in temperature and photosynthetic rates. Therefore, a dissolved oxygen measurement taken at one time of the day may

not truly represent the oxygen regime in the water body. Nevertheless, the low dissolved oxygen levels during Autumn 2011 for Terrys, Shrimptons, Buffalo and Archers creeks are an area of concern. These sites are continually showing impacted dissolved oxygen saturation levels, particularly during periods of extended low flow. Shrimptons Creek has the lowest historical average saturation level at 41.5 % saturation.

Dissolved oxygen saturation levels in Porters Creek at both the core site and additional sites were at acceptable levels, with the exception of the Spur Branch site in April. Porters Creek has the healthiest historical average for dissolved oxygen saturation (88.8 % saturation) of the five creeks, having the only historical average falling within the guideline range. This is partly related to more efficient run-off transport during both wet and dry periods.

Turbidity can be caused by soil erosion, waste discharge, urban runoff, algal growth and other disturbances in the water channel. Particles can smother aquatic insects, clog fish gills, prevent egg and larval development, reduce aquatic flora and fauna growth rates and generally decrease resistance to disease.

No sites on Terrys, Shrimptons, Buffalo, Porters or Archers creeks had elevated turbidity levels during Autumn 2011. These results are consistent with the historical averages for these sites of which none exceed the recommended guideline of 50 NTU.

5.2 Macroinvertebrates

The macroinvertebrate results indicated that Archers, Shrimptons, Buffalo, Porters and Terrys creeks had impaired macroinvertebrate communities, and the results from Autumn 2011 reflected this.

The City of Ryde Monitoring Strategy uses comparable data from all five creeks, each of which experiences natural variations in macroinvertebrate assemblages. The monitoring program resulted in a total of twelve seasons of comparable data for all five creeks since sampling began in Spring 2004. ANZECC (2000) recommends that to make informed management judgements adequate baseline data is required. Baseline data allows for the determination of an acceptable level of change in an environment accounting for the natural variability of an indicator (in this case, macroinvertebrates). It is suggested by ANZECC (2000) that three to five years of data be gathered from control or reference locations for comparison. The current baseline data will allow for tracking of any significant changes in macroinvertebrate assemblages during any future monitoring programs.

The Sydney specific SIGNAL-SF index and the NSW AUSRIVAS predictive models provide this data by the statistically defined 10th percentile of mean reference condition values. The range of each measure of stream health has been plotted in this report with a +/-1 standard deviation of the mean for basing ecological decisions. Presenting data in this way attempts to take account of variation at study sites and provide a basis for management tracking and ecological decision making (ANZECC, 2000).

A total of 2,044 macroinvertebrates were collected during the Autumn 2011 sampling season. The total number of macroinvertebrates collected since Spring 2004 has fluctuated between seasons, this variation can be influenced by several factors. Changes at sampling sites and replicate numbers sampled each season can have an influence. Also, numbers can be reflective of environmental cues that influence the development of macroinvertebrate taxa rather than water quality or other in-stream factors. This means that taxa may not be present in the water at the time of sampling or the cohort (age class) may be too small to be retained by the 0.25 mm mesh of the net.

The measure of sensitive taxa, EPT richness, indicated very low diversity in Autumn 2011. The highest average diversity of EPT taxa was at Porters Creek but was only one EPT taxa per sample. There were three families of EPT taxa sampled in Autumn 2011. Consisting of two Trichoptera; Hydroptilidae and Leptoceridae and the Ephemeroptera, Baetidae.

EPT taxa collected during the monitoring program from the five creeks have been in low abundances and found sporadically. Throughout the monitoring program only five families of EPT taxa have been sampled from the five creeks. The only consistent EPT taxa present is the Trichopteran Hydroptilidae, which has a SIGNAL2 score of 4. Although it is an EPT taxa it is considered a tolerant macroinvertebrate, as is the next most common EPT taxon the Ephemeroptera, Baetidae.

With these factors in mind EPT richness as a measure of stream health is limited in its ability to suggest or indicate any future positive impacts on stream health. The Spring 2007 report suggested that a return to average or above average

rainfall conditions might influence the presence of EPT taxa. Nominal average rainfalls have returned, yet the presence of EPT taxa has remained consistently low.

Considering the relatively low occurrence and type of EPT taxa found during the current monitoring program, reference to EPT indicator taxa from the AUSRIVAS model (as per criteria of Section 3.6) is recommended. This would be done before assessing positive changes in this measure and attributing them to management activities. EPT indicator taxa are considered sensitive and their presence/absence would be a more appropriate indicator of improved stream health.

Direct measurement of stream health using SIGNAL-SF and measurement via AUSRIVAS predictive model OE50 and OE0 SIGNAL2 outcomes reflected impaired stream health for all five creeks. The multivariate analysis tools complement univariate analyses in exploring patterns of macroinvertebrate communities by looking at the chosen array of samples and all taxa recorded. The univariate analyses indicated that all five creeks had relatively similar stream health. Likewise, the multivariate analyses indicated that all creeks had relatively similar macroinvertebrate community assemblage. The exception to this was Shrimptons Creek, which showed greater variation over time and when compared to the other creeks of the monitoring program.

Archers Creek

The SIGNAL-SF index and AUSRIVAS OE50 Spring and combined season models indicated that Archers Creek was slightly healthier than the other four creeks. The AUSRIVAS OE0 SIGNAL2 Spring and combined season models place Archers Creek within the range of the other four creeks.

The analysis of Archers Creek using AUSRIVAS is greatly restricted by the absence of autumn and combined season data points since Autumn 2007 (explained further in this section). There has not been an issue when running the Spring AUSRIVAS models.

The Archers Creek SIGNAL-SF average score increased in Autumn 2011 compared to the previous Spring 2010 samples when it recorded its lowest average score of the program. This result is reflective of the observed trend of slightly higher stream health in autumn than in spring. However, Autumn 2011 equalled the lowest autumn score recorded during the program. The previous autumn result was the second highest recorded during the monitoring program, and these changes would be related to slight seasonal variation in stream health.

Results from SIMPER indicated a seasonal change in macroinvertebrate assemblages in Archers Creek. Archers Creek has higher contributions from tolerant insects in autumn than in spring. The dominant insect and non-insect taxa of Archers Creek are all tolerant. The insects tend to have slightly higher SIGNAL S-F and SIGNAL2 grades than the non-insects and this may explain the trend that appears in the univariate analyses.

The Archers Creek MDS ordination plots and SIMPROF dendrogram generally grouped seasons separately. Despite the marginal seasonal variation, no significant shift in community assemblages has been observed during the program by the multivariate or univariate analyses. With this in mind it is not

possible to link any currently observed in-stream health shifts to past rehabilitation work on Archers Creek.

Shrimptons Creek

The univariate analyses suggest that Shrimptons Creek has, marginally, the poorest stream health of the five creeks in the program. The exception is the AUSRIVAS OE0 SIGNAL2 spring model output. The average score range overlaps with that of the other four creeks. It does, however, have the largest average score range through time of the five creeks for this model output.

In Autumn 2005, Shrimptons Creek returned the lowest SIGNAL S-F and AUSRIVAS OE50 autumn average stream health for any creek during the program. Shrimptons Creek average stream health then improved each season after the Autumn 2005 result, peaking in Autumn 2007. The average scores in Autumn 2007 were the highest for that creek during the program and fell within the upper range of stream health for the four other creeks. Stream health dropped significantly in Spring 2007 and Autumn 2008, results since have remained similar for the duration of the program.

Despite the recent stability in stream health, Shrimptons Creek has the most varied health through time of the five creeks. This variation was clearly indicated by the MDS ordination of all five creeks. Shrimptons Creek samples are clearly separated from those of other creeks'. The SIMPROF ordination separates Shrimptons Creek samples from all other creeks in what can be considered a 'real' difference in community structure.

Dissolved oxygen concentrations in a water body are one of the best indicators for the survival potential of aquatic organisms. It is significant that all of Shrimptons Creek sites have historically had very low dissolved oxygen levels making it the poorest of the five creeks of the program. Improvement in stormwater transportation and in-stream conditions of Shrimptons Creek may improve dissolved oxygen concentrations and stream health, as observed by macroinvertebrates and water quality results. However dissolved oxygen concentrations will not be the only driver limiting stream health and an improvement in this measure may not be reflected in macroinvertebrate assemblages.

Buffalo Creek

Buffalo Creek's SIGNAL S-F average score in Autumn 2011 showed a continuing improvement in stream health that has been observed since its poorest recording in Spring 2008. The AUSRIVAS OE50 spring and autumn models have also indicated an improvement in stream health since Spring 2008, when it recorded its poorest result (the only time the average score fell into Band D).

SIMPER results indicated a change in community structure in Spring 2008, with just three taxa contributing to 80% of the overall macroinvertebrate assemblage. These taxa were the Aquatic Snails (Physidae and Hydrobiidae) and the non-biting midge (Chironominae), all tolerant taxa. SIMPER results since showed that the range of taxa contributing to the overall macroinvertebrate assemblages has reflected what was previously recorded.

These results indicated that a new impact had limited stream health in Buffalo Creek to levels not previously observed by the program. In Autumn 2008, elevated levels of turbidity were present and observed during a site inspection. A significant build-up of sediment at the core Buffalo Creek site was observed the following Spring 2008 season.

It was suggested in the Spring 2008 report (Sydney Water, 2008b) that the loss of taxa and decline in stream health resulted from a smothering effect by fine sediment that had run-off from development in the upper catchment. This smothering has been linked to the loss of certain taxa in streams that have had an influx of fine sediment within forestry areas (Vuori & Joensuu, 1996; Death et al., 2003), which coincided with the dominance of new taxa. Death et al. (2003) found that dominant sensitive mayfly taxa were lost and that tolerant (including Chironomidae and Hydrobiidae) taxa achieved dominance when elevated levels of fine sediment were introduced to streams.

The loss of taxa and drop in stream health in Buffalo Creek could be reversed if the source of sediment was controlled or was only a short-term impact. Wood & Armitage (1997) suggested that short-term increases in fine sediment due to human disturbances, such as construction developments, could precede a rapid recovery. Results since Spring 2008 suggest that a recovery has occurred and that the impact was short term.

Porters Creek

Porters Creek Autumn 2011 SIGNAL S-F average score was the highest returned for that creek during the sampling program. The Autumn 2011 AUSRIVAS OE0 SIGNAL2 combined season average score was also the highest during the sampling program for not only Porters Creek but for all creeks. The AUSRIVAS OE50 combined season average score place it in the significantly impaired band for the second year in a row.

Porters Creek has shown a seasonal trend of marginally higher stream health in Autumn than in Spring. This trend is evident in both SIGNAL S-F average scores and AUSRIVAS OE50, the latter showing this trend for all five creeks of the program. SIMPER results indicate higher abundances of tolerant insects in Autumn and higher abundances of tolerant non-insects in Spring. As for the trend in Archers Creek, there are slightly higher SIGNAL S-F scores for insects than for non-insects in Porters Creek. Multivariate results for Porters Creek suggest that there is little variation in its macroinvertebrate assemblages through time. The variation that does occur is a general separation of samples from autumn and spring.

Terrys Creek

The macroinvertebrate results for Terrys Creek have shown very little variation through time during the monitoring program. The SIGNAL S-F average score decreased in Autumn 2011, recording its lowest autumn score since the start of the program. The same trend occurred in both the Autumn edge AUSRIVAS OE50 and AUSRIVAS OE0 SIGNAL2 models, with the autumn results returned for Terrys Creek being the lowest of the program.

The multivariate results in the form of the MDS ordination and SIMPROF dendrogram show little variation in Terrys Creek macroinvertebrate assemblages. In Autumn 2010 samples separated in the SIMPROF dendrogram and in the respective MDS ordination, however, Autumn 2011 samples were similar to those previously sampled during the monitoring program. SIMPER results for Autumn 2011 also complemented these observations.

Combined Creeks

The univariate and multivariate results indicate that all five creeks of the monitoring program have shown similar stream health when compared to one another and amongst seasons. Similarly, not many significant shifts in macroinvertebrate community assemblages have occurred. Exceptions have occurred in both Shrimptons and Buffalo creeks, which have been indicated by most of the data analyses. These were not linked to capital works on the creeks. This means that, as yet, no significant impact has been observed from creek rehabilitation work carried out by Ryde Council.

Some observations of the univariate analyses produce limitations on reporting. AUSRIVAS OE0 SIGNAL2 is at times contradictory to the other analyses. The Spring/Autumn seasonal trend in Archers and Porters creeks is an example of this. The most notable limitation occurs with the AUSRIVAS autumn edge model output for Archers Creek. The output describes the data being outside the experience of the model, resulting in three missing data points for the Autumn model and two combined season outputs. This limits the ability to compare and track changes in stream health for that creek. The combination of the physical and biological data was not typical of reference material used by the AUSRIVAS Autumn eastern edge model. Changes to the stream channel of Archers Creek, combined with few AUSRIVAS reference sites situated in the Sydney region, may explain the result.

The attempt to link water quality patterns to macroinvertebrate patterns using the multivariate BIOENV routine produced weak to mild correlations for each individual creek, and the highlighted variables were varied.

The strongest BIOENV result of the Autumn 2011 period was at Porters Creek, returning 0.364. This is only a mild correlation, the rest of the creeks returned only weak correlations. A stronger correlation would be needed to suggest any direct connection between the water quality variables and the macroinvertebrate community assemblages, as assessed in the program. The BIOENV result for all five creeks was mild (0.396) and highlighted total oxidised nitrogen, conductivity, dissolved oxygen, ratio of total length of pipe/catchment area, and ratio of number of outlets/catchment area.

The weak to mild correlations of these extrinsic variables suggest that the respective macroinvertebrate community structures of each creek are not predominantly influenced by these water quality variables as measured. This suggests that physico-chemical analytes measured under the strategy are not the only drivers of the shifts recorded in macroinvertebrate community structure. As such, efforts to improve water quality should not solely concentrate on the variables measured during the monitoring program.

Archers and Shrimptons Creeks have had relatively extensive stream and riparian zone restoration during the monitoring program, yet macroinvertebrate and water quality results have not indicated any significant improvement. Selvakumar et al (2010), whilst conducting a study into the effectiveness of stream and bank restoration of a section of an urban catchment found no significant improvement into stream health. Managing the health of our aquatic ecosystems with stream and riparian restoration as a main driver may not deliver improvements in stream health without a consideration of the whole catchment.

Research conducted in the greater Melbourne area that looked at water quality, epilithic diatoms, benthic algae and macroinvertebrate indicators suggested that minimisation of directly piped stormwater drainage connection of impervious surfaces was beneficial in mitigation of urban impacts on receiving streams (Hatt et al., 2004; Walsh, 2004; Taylor et al. 2004; Newall & Walsh, 2005). The primary degrading process in urban streams was suggested to be effective imperviousness (the proportion of a catchment covered by impervious surfaces directly connected to the stream by stormwater pipes) (Walsh et al., 2005a). This is based on the condition that sewer overflows, sewage treatment plant discharges, or long-lived pollutants from earlier land uses are not operable, as these can obscure stormwater impacts (Walsh et al., 2005b). Walsh (2004) determined that community composition was strongly explained by the gradient of urban density, observing that most sensitive taxa were absent from urban sites with greater than 20% connection of impervious surfaces to streams by pipes.

The direct connection of impervious surfaces to a stream allows small rainfall events to produce surface runoff that causes frequent disturbance to the stream through regular delivery of water and pollutants (Walsh et al., 2005a). In catchments with existing drainage networks such as those in the City of Ryde, policies that facilitate infiltration, evaporation and transpiration or storage for later in-house use will gradually benefit stream health in the longer term, based on outcomes of the research conducted in Melbourne.

6 Comments on progress of strategy aims

This section places the current knowledge of sampling data consolidated in this report within the context of the aims detailed in the City of Ryde's request for the engagement of consultants No COR-RFQ-29/09.

- Evaluate chemical and biological water quality monitoring both for short and long term interpretation and temporal evaluation over the duration of the strategy;

Consolidation of available comparable data was conducted in the Spring 2006 report. Analysis of all data in reports after Spring 2006 has also incorporated this available comparable historical data. Sampling across all five streams during the monitoring program has allowed statistical analysis to identify temporal shifts in community structure across seasons and under varying climatic conditions. Providing a better understanding of variation between autumn and spring seasons and between different climatic conditions. This has provided better base line data to assess changes in community structure that may result from future City of Ryde management actions for the duration of the program and also for future programs.

- Detail where, when and how often samples should be taken from creeks within the Ryde Local Government Area based on existing site data, catchment position, accessibility and trends identified;

Recommendations made in the Spring 2006 report to sample all creeks in each sample session have been implemented. Allowing capture of the natural variation through time and under different weather conditions that influence the five study streams. Benefits of sampling all five creeks are detailed above and in the last paragraph of this section. It was also suggested to move from three sampling occasions per season to two, and this was implemented.

- Prescribe how to sample for macroinvertebrates, building on the standard protocols designed by AUSRIVAS;

Adoption of a standard methodology under the strategy allows for collection of comparable data and in turn statistical analysis of comparable measures, which facilitates interpretation of collected data.

- Provide for a series of options for identification of key indicator taxa to family and or morphospecies;

This is provided by EPT indicator taxa from AUSRIVAS predicted model output. SIGNAL-SF grades could also be used to assess key indicator taxa. With only two EPT indicator taxa recorded during the monitoring program the use of SIGNAL-SF scores in this way were not considered.

- Identify a standard suite of analyses to determine status and trends in water quality including calculation of the AUSRIVAS index;

Suitable indices, such as SIGNAL SF to assess water quality status including calculation of the Observed/Expected (OE50 and OE0 SIGNAL2) ratios from the respective AUSRIVAS predictive models for autumn, spring, and combined seasons were evaluated in Spring 2006 with subsequent recommendation made and these have been implemented. Multivariate statistical analysis techniques have also been incorporated into Spring 2006 to the current Autumn 2011 reports. A change was made to the routine used to assess water quality and macroinvertebrates linkages in Spring 2006 with the BIOENV routine employed instead of the BVSTEP routine which conducts a less thorough search. This change was made given the relatively small amount of water quality variables and suitable computing power was available to conduct a full search of the data with BIOENV. In the Spring 2008 report the SIMPROF test was added, due to recent advances in multivariate statistical software.

- Provide the basis for an appraisal of the capacity of a standard monitoring strategy to be integrated into a community-monitoring program eg. Streamwatch.

Suggestions were put forward in the Spring 2006 and Autumn 2007 reports for use of SIGNAL2 in a format that could be calculated by community groups without access to the AUSRIVAS predictive models. In the 2007 report, calibration was made for boundary points of water quality status so community groups could use this analysis in the City of Ryde area. Standard collection methods would need to be used and suitable quality control of data would need to be implemented to provide comparability of data through time.

- Provide the foundation to augment the Streamwatch capacity within the City of Ryde including options for improved education awareness of water quality issues within schools and community groups.

As above.

- Provide information and direction on potential infrastructural works to complement water quality monitoring and improve overall creek health.

Suitable baseline data has been collated from spring and autumn season during varying weather conditions. This base line data has been used in part to assess the effectiveness of completed infrastructural work completed in creeks in the City of Ryde LGA. It will also provide the City of Ryde the ability to assess the effectiveness of future management actions.

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Appendix 1 Quality Assurance

Sydney Water Analytical Services is a quality business organisation, certified to AS/NZS ISO 9001:2008 Quality management systems - requirements certification number 18533-2008-AQ-AUS-JAS-ANZ, issued by Det Norske Veritas (DNV) on 28th December 2007 for the Monitoring Process Management System. All investigations performed for the production of this report, and all business operations of the organisation, have been conducted to the requirements of this standard including project management, macroinvertebrate sampling, water quality sampling and interpretive reporting.

Macroinvertebrates have been identified and enumerated to the genus or species taxonomic level, (as appropriate for the study) by the Aquatic Ecology team. The method used SSWI433 In-house test method macroinvertebrate cataloguing, identification and counting is in compliance with the requirements of AS ISO/IEC 17025:2005 General Requirements for the Competence of Testing and Calibration Laboratories was added under technical accreditation number 610 issued by National Association of Testing Authorities (NATA) in 1997. In particular macroinvertebrate identification was performed with appropriate published keys listed in Hawking (2000), internal keys to the macroinvertebrate collection, unpublished descriptions and voucher specimens. Where a specimen could not be keyed to a formally described species, a morphospecies number has been assigned as per in-house test method SSWI433. Terrestrial macroinvertebrate morphospecies have been shown to produce similar patterns to those obtained using formally described species (Oliver and Beattie, 1996).

Quality assurance was conducted as per SSWI434 In-house test method quality control of macroinvertebrate identification, counting and archiving of collections in compliance with the requirements of AS ISO/IEC 17025:2005 General Requirements for the Competence of Testing and Calibration Laboratories was added under technical accreditation number 610 in 1997. Quality assurance was conducted on at least 5% of samples collected for this study, and identification and counting errors on average are less than 10% for the study.

Appendix 2 Water Quality Results

Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Terrys Ck	Site 1	Autumn 2011	08/04/11	210	5	170	33	260	430	42.9	1.58	243	138	7.19	7.9	16.8
Shrimptons Ck	Site 2	Autumn 2011	08/04/11	360	5	190	40	240	430	38.9	2	196	111	6.98	6.2	17.4
Porters Ck	Site 3	Autumn 2011	08/04/11	590	330	930	34	620	1550	77.1	1.08	389	228	7.16	10.2	19
Buffalo Ck	Site 4	Autumn 2011	08/04/11	350	20	210	32	310	520	52.6	2.57	369	209	7.12	6.8	17.1
Archers Ck	Site 5	Autumn 2011	08/04/11	310	5	190	42	300	490	68.7	1.14	376	211	6.98	5.2	16.5
Terrys Ck	Site 1	Autumn 2011	04/03/11	120	1030	30	243	1650	1680	94.5	2.72	552	309	7.25	5.5	20
Shrimptons Ck	Site 2	Autumn 2011	04/03/11	400	10	60	35	340	400	35.4	0.75	186	113	7.28	6.2	20.6
Porters Ck	Site 3	Autumn 2011	04/03/11	240	210	1270	19	610	1880	58.8	1.93	299	184	7.56	8.2	20.2
Buffalo Ck	Site 4	Autumn 2011	04/03/11	40	40	70	39	410	480	65.8	2.18	792	478	7.22	5.8	20.8
Archers Ck	Site 5	Autumn 2011	04/03/11	340	50	10	65	600	610	82.8	2.02	404	240	7.15	6.3	24.8
Terrys Ck	Site 1	Spring 2010	04/11/10	230	30	480	35	350	830	61.9	5.05	420	251	7.09	8.6	15.8
Shrimptons Ck	Site 2	Spring 2010	04/11/10	330	50	670	44	580	1250	65	5.89	458	280	6.88	6.6	16.9
Porters Ck	Site 3	Spring 2010	04/11/10	350	400	1050	26	970	2020	89.0	3.05	451	260	7.34	9.1	16.4
Buffalo Ck	Site 4	Spring 2010	04/11/10	270	30	1090	29	470	1560	69.2	6.92	523	332	7.37	9.1	16.4
Archers Ck	Site 5	Spring 2010	04/11/10	690	10	980	44	490	1470	71.8	5.42	700	418	6.56	6.8	16.1
Terrys Ck	Site 1	Spring 2010	30/09/10	280	870	90	136	1360	1350	69.7	2.27	582	461	7.40	7.6	12.4
Shrimptons Ck	Site 2	Spring 2010	30/09/10	450	50	190	41	340	530	57.8	3.33	325	189	7.20	4.6	17.7
Porters Ck	Site 3	Spring 2010	30/09/10	40	940	1420	30	1350	2770	85.3	3.89	439	360	7.65	10.5	13.7
Buffalo Ck	Site 4	Spring 2010	30/09/10	240	10	250	28	520	770	73.0	4.36	751	538	7.30	8.3	13.2
Archers Ck	Site 5	Spring 2010	30/09/10	29	50	90	20	470	560	81.3	1.64	1562	1159	6.86	6.8	12.8
Terrys Ck	Site 1	Autumn 2010	15/04/10	38	10	230	31	310	540	59	1.51	328	208	7.21	6.9	15.2
Shrimptons Ck	Site 2	Autumn 2010	15/04/10	200	40	90	57	310	400	70	3.54	306	177	7.16	4.6	16.2
Porters Ck	Site 3	Autumn 2010	15/04/10	600	560	1950	18	800	2750	91	4.29	478	309	7.75	9.0	17.5
Buffalo Ck	Site 4	Autumn 2010	15/04/10	30	40	170	18	260	430	69	2.24	694	412	7.44	8.0	16.6
Archers Ck	Site 5	Autumn 2010	15/04/10	240	30	30	14	260	290	86	2.42	445	261	7.16	5.9	20.9
Terrys Ck	Site 1	Autumn 2010	15/03/10	430	60	500	21	460	960	67.9	2.03	690	433	7.13	6.5	17.9
Shrimptons Ck	Site 2	Autumn 2010	15/03/10	6200	50	200	46	500	700	71.2	4.91	515	320	7.04	4.3	18.8
Porters Ck	Site 3	Autumn 2010	15/03/10	~88000	550	1070	39	1160	2230	82.2	2.92	377	240	7.75	8.3	21.6
Buffalo Ck	Site 4	Autumn 2010	15/03/10	1800	60	190	48	460	650	52.2	3.17	400	250	7.06	6.7	20.3
Archers Ck	Site 5	Autumn 2010	15/03/10	290	30	310	22	380	690	72	1.3	410	240	7.05	5.2	19.7
Terrys Ck	Site 1	Spring 2009	02/11/09	320	190	200	70	540	740	57.8	40.1	329	187	7.58	6.5	13.0
Shrimptons Ck	Site 2	Spring 2009	02/11/09	490	<10	<10	243	1290	1290	69.6	8.7	381	219	7.54	3.7	15.2

Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Porters Ck	Site 3	Spring 2009	02/11/09	280	810	1510	16	1050	2560	73.2	3.7	388	219	8.24	9.6	17.3
Buffalo Ck	Site 4	Spring 2009	02/11/09	~160	20	60	53	370	430	84.2	4.3	880	486	8.01	7.9	17.3
Archers Ck	Site 5	Spring 2009	02/11/09	500	100	20	39	380	400	57.2	3.1	280	161	6.94	3.4	13.5
Terrys Ck	Site 1	Spring 2009	30/09/09	39	20	170	31	260	430	61.9	4.0	482	263	7.21	6.2	18.0
Shrimptons Ck	Site 2	Spring 2009	30/09/09	280	50	280	48	400	680	74.3	4.6	462	275	7.18	5.6	19.6
Porters Ck	Site 3	Spring 2009	30/09/09	6700	810	1200	39	1180	2380	92.2	8.6	442	199	7.81	8.4	19.6
Buffalo Ck	Site 4	Spring 2009	30/09/09	570	70	290	37	430	720	87.7	4.7	758	424	7.40	7.5	22.2
Archers Ck	Site 5	Spring 2009	30/09/09	640	40	390	34	340	730	53.6	2.9	327	187	7.51	9.3	25.0
Terrys Ck	Site 1	Autumn 2009	19/03/09	67	10	260	25	350	610	72.0	2.9	525	282	7.60	7.2	18.0
Shrimptons Ck	Site 2	Autumn 2009	19/03/09	1200	<10	90	43	510	600	70.1	2.8	377	220	7.34	0.2	19.4
Porters Ck	Site 3	Autumn 2009	19/03/09	3000	820	1290	27	1490	2780	106.0	2.9	487	266	7.75	8.3	20.4
Buffalo Ck	Site 4	Autumn 2009	19/03/09	240	20	580	31	520	1100	89.0	7.0	886	490	7.33	4.7	17.8
Archers Ck	Site 5	Autumn 2009	19/03/09	4800	1220	1380	171	1760	3140	78.5	2.2	517	278	7.42	5.8	17.8
Terrys Ck	Site 1	Autumn 2009	01/05/09	140	<10	180	20	240	420	64.8	2.1	518	300	7.55	7.9	12.5
Shrimptons Ck	Site 2	Autumn 2009	01/05/09	350	<10	140	34	340	480	81.5	2.1	481	289	7.45	7.4	14.5
Porters Ck	Site 3	Autumn 2009	01/05/09	~190	860	1350	21	1010	2360	86.3	4.0	449	268	7.75	9.4	16.0
Buffalo Ck	Site 4	Autumn 2009	01/05/09	92	<10	330	20	310	640	72.2	4.3	708	408	7.53	7.8	14.0
Archers Ck	Site 5	Autumn 2009	01/05/09	~1700	<10	860	31	270	1130	67.3	2.7	472	269	7.84	8.5	12.8
Terrys Ck	Site 1	Spring 2008	16/09/08	~820	10	120	35	370	490	41.5	11.5	254	149	7.20	7.8	14.6
Shrimptons Ck	Site 2	Spring 2008	16/09/08	240	20	250	54	440	690	51.0	8.9	278	155	7.10	3.8	16.1
Porters Ck	Site 3	Spring 2008	16/09/08	260	4000	1660	24	4520	6180	130.0	5.5	611	336	7.70	9.6	14.7
Buffalo Ck	Site 4	Spring 2008	16/09/08	820	10	450	42	400	850	79.5	10.8	524	293	7.34	7.2	14.9
Archers Ck	Site 5	Spring 2008	16/09/08	270	10	670	19	350	1020	82.5	2.7	555	311	7.67	10.4	13.7
Terrys Ck	Site 1	Spring 2008	13/10/08	~80	20	140	52	440	580	74.0	3.0	509	281	7.13	3.6	14.1
Shrimptons Ck	Site 2	Spring 2008	13/10/08	420	120	30	197	900	930	67.0	3.9	301	171	7.14	0.0	16.8
Porters Ck	Site 3	Spring 2008	13/10/08	48	980	1870	26	1410	3280	91.5	4.9	456	251	7.40	7.3	16.3
Buffalo Ck	Site 4	Spring 2008	13/10/08	~84	130	90	41	540	630	96.5	13.2	1008	573	7.16	0.3	17.1
Archers Ck	Site 5	Spring 2008	13/10/08	220	50	380	33	370	750	85.5	2.7	501	279	7.25	3.4	16.5
Terrys Ck	Site 1	Autumn 2008	03/05/08	150	10	270	24	310	580	71.5	3.2	474	284	8.00	8.4	21.9
Shrimptons Ck	Site 2	Autumn 2008	03/05/08	200	10	10	53	670	680	74.0	3.2	358	214	7.40	5.8	17.3
Porters Ck	Site 3	Autumn 2008	03/05/08	530	250	430	38	1100	1530	81.0	15.2	650	444	7.60	6.7	19.3
Buffalo Ck	Site 4	Autumn 2008	03/05/08	620	40	450	35	370	820	91.0	37.2	885	552	8.10	6.8	21.0

Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Archers Ck	Site 5	Autumn 2008	03/05/08	170	30	370	20	290	660	77.5	2.2	513	310	7.30	6.5	19.8
Terrys Ck	Site 1	Autumn 2008	04/03/08	250	10	120	25	200	320	64.0	3.1	351	160	7.32	8.3	15.7
Shrimptons Ck	Site 2	Autumn 2008	04/03/08	700	10	10	92	620	620	73.0	6.2	291	130	7.16	3.8	16.8
Porters Ck	Site 3	Autumn 2008	04/03/08	370	750	300	27	1100	4100	100.0	4.0	505	290	7.56	9.3	16.9
Buffalo Ck	Site 4	Autumn 2008	04/03/08	120	50	220	33	260	480	77.0	4.7	654	389	7.30	8.0	15.8
Archers Ck	Site 5	Autumn 2008	04/03/08	160	40	110	22	230	340	83.0	1.5	470	253	7.28	7.1	16.7
Terrys Ck	Site 1	Spring 2007	27/09/07	87	20	190	21	290	480	67.0	2.0	503	276	7.30	6.0	14.0
Shrimptons Ck	Site 2	Spring 2007	26/09/07	300	160	30	54	650	680	72.0	2.6	403	232	7.10	2.4	16.9
Porters Ck	Site 3	Spring 2007	27/09/07	1000	2600	3200	60	3110	6310	122.0	6.7	671	372	7.80	6.5	15.0
Buffalo Ck	Site 4	Spring 2007	27/09/07	54	40	170	37	440	610	90.0	7.3	960	484	7.30	5.7	19.0
Archers Ck	Site 5	Spring 2007	26/09/07	270	20	480	26	680	1160	59.0	3.2	527	304	7.50	6.3	15.1
Terrys Ck	Site 1	Spring 2007	23/10/07	6	40	80	35	730	810	88.0	1.6	712	437	7.00	4.0	15.6
Shrimptons Ck	Site 2	Spring 2007	22/10/07	150	<10	<10	111	1000	1000	77.0	11.9	519	350	6.70	2.9	19.8
Porters Ck	Site 3	Spring 2007	23/10/07	160	1020	2600	68	1580	4180	90.0	8.2	505	326	7.70	7.3	19.3
Buffalo Ck	Site 4	Spring 2007	23/10/07	140	110	60	73	790	850	108.0	7.7	1001	621	7.20	7.0	20.4
Archers Ck	Site 5	Spring 2007	22/10/07	90	150	50	57	480	530	74.0	7.1	378	220	6.70	3.9	17.3
Terrys Ck	Site 1	Autumn 2007	14-15/03/07	300	<10	370	30	280	650	64.0	1.6	472	358	7.20	5.1	18.1
Shrimptons Ck	Site 2	Autumn 2007	14-15/03/07	600	<10	550	58	330	880	64.0	2.9	362	276	7.10	3.2	20.6
Porters Ck	Site 3	Autumn 2007	14-15/03/07	600	580	1310	51	1040	2350	97.0	1.3	3030	2010	7.90	8.4	19.3
Buffalo Ck	Site 4	Autumn 2007	14-15/03/07	68	90	120	48	440	560	75.0	2.1	646	442	7.30	5.1	19.5
Archers Ck	Site 5	Autumn 2007	14-15/03/07	290	<10	170	89	270	440	64.0	0.9	397	300	7.20	4.6	20.8
Terrys Ck	Site 1	Autumn 2007	17-18/04/07	900	110	200	53	530	730	57.0	2.7	438	.	7.10	5.3	17.2
Shrimptons Ck	Site 2	Autumn 2007	17-18/04/07	550	30	160	45	490	650	81.0	8.4	397	.	6.90	3.8	17.6
Porters Ck	Site 3	Autumn 2007	17-18/04/07	10000	710	1590	20	1200	2790	98.0	3.2	3130	.	7.80	7.7	18.0
Buffalo Ck	Site 4	Autumn 2007	17-18/04/07	740	130	120	48	540	660	81.0	8.6	912	.	6.70	3.8	17.2
Archers Ck	Site 5	Autumn 2007	17-18/04/07	210	30	50	58	520	570	70.0	4.2	322	.	7.20	4.1	18.7
Shrimptons Ck	Site 2	Spring 2006	28/09/06	69	130	140	64	580	720	94.0	7.8	717	420	7.12	4.3	17.3
Archers Ck	Site 5	Spring 2006	28/09/06	160	<10	<10	104	520	520	83.0	2.0	509	293	7.37	6.5	15.4
Shrimptons Ck	Site 2	Spring 2006	18/10/06	560	10	20	136	1180	1200	66.0	6.3	481	311	6.54	2.2	17.2
Archers Ck	Site 5	Spring 2006	18/10/06	340	<10	10	90	500	510	70.0	2.3	448	295	6.93	3.9	18.3
Shrimptons Ck	Site 2	Spring 2006	10/11/06	880	70	1200	68	800	2000	58.0	96.7	384	265	7.41	4.2	17.5
Archers Ck	Site 5	Spring 2006	10/11/06	1700	20	40	50	360	400	84.0	1.8	502	310	7.21	7.2	18.6

Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Terrys Ck	Site 1	Autumn 2006	9-10/03/06	160	<10	60	30	310	370	50.0	2.3	381	180	6.80	5.0	20.2
Shrimptons Ck	Site 2	Autumn 2006	9-10/03/06	330	40	<10	50	380	390	85.0	4.6	435	230	6.70	2.1	21.1
Porters Ck	Site 3	Autumn 2006	9-10/03/06	9800	820	760	20	1500	2300	48.0	1.9	3712	2200	7.40	7.4	25.2
Buffalo Ck	Site 4	Autumn 2006	9-10/03/06	220	130	470	70	500	1000	90.0	8.0	738	390	7.20	4.4	22.1
Archers Ck	Site 5	Autumn 2006	9-10/03/06	140	90	80	100	520	600	95.0	2.5	1482	830	7.00	4.1	20.6
Terrys Ck	Site 1	Autumn 2006	19-20/04/06	560	450	90	100	1100	1200	45.0	3.2	306	180	7.00	2.4	15.7
Shrimptons Ck	Site 2	Autumn 2006	19-20/04/06	860	30	30	80	480	510	40.0	5.0	281	160	6.70	4.6	16.8
Porters Ck	Site 3	Autumn 2006	19-20/04/06	290	350	630	20	700	1300	45.0	2.3	3792	2100	7.60	8.3	19.8
Buffalo Ck	Site 4	Autumn 2006	19-20/04/06	170	90	450	60	470	920	70.0	5.1	749	400	7.20	4.6	19.2
Archers Ck	Site 5	Autumn 2006	19-20/04/06	240	90	470	70	390	860	45.0	4.1	259	150	7.10	4.4	18.4
Terrys Ck	Site 1	Autumn 2006	9-10/05/06	66	70	240	50	380	620	60.0	2.4	358	220	7.10	4.0	11.9
Shrimptons Ck	Site 2	Autumn 2006	9-10/05/06	750	20	40	80	340	380	35.0	7.7	264	140	6.80	5.0	13.1
Porters Ck	Site 3	Autumn 2006	9-10/05/06	40	400	650	10	800	1400	1.0	1.2	2916	1700	7.30	8.3	15.3
Buffalo Ck	Site 4	Autumn 2006	9-10/05/06	110	60	480	60	240	720	90.0	4.4	667	400	7.30	4.7	11.7
Archers Ck	Site 5	Autumn 2006	9-10/05/06	28	50	370	40	300	670	55.0	5.1	245	120	7.20	6.3	12.4
Terrys Ck	Site 1	Spring 2005	6-7/09/05	300	59	48	10	900	140	43.0	6.5	187	140	6.70	8.1	11.1
Shrimptons Ck	Site 2	Spring 2005	6-7/09/05	90	5	37	40	280	65	42.0	7.0	164	140	6.70	4.3	12.9
Porters Ck	Site 3	Spring 2005	6-7/09/05	500	110	58	20	2400	300	37.0	3.0	6141	4000	7.00	8.7	12.8
Buffalo Ck	Site 4	Spring 2005	6-7/09/05	16	10	50	80	270	77	79.0	5.5	620	380	7.00	6.2	13.2
Archers Ck	Site 5	Spring 2005	6-7/09/05	2000	17	26	110	560	82	56.0	10.0	245	160	6.80	5.6	14.7
Terrys Ck	Site 1	Spring 2005	11-12/10/05	2000	10	33	10	520	85	47.0	2.2	245	180	7.10	4.5	13.6
Shrimptons Ck	Site 2	Spring 2005	11-12/10/05	32000	16	36	100	540	90	43.0	3.9	246	150	7.20	3.3	15.7
Porters Ck	Site 3	Spring 2005	11-12/10/05	16000	54	51	50	1300	180	31.0	4.5	3965	2600	7.60	8.7	17.9
Buffalo Ck	Site 4	Spring 2005	11-12/10/05	6500	26	63	200	700	130	44.0	29.0	472	210	7.60	9.2	16.1
Archers Ck	Site 5	Spring 2005	11-12/10/05	3800	6	54	100	500	100	30.0	5.1	206	100	7.30	4.6	20.6
Terrys Ck	Site 1	Spring 2005	02/11/05	380	<1	2	40	370	39	37.0	1.0	159	110	6.50	5.4	20.8
Shrimptons Ck	Site 2	Spring 2005	02/11/05	500	6	19	60	450	64	50.0	6.1	226	150	6.60	5.2	22.2
Porters Ck	Site 3	Spring 2005	02/11/05	260	83	42	<10	2100	250	30.0	6.4	5633	3500	7.10	7.9	23.4
Buffalo Ck	Site 4	Spring 2005	02/11/05	2000	5	28	50	350	63	60.0	4.1	299	200	7.00	5.7	21.0
Archers Ck	Site 5	Spring 2005	02/11/05	640	6	18	40	560	74	79.0	12.6	350	210	6.90	5.6	25.1
Terrys Ck	Site 1	Autumn 2005	30-31/03/05	60000	590	170	100	800	970	40.0	42.0	315	130	7.20	8.4	16.9
Shrimptons Ck	Site 2	Autumn 2005	30-31/03/05	3400	20	240	40	280	520	52.0	9.0	305	170	6.70	4.5	17.1

Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Porters Ck	Site 3	Autumn 2005	30-31/03/05	1000	670	820	40	1100	1900	99.0	18.9	1719	1100	7.30	7.6	18.3
Buffalo Ck	Site 4	Autumn 2005	30-31/03/05	36	130	290	30	370	660	59.0	17.4	241	140	7.60	8.4	17.8
Archers Ck	Site 5	Autumn 2005	30-31/03/05	360	20	50	60	350	400	68.0	22.2	183	180	7.10	7.5	19.6
Terrys Ck	Site 1	Autumn 2005	26-27/04/05	90	70	140	40	300	440	62.0	1.7	264	180	6.60	6.6	15.8
Shrimptons Ck	Site 2	Autumn 2005	26-27/04/05	940	40	100	30	270	370	65.0	3.2	236	160	6.40	5.7	17.3
Porters Ck	Site 3	Autumn 2005	26-27/04/05	220	400	590	20	1100	1700	35.0	3.6	2520	1800	7.20	8.8	18.3
Buffalo Ck	Site 4	Autumn 2005	26-27/04/05	520	80	940	40	.	770	95.0	7.6	548	390	6.70	5.4	16.6
Archers Ck	Site 5	Autumn 2005	26-27/04/05	300	40	20	10	240	260	78.0	1.4	261	160	6.80	5.8	17.4
Terrys Ck	Site 1	Autumn 2005	26-27/05/05	130	40	110	30	260	370	61.0	1.8	325	180	7.30	8.3	10.8
Shrimptons Ck	Site 2	Autumn 2005	26-27/05/05	400	40	290	30	.	560	65.0	4.9	333	180	7.20	5.7	11.9
Porters Ck	Site 3	Autumn 2005	26-27/05/05	59	350	640	20	1100	1700	30.0	1.5	2305	1500	7.80	10.0	15.6
Buffalo Ck	Site 4	Autumn 2005	26-27/05/05	170	90	350	40	300	650	92.0	7.1	641	360	7.50	7.4	12.6
Archers Ck	Site 5	Autumn 2005	26-27/05/05	360	60	70	20	310	380	99.0	3.3	376	200	7.40	8.1	10.8
Terrys Ck	Site 1	Spring 2004	14-15/09/04	80	.	.	110	.	.	50.0	2.4	.	150	6.80	5.1	10.6
Shrimptons Ck	Site 2	Spring 2004	14-15/09/04	880	.	.	90	.	.	58.0	3.1	.	140	6.80	2.2	11.8
Archers Ck	Site 5	Spring 2004	14-15/09/04	650	.	.	150	.	.	70.0	0.6	.	110	7.00	6.5	13.3
Terrys Ck	Site 1	Spring 2004	11-12/10/04	44	.	.	30	.	.	64.0	0.3	.	310	7.60	5.0	16.1
Shrimptons Ck	Site 2	Spring 2004	11-12/10/04	110	.	.	60	.	.	76.0	0.5	.	260	7.40	5.7	18.5
Archers Ck	Site 5	Spring 2004	11-12/10/04	1500	.	.	50	.	.	82.0	0.8	.	230	7.50	4.3	18.6
Terrys Ck	Site 1	Spring 2004	23-24/11/04	150	.	.	40	.	.	56.0	2.6	.	180	6.70	6.9	15.5
Shrimptons Ck	Site 2	Spring 2004	23-24/11/04	1000	.	.	90	.	.	75.0	11.5	.	190	6.40	2.9	17.0
Archers Ck	Site 5	Spring 2004	23-24/11/04	1700	.	.	40	.	.	84.0	4.7	.	270	6.60	8.0	17.2

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Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2008	05/03/08	650	30	40	41	330	370	37.5	6.02	29.1	170	6.8	1.9	19
Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2008	03/04/08	~25	<10	20	34	230	250	37	2.58	18.4	86	7.2	6.3	16.3
Shrimpton's Creek at Kent Rd	CR1SA	Spring 2008	16/09/08	~1100	40	210	59	410	620	41.5	10.1	28.7	157	7.3	5.2	14.4
Shrimpton's Creek at Kent Rd	CR1SA	Spring 2008	13/10/08	220	120	20	101	690	710	63.5	4.4	36.5	208	6.9	0.2	17
Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2009	19/03/09	2300	10	60	41	440	500	60.1	3.88	46.9	255	7.2	4.5	18.8
Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2009	01/05/09	570	<10	50	25	280	330	69.5	1.72	59.7	350	7.2	5.9	14.8
Shrimpton's Creek at Kent Rd	CR1SA	Spring 2009	30/09/09	74	10	10	144	980	990	55.2	7.92	30	157	7.07	1.7	13.3
Shrimpton's Creek at Kent Rd	CR1SA	Spring 2009	02/11/09	240	60	100	32	420	520	66.5	4.32	64.7	364	7.06	5.4	20.3
Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2010	15/03/10	430	40	150	28	500	650	71.2	5.23	62	360	7	4.7	19
Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2010	15/04/10	240	6400	120	76	6760	6880	131	4.23	60.2	308	7.24	4	16.1
Shrimpton's Creek at Kent Rd	CR1SA	Spring 2010	30/09/10	25	20	90	22	500	590	74.8	1.88	74.6	504	7.25	6.9	14.5
Shrimpton's Creek at Kent Rd	CR1SA	Spring 2010	04/11/10	490	40	620	32	450	1070	64.7	5.36	56.2	322	6.92	8.5	16.9
Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2011	04/03/11	1000	20	10	186	1200	1210	74.7	1.14	40	222	7.01	1.7	20.1
Shrimpton's Creek at Kent Rd	CR1SA	Autumn 2011	08/04/11	470	<10	120	45	280	400	34	2.05	21.2	122	6.95	5.9	17.6
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2008	05/03/08	490	20	970	40	420	1390	101	7	88.9	548	-	8.5	-
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2008	03/04/08	4200	10	30	20	330	360	134	3.5	90.3	487	-	5.2	-
Shrimpton's Creek at Bridge St	CR1SB	Spring 2008	16/09/08	110	30	170	26	370	540	70.5	4.58	58.3	336	6.9	9.1	17.7
Shrimpton's Creek at Bridge St	CR1SB	Spring 2008	13/10/08	~110	20	10	24	350	360	93.5	5.3	61.2	345	7	2	18.2
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2009	19/03/09	38	<10	20	24	350	370	80	3.4	58.7	320	7.3	5.8	18.8
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2009	01/05/09	~90	10	50	27	320	370	87.5	3.96	73.5	426	7.2	7.7	14.8
Shrimpton's Creek at Bridge St	CR1SB	Spring 2009	30/09/09	240	40	70	365	2320	2390	63.9	447	39.4	303	6.85	5.5	15.8
Shrimpton's Creek at Bridge St	CR1SB	Spring 2009	02/11/09	500	120	170	69	700	870	76.7	18.2	99.1	560	7.19	7.1	23.9
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2010	15/03/10	510	40	300	37	580	880	70.4	8.53	48	303	7.05	5.9	19.4
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2010	15/04/10	9200	11000	20	600	12000	12000	184	5.32	68.8	358	7.26	4	16
Shrimpton's Creek at Bridge St	CR1SB	Spring 2010	30/09/10	250	30	20	10	500	520	85.5	6.09	96.5	620	7.22	8	13.4
Shrimpton's Creek at Bridge St	CR1SB	Spring 2010	04/11/10	230	30	490	17	330	820	75.2	3.26	65	380	6.87	7.4	16.6
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2011	04/03/11	54	<10	<10	14	380	380	110	1.15	103.5	616	7.14	3.8	20.3
Shrimpton's Creek at Bridge St	CR1SB	Autumn 2011	08/04/11	270	<10	20	25	250	270	59.9	1.94	39.2	211	6.85	4.4	17.4

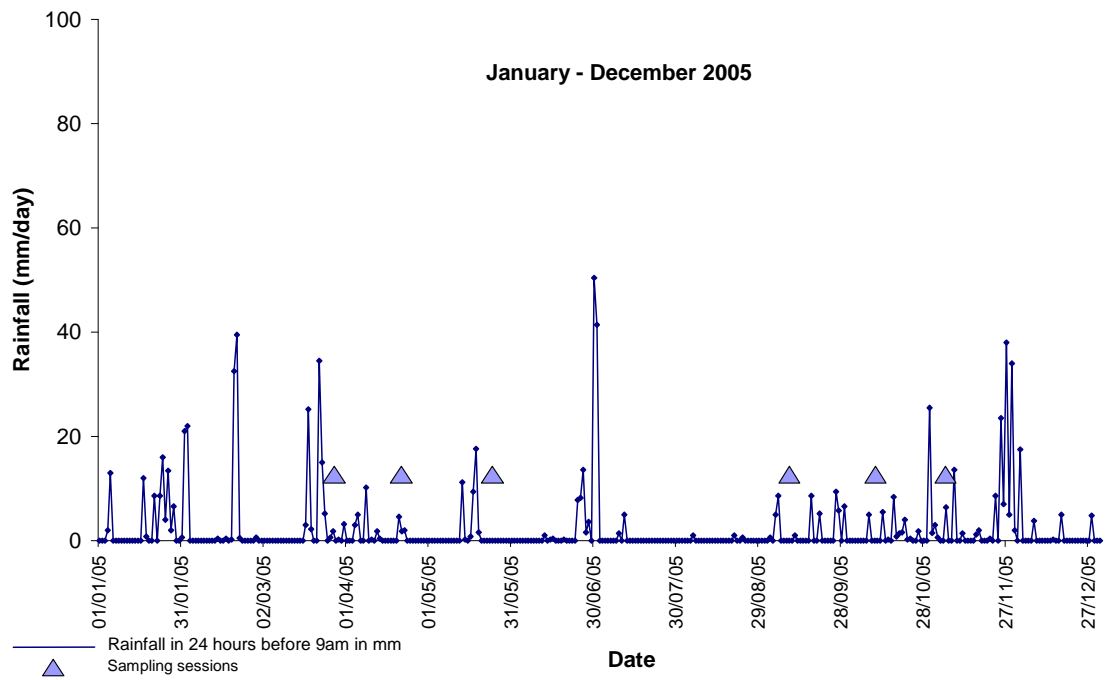
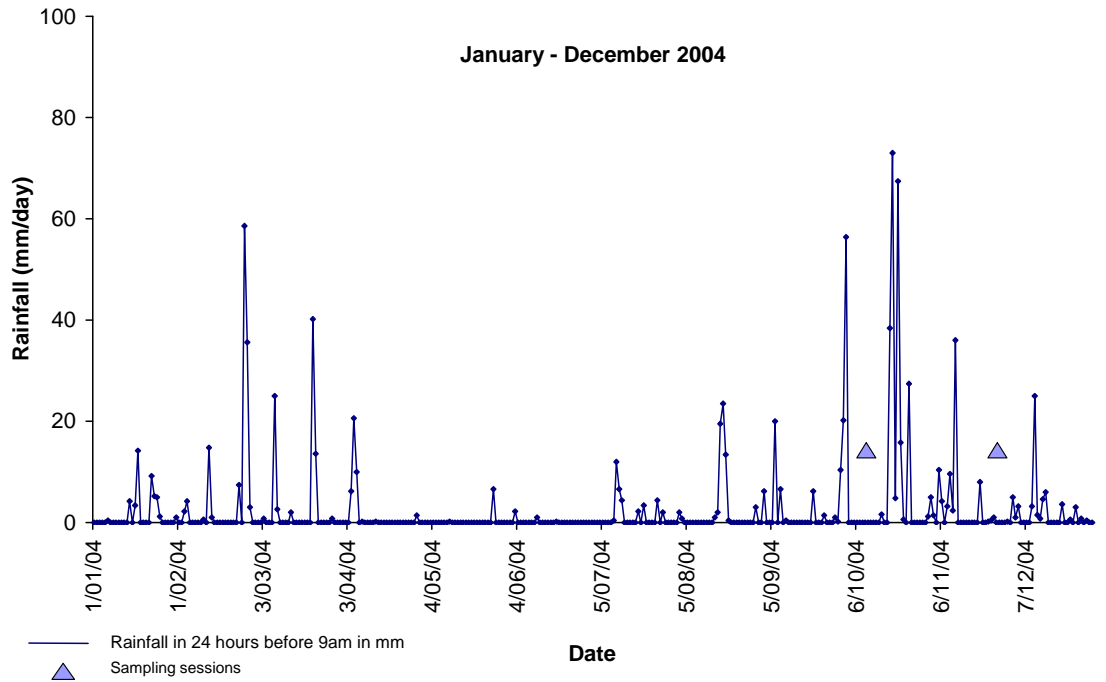
Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2008	03/04/08	4200	10	30	20	330	360	134	3.5	90.3	487	7.7	5.8	22.2
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2008	05/03/08	490	20	970	40	420	1390	101	7	88.9	548	7.2	6.7	18.1
Shrimpton's Creek at Quarry Rd	CR1SC	Spring 2008	16/09/08	~100	180	510	35	530	1040	75.5	5.66	89.9	517	7.1	6.3	17.7
Shrimpton's Creek at Quarry Rd	CR1SC	Spring 2008	13/10/08	25	270	440	165	1120	1560	112	12.4	144.9	826	6.9	0.1	17.6
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2009	19/03/09	67000	5800	670	625	6390	7060	104	2.6	76.9	396	7.5	4.4	20.4
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2009	01/05/09	~1900	<10	410	69	440	850	79.7	4.11	86.4	504	7.5	5.5	16.6
Shrimpton's Creek at Quarry Rd	CR1SC	Spring 2009	30/09/09	400	110	10	600	2620	2630	130	12	168.7	940	6.8	1.3	12.9
Shrimpton's Creek at Quarry Rd	CR1SC	Spring 2009	02/11/09	510	20	850	48	330	1180	76.6	3.58	101.3	590	7.69	8.8	24.2
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2010	15/03/10	6400	70	670	62	500	1170	99.6	3.17	87.6	530	7.33	6.5	19.6
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2010	15/04/10	23000	1000	230	416	1970	2200	88	2.58	98.4	598	7.84	9.2	17.2
Shrimpton's Creek at Quarry Rd	CR1SC	Spring 2010	30/09/10	250	<10	890	31	480	1370	69.3	1.33	127.6	848	7.64	12.1	13.2
Shrimpton's Creek at Quarry Rd	CR1SC	Spring 2010	04/11/10	~950	20	1100	28	420	1520	67	4.42	84.9	496	6.95	9.2	16.9
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2011	04/03/11	300	40	1110	102	630	1740	60.1	2.08	93.9	546	7.27	8	19.3
Shrimpton's Creek at Quarry Rd	CR1SC	Autumn 2011	08/04/11	460	30	1070	168	390	1460	75.5	1.45	79.6	444	7.06	7.4	16.7

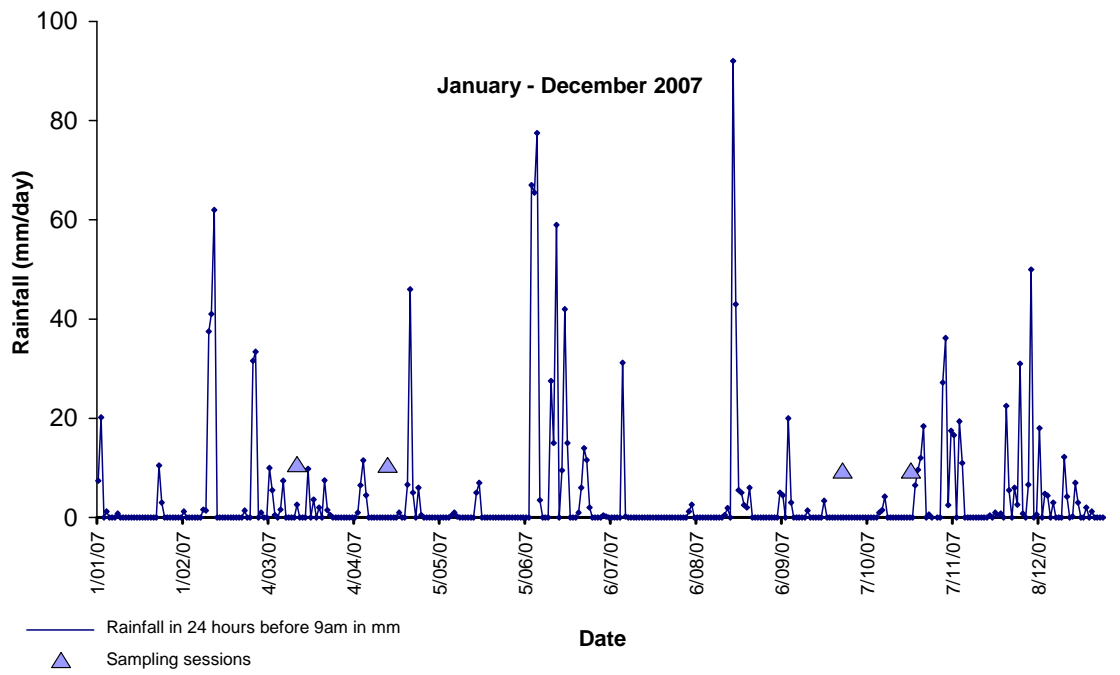
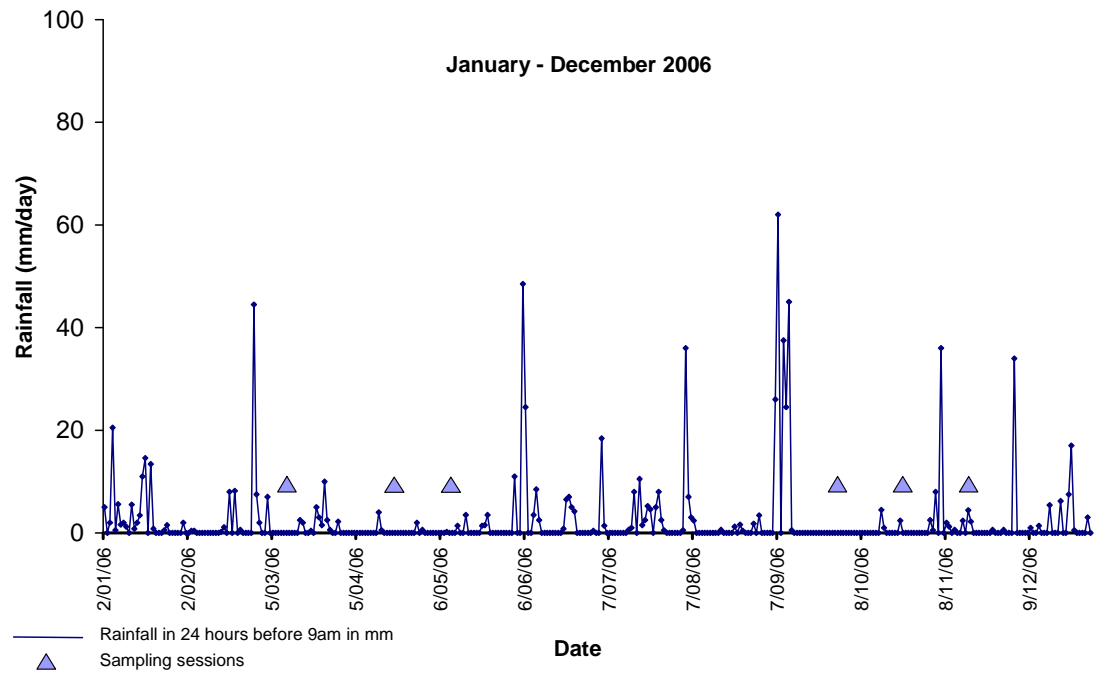
Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO ₃ /L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2008	05/03/08	~4500	<10	540	45	410	950	84	72.2	217.3	1320	7.3	6.9	21.8
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2008	03/04/08	~860	-	-	-	-	-	-	-	-	-	7.1	9.2	17.7
Buffalo Creek d/s of Burrows Park	CR4BA	Spring 2008	16/09/08	1200	40	1790	51	530	2320	86.5	30.4	123.9	696	7.2	7.2	15.7
Buffalo Creek d/s of Burrows Park	CR4BA	Spring 2008	13/10/08	760	50	60	63	670	730	116	17	182	1034	6.9	7.8	17.2
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2009	19/03/09	660	60	900	43	510	1410	95.1	7.69	157.2	882	7.3	6.3	18.5
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2009	01/05/09	~820	<10	680	35	330	1010	106	7.25	140.3	765	7.3	8.5	16.1
Buffalo Creek d/s of Burrows Park	CR4BA	Spring 2009	30/09/09	490	<10	30	107	430	460	120	7.93	167.9	934	7.73	6.8	16.8
Buffalo Creek d/s of Burrows Park	CR4BA	Spring 2009	02/11/09	460	100	1030	69	490	1520	109	11.7	108.9	626	7.18	8	21.7
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2010	15/03/10	~950	20	130	83	570	700	73.4	5.3	80.1	498	6.87	6	21.2
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2010	15/04/10	240	40	120	27	390	510	101	5.32	113.6	712	7.04	7.6	18.2
Buffalo Creek d/s of Burrows Park	CR4BA	Spring 2010	30/09/10	5200	<10	550	33	660	1210	85.1	5.88	109.6	786	7.23	8.6	13.8
Buffalo Creek d/s of Burrows Park	CR4BA	Spring 2010	04/11/10	~980	30	1660	35	450	2110	78.3	5.44	79.9	512	7.13	9.6	17.1
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2011	04/03/11	590	<10	<10	44	490	490	105	0.75	180.6	1076	7.03	7.5	22.4
Buffalo Creek d/s of Burrows Park	CR4BA	Autumn 2011	08/04/11	~950	<10	510	43	300	810	79.5	3.08	79.6	454	7	7.6	17.9
Buffalo Creek u/s of Burrows Park	CR4BB	Autumn 2008	05/03/08	>10000	20	890	71	520	1410	88	87.3	76.3	468	7.5	6.5	21.7
Buffalo Creek u/s of Burrows Park	CR4BB	Autumn 2008	03/04/08	600	-	-	-	-	-	-	-	-	-	7.7	11.1	17
Buffalo Creek u/s of Burrows Park	CR4BB	Spring 2008	16/09/08	410	50	1610	31	380	1990	87.5	11.4	107.1	602	7.8	9.3	14
Buffalo Creek u/s of Burrows Park	CR4BB	Spring 2008	13/10/08	410	100	680	106	780	1460	124	9.72	151.1	840	7.3	2.4	17.2
Buffalo Creek u/s of Burrows Park	CR4BB	Autumn 2009	19/03/09	490	60	1240	48	430	1670	102	2.68	130.8	742	8.9	7.8	18.3
Buffalo Creek u/s of Burrows Park	CR4BB	Autumn 2009	01/05/09	1500	<10	850	62	300	1150	104	8.19	118.2	658	7.7	8.1	15.6
Buffalo Creek u/s of Burrows Park	CR4BB	Spring 2009	30/09/09	340	40	370	61	380	750	92.5	2.85	100.2	530	8.17	8.3	15.5
Buffalo Creek u/s of Burrows Park	CR4BB	Spring 2009	02/11/09	400	20	1420	118	690	2110	118	32.1	110.9	630	8.03	7.9	20.4
Buffalo Creek u/s of Burrows Park	CR4BB	Autumn 2010	15/03/10	540	<10	480	75	410	890	108	1.54	93.4	568	7.65	7.7	20.1
Buffalo Creek u/s of Burrows Park	CR4BB	Autumn 2010	15/04/10	440	10	510	53	340	850	107	1.52	84.6	532	7.75	8.4	17.7
Buffalo Creek u/s of Burrows Park	CR4BB	Spring 2010	30/09/10	260	<10	1230	35	450	1680	84.9	4.32	88.4	634	7.31	8.4	13.9
Buffalo Creek u/s of Burrows Park	CR4BB	Spring 2010	04/11/10	790	20	1870	33	460	2330	84.3	2.94	75.6	486	7.21	9.2	16.3
Buffalo Creek u/s of Burrows Park	CR4BB	Autumn 2011	04/03/11	270	<10	<10	61	490	490	102	1.74	103.8	628	7.36	6.4	20.9
Buffalo Creek d/s of Burrows Park	CR4BB	Autumn 2011	08/04/11	~1700	<10	620	50	440	1060	88.1	0.71	70.2	392	7.36	9.5	17.7

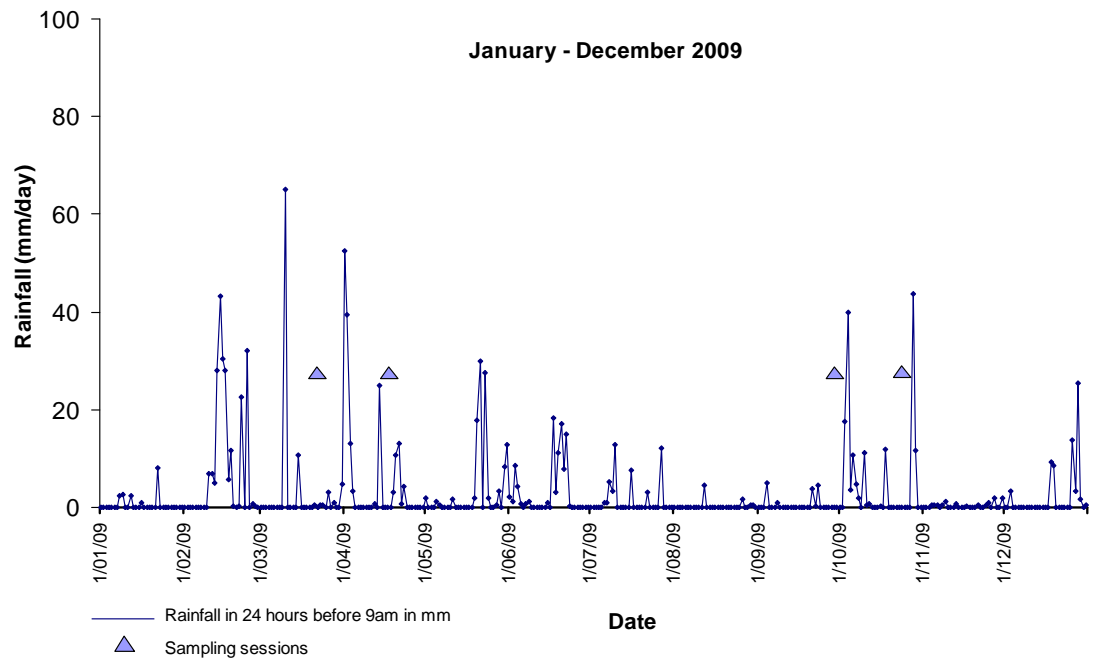
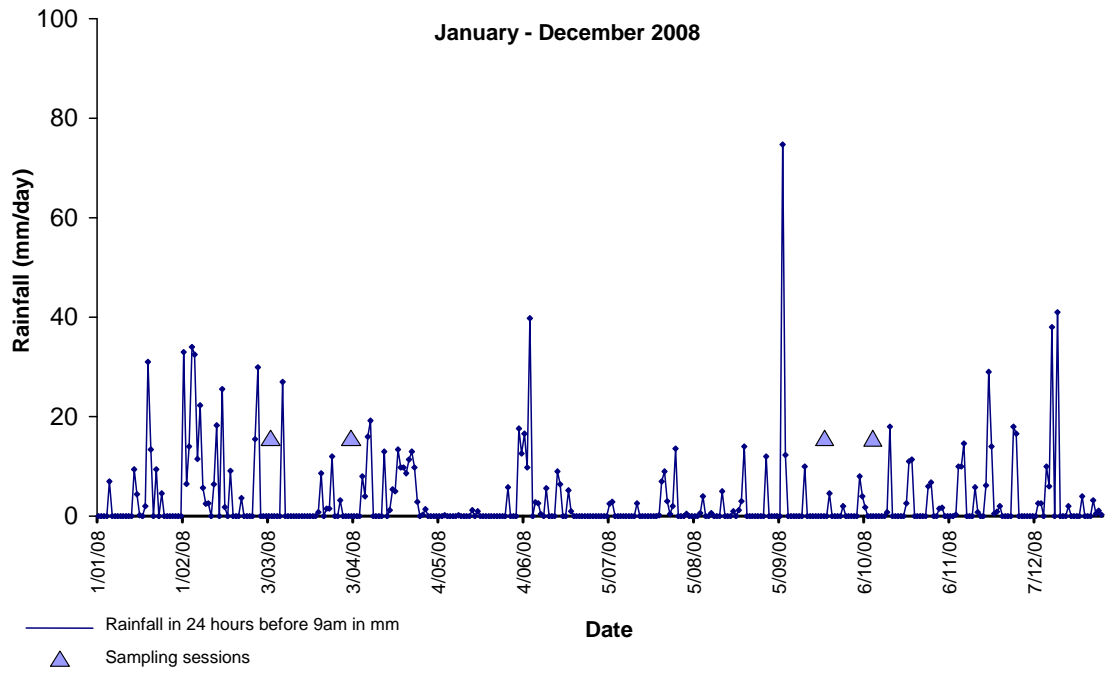
Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Porters Creek main branch	CR5Pa	Autumn 2008	05/03/08	210	110	120	43	370	490	55.5	19.5	15.2	92	7.1	8.1	23
Porters Creek main branch	CR5Pa	Autumn 2008	03/04/08	62	70	80	37	420	500	60	2.88	15.4	70	7	6.3	17.2
Porters Creek main branch	CR5Pa	Spring 2008	16/09/08	46	250	350	41	920	1270	240	12.7	61.1	350	7	9.1	13.8
Porters Creek main branch	CR5Pa	Spring 2008	13/10/08	~4	60	170	24	750	920	414	3.13	97.4	569	7.4	6.6	17.6
Porters Creek main branch	CR5Pa	Autumn 2009	19/03/09	210	40	150	39	320	470	32.9	11	10.5	61	7	7.6	20.9
Porters Creek main branch	CR5Pa	Autumn 2009	01/05/09	~31	220	30	31	430	460	119	1.99	35.3	219	7.2	2.5	15.1
Porters Creek main branch	CR5PA	Spring 2009	30/09/09	~14	130	550	10	580	1130	392	2.63	96.5	544	8.16	11.2	17.8
Porters Creek main branch	CR5PA	Spring 2009	02/11/09	47	30	160	27	300	460	41.1	3.77	15	84	7.43	10.9	20.3
Porters Creek main branch	CR5PA	Autumn 2010	15/03/10	47	220	30	32	760	790	82.2	2.63	23.5	140	6.84	4.8	20.8
Porters Creek main branch	CR5PA	Autumn 2010	15/04/10	280	10	380	67	290	670	90	2.24	37.3	232	7.27	7.6	16.9
Porters Creek main branch	CR5PA	Spring 2010	30/09/10	120	160	220	49	1110	1330	248	13.4	75.7	503	6.89	9.7	14.9
Porters Creek main branch	CR5PA	Spring 2010	04/11/10	340	60	250	48	420	670	32.7	9.71	11.5	50	7.16	8.6	17.2
Porters Creek main branch	CR5PA	Autumn 2011	04/03/11	~6	<10	<10	37	580	580	177	1.51	58.4	335	7.46	10	21.8
Porters Creek main branch	CR5PA	Autumn 2011	08/04/11	38	270	20	38	1000	1020	<2	3.3	80	492	6.94	3.4	18.1
Porters Creek spur branch	CR5Pb	Autumn 2008	05/03/08	410	110	260	49	540	800	106	8.63	47.4	302	7.8	7.8	21.4
Porters Creek spur branch	CR5Pb	Autumn 2008	03/04/08	~920	100	220	1530	960	1180	68	625	31.8	172	7.6	11.7	17.6
Porters Creek spur branch	CR5Pb	Spring 2008	16/09/08	~94	60	310	35	380	690	73	3.61	34.3	139	7.7	9.6	15
Porters Creek spur branch	CR5Pb	Spring 2008	13/10/08	380	40	180	42	340	520	64	2.22	31.8	191	7.7	8.2	17.2
Porters Creek spur branch	CR5Pb	Autumn 2009	19/03/09	150	70	280	20	450	730	79.5	3.35	37.9	221	7.5	9.6	20.8
Porters Creek spur branch	CR5Pb	Autumn 2009	01/05/09	46	90	330	15	380	710	65.5	5.1	34.2	195	7.6	9.8	16.7
Porters Creek spur branch	CR5PB	Spring 2009	30/09/09	~13	200	480	13	410	890	124	2.7	69.3	388	8.21	10.5	15.2
Porters Creek spur branch	CR5PB	Spring 2009	02/11/09	80	170	400	42	500	900	119	7.18	59.8	356	7.54	8.2	20.7
Porters Creek spur branch	CR5PB	Autumn 2010	15/03/10	250	250	210	49	750	960	72.1	9.63	36.5	220	7.37	8.7	20.2
Porters Creek spur branch	CR5PB	Autumn 2010	15/04/10	2000	20	150	93	1470	1620	166	22.5	72	466	8.37	9	18.7
Porters Creek spur branch	CR5PB	Spring 2010	30/09/10	~18	70	250	55	510	760	46.9	6.46	25.1	195	7.51	10.4	15.8
Porters Creek spur branch	CR5PB	Spring 2010	04/11/10	230	50	260	30	340	600	57.2	2.89	26.8	143	7.08	9.7	16.8
Porters Creek spur branch	CR5PB	Autumn 2011	04/03/11	44	30	200	18	260	460	46.6	1.11	23.9	144	7.7	8.9	20.9
Porters Creek spur branch	CR5PB	Autumn 2011	08/04/11	~89	80	280	20	260	540	47.4	1.05	24.8	141	7.14	9.3	19.8

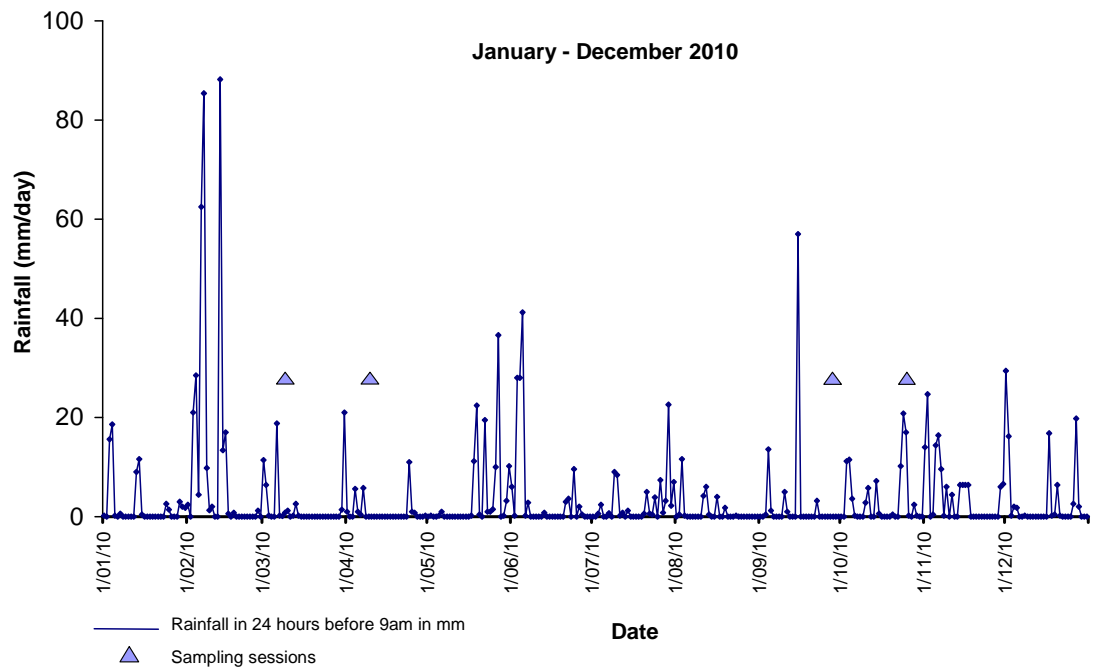
Stream	Site Code	Season	Sample Date	Faecal Coliforms CFU/100mL	Ammonia µg/L	Oxidised Nitrogen NOx µg/L	Total Phosphorus TP µg/L	Total Kjeldahl Nitrogen TKN µg/L	Total Nitrogen TN µg/L	Alkalinity mg CaCO3/L	Turbidity NTU	Conductivity µS/cm	Total Dissolved Solids mg/L	pH	Dissolved Oxygen DO mg/L	Temp. °C
Porters Creek Wicks Rd	CR5Pc	Autumn 2008	05/03/08	540	<10	110	32	630	740	74	20.7	66.1	454	8.5	7.2	22.6
Porters Creek Wicks Rd	CR5Pc	Autumn 2008	03/04/08	~120	70	1150	51	520	1670	54	31.6	71.3	385	9.1	10.8	19.5
Porters Creek Wicks Rd	CR5Pc	Spring 2008	16/09/08	510	80	1040	19	300	1340	82	8.64	51.1	591	7.7	9.5	14
Porters Creek Wicks Rd	CR5Pc	Spring 2008	13/10/08	730	100	900	116	660	1560	97.5	59.8	53.3	318	7.5	6.9	16.3
Porters Creek Wicks Rd	CR5Pc	Autumn 2009	19/03/09	~840	30	1480	27	350	1830	86	3.34	55.6	326	7.9	8.7	21.4
Porters Creek Wicks Rd	CR5Pc	Autumn 2009	01/05/09	160	30	1500	22	200	1700	83.8	2.36	54.3	334	7.7	8.5	15.7
Porters Creek Wicks Rd	CR5PC	Spring 2009	30/09/09	240	80	370	36	390	760	54.5	8.34	28.4	165	8.41	10.1	17.1
Porters Creek Wicks Rd	CR5PC	Spring 2009	02/11/09	45	70	820	23	260	1080	56.9	3.45	34.1	201	7.7	8.8	19.4
Porters Creek Wicks Rd	CR5PC	Autumn 2010	15/03/10	26	100	500	11	360	860	49	1.81	26.9	175	7.4	8.6	20.8
Porters Creek Wicks Rd	CR5PC	Autumn 2010	15/04/10	580	30	970	34	460	1430	76	65.3	41.8	266	7.85	8.9	18.8
Porters Creek Wicks Rd	CR5PC	Spring 2010	30/09/10	3800	30	1140	26	560	1700	117	2.39	65.9	545	7.52	9.4	14.1
Porters Creek Wicks Rd	CR5PC	Spring 2010	04/11/10	7200	50	1330	30	320	1650	77.7	3.42	49.5	301	7.2	9.5	16.8
Porters Creek Wicks Rd	CR5PC	Autumn 2011	04/03/11	390	<10	430	27	240	670	80.1	1.23	38.9	221	7.67	7.5	20.2
Porters Creek Wicks Rd	CR5PC	Autumn 2011	08/04/11	>10000	<10	770	220	610	1380	88.7	1.64	51.3	263	7.11	9.2	18.5

Appendix 3 Rainfall 2004 - 2010









Appendix 4 Macroinvertebrate results

Appendix 5 SIMPER output

SIMPER all five creeks reps merged 2005 – 2011

Data worksheet

Name: All five Cks Sqrt
Data type: Abundance
Sample selection: All
Variable selection: All

Parameters

Resemblance: S17 Bray Curtis similarity
Cut off for low contributions: 90.00%

Factor Groups

Sample Creeks
S5 Archers Ck
S4 Buffalo Ck
S3 Porters Ck
S2 Shrimptons Ck
S1 Terrys Ck

Group Archers Ck

Average similarity: 63.36

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	11.66	10.61	3.88	16.74	16.74
Oligochaeta	6.28	6.05	4.59	9.54	26.28
Physidae	6.45	5.35	3.32	8.44	34.73
Dugesidae	5.82	4.79	2.60	7.55	42.28
Libellulidae	4.62	3.53	2.36	5.58	47.85
Hydrobiidae	4.75	2.98	1.06	4.70	52.56
Hemicorduliidae	3.98	2.96	1.64	4.66	57.22
Coenagrionidae	3.62	2.73	1.86	4.31	61.53
s-f Tanypodinae	3.32	2.72	2.84	4.29	65.83
Notonectidae	3.70	2.40	1.30	3.79	69.62
Stratiomyidae	2.84	2.19	3.45	3.45	73.07
Veliidae	2.96	2.09	1.88	3.30	76.37
Megapodagrionidae	2.46	1.80	1.74	2.84	79.21
s-f Orthoclaeiinae	3.46	1.72	1.01	2.71	81.92
Hydroptilidae	3.40	1.56	0.69	2.46	84.37
Glossiphoniidae	2.33	1.43	1.20	2.26	86.63
Aeshnidae	2.04	1.21	0.85	1.90	88.54
Acarina	1.73	1.04	1.19	1.64	90.17

Group Buffalo Ck

Average similarity: 65.58

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	9.57	10.16	3.78	15.49	15.49
Hydrobiidae	6.42	6.78	2.53	10.33	25.83
Physidae	6.50	6.36	2.21	9.70	35.53
Megapodagrionidae	5.18	5.58	7.16	8.51	44.04
Notonectidae	5.09	4.56	2.92	6.95	50.99
Oligochaeta	4.09	3.93	2.63	5.99	56.98
Dugesidae	3.54	3.41	4.11	5.21	62.19
Planorbidae	3.16	2.80	1.70	4.28	66.46
s-f Tanypodinae	2.65	2.39	1.89	3.64	70.10
Coenagrionidae	3.38	2.35	1.20	3.58	73.68
Hemicorduliidae	3.21	2.10	1.22	3.20	76.88
Isostictidae	2.31	1.97	3.21	3.00	79.88
Hydroptilidae	2.69	1.73	1.11	2.64	82.52
Lymnaeidae	2.06	1.72	1.37	2.62	85.15
Libellulidae	2.36	1.60	1.20	2.44	87.58
Sphaeriidae	2.36	1.57	0.91	2.39	89.98
Stratiomyidae	1.53	1.09	1.22	1.66	91.64

Group Porters Ck

Average similarity: 62.76

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	9.59	10.01	3.34	15.95	15.95
Hydrobiidae	8.52	9.87	7.79	15.73	31.69
Oligochaeta	4.14	4.33	3.52	6.90	38.58
Megapodagrionidae	4.33	4.22	3.01	6.72	45.31
Notonectidae	4.01	3.82	2.00	6.08	51.39
Isostictidae	4.12	3.73	2.66	5.94	57.33
Physidae	4.15	3.67	1.95	5.85	63.18
Coenagrionidae	3.65	3.10	2.14	4.93	68.12
s-f Orthoclaadiinae	3.26	2.52	1.14	4.01	72.13
s-f Tanypodinae	2.52	1.75	1.09	2.80	74.92
Stratiomyidae	1.86	1.60	1.75	2.55	77.48
Hemicorduliidae	2.61	1.50	0.88	2.38	79.86
Dugesiidae	1.81	1.46	1.35	2.33	82.19
Libellulidae	2.13	1.44	1.23	2.29	84.48
Planorbidae	1.89	1.34	0.99	2.14	86.62
Hydroptilidae	2.06	1.31	0.80	2.08	88.70
Glossiphoniidae	1.86	1.06	0.91	1.69	90.39

Group Shrimptons Ck

Average similarity: 61.47

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	7.39	9.90	5.61	16.10	16.10
Dugesiidae	6.95	9.81	3.95	15.95	32.05
Oligochaeta	5.18	7.00	2.47	11.39	43.45
s-f Chironominae	5.69	4.65	1.30	7.56	51.00
Coenagrionidae	3.47	4.06	3.13	6.61	57.61
Glossiphoniidae	3.41	3.77	1.21	6.13	63.74
Acarina	3.17	3.70	2.35	6.02	69.76
Hemicorduliidae	2.69	2.69	1.53	4.37	74.13
Notonectidae	2.58	2.28	1.02	3.72	77.85
Libellulidae	1.68	1.85	1.29	3.00	80.85
Lymnaeidae	1.73	1.72	1.77	2.79	83.65
Megapodagrionidae	2.26	1.64	0.97	2.67	86.32
Hydrobiidae	1.74	1.02	0.80	1.66	87.98
Stratiomyidae	1.07	1.00	1.04	1.62	89.60
Corixidae	1.12	0.84	0.89	1.36	90.96

Group Terrys Ck

Average similarity: 68.99

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	8.26	8.84	4.18	12.81	12.81
Megapodagrionidae	6.99	7.06	2.69	10.23	23.04
s-f Chironominae	7.05	6.48	2.30	9.39	32.43
Physidae	5.49	5.80	5.01	8.40	40.83
Oligochaeta	4.82	5.21	9.58	7.55	48.38
Dugesiidae	4.60	4.71	4.25	6.82	55.20
s-f Tanypodinae	3.86	3.56	2.63	5.16	60.36
Isostictidae	3.27	3.19	3.17	4.62	64.98
Notonectidae	3.53	2.95	1.31	4.27	69.25
Hemicorduliidae	3.32	2.58	1.92	3.74	72.99
Elmidae	1.75	1.68	3.59	2.43	75.41
Planorbidae	1.86	1.64	1.85	2.37	77.79
Sphaeriidae	2.24	1.60	1.02	2.33	80.11
Acarina	1.94	1.42	1.27	2.06	82.18
Coenagrionidae	1.92	1.39	1.23	2.02	84.19
Stratiomyidae	1.49	1.32	1.70	1.91	86.10
Veliidae	1.59	1.28	1.33	1.85	87.96
s-f Orthoclaadiinae	1.61	1.21	1.35	1.76	89.71
Glossiphoniidae	1.46	0.99	0.99	1.43	91.15

Groups Archers Ck & Buffalo Ck

Average dissimilarity = 42.60

Species	Archers Ck Av.Abund	Buffalo Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum. %
s-f Chironominae	1.66	9.57	1.92	1.40	4.52	4.52
Hydroptilidae	3.40	2.69	1.73	1.42	4.06	8.57
Planorbidae	0.15	3.16	1.71	2.16	4.02	12.59
Hydrobiidae	4.75	6.42	1.70	0.99	3.98	16.57
s-f Orthoclaadiinae	3.46	1.87	1.65	1.07	3.87	20.44
Megapodagrionidae	2.46	5.18	1.62	2.26	3.79	24.23
Dugesidae	5.82	3.54	1.60	1.72	3.77	28.00
Libellulidae	4.62	2.36	1.57	1.35	3.69	31.69
Notonectidae	3.70	5.09	1.52	1.40	3.56	35.25
Oligochaeta	6.28	4.09	1.46	1.71	3.43	38.68
Physidae	6.45	6.50	1.45	1.31	3.41	42.09
Hemicorduliidae	3.98	3.21	1.45	1.41	3.41	45.51
Veliidae	2.96	1.07	1.31	1.36	3.09	48.59
Sphaeriidae	0.83	2.36	1.30	1.37	3.05	51.64
Coenagrionidae	3.62	3.38	1.30	1.32	3.05	54.69
Isostictidae	0.00	2.31	1.29	2.17	3.02	57.71
Baetidae	1.85	0.74	1.12	0.88	2.62	60.32
Glossiphoniidae	2.33	0.92	1.10	1.33	2.59	62.92
Aeshnidae	2.04	1.73	1.10	1.28	2.59	65.51
Culicidae	1.87	1.04	1.08	1.17	2.53	68.04
Stratiomyidae	2.84	1.53	0.89	1.19	2.09	70.12
Atyidae	1.50	0.12	0.88	0.77	2.06	72.19
Lymnaeidae	0.97	2.06	0.83	1.77	1.96	74.15
Acarina	1.73	0.76	0.82	1.40	1.92	76.07
s-f Tanypodinae	3.32	2.65	0.79	1.18	1.87	77.93
Corixidae	1.41	0.25	0.75	0.97	1.75	79.68
Corbiculidae	0.27	1.04	0.68	0.69	1.59	81.28
Simuliidae	1.21	0.17	0.66	0.94	1.54	82.82
Ceratopogonidae	1.11	0.57	0.61	1.11	1.43	84.25
Scyphacidae	1.13	0.67	0.60	1.16	1.41	85.65
Tipulidae	1.16	0.25	0.59	1.26	1.40	87.05
Gerridae	0.32	1.05	0.57	0.96	1.35	88.40
Hydraenidae	0.70	0.59	0.53	0.91	1.24	89.64
Dytiscidae	0.81	0.12	0.45	1.02	1.06	90.70

Groups Archers Ck & Porters Ck

Average dissimilarity = 44.63

Species	Archers Ck Av.Abund	Porters Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum. %
Isostictidae	0.00	4.12	2.33	2.50	5.22	5.22
Hydrobiidae	4.75	8.52	2.33	1.14	5.22	10.44
Dugesidae	5.82	1.81	2.33	1.94	5.22	15.66
s-f Chironominae	1.66	9.59	2.23	1.40	5.00	20.66
Physidae	6.45	4.15	1.75	1.54	3.93	24.59
Hydroptilidae	3.40	2.06	1.74	1.47	3.90	28.49
s-f Orthoclaadiinae	3.46	3.26	1.69	1.22	3.79	32.28
Libellulidae	4.62	2.13	1.68	1.43	3.77	36.05
Hemicorduliidae	3.98	2.61	1.53	1.37	3.42	39.47
Oligochaeta	6.28	4.14	1.36	1.70	3.04	42.51
Veliidae	2.96	0.98	1.35	1.38	3.02	45.53
Megapodagrionidae	2.46	4.33	1.33	1.53	2.98	48.51
Notonectidae	3.70	4.01	1.29	1.35	2.88	51.39
Coenagrionidae	3.62	3.65	1.17	1.39	2.63	54.02
Atyidae	1.50	1.56	1.13	1.09	2.53	56.55
s-f Tanypodinae	3.32	2.52	1.11	1.45	2.49	59.05
Aeshnidae	2.04	0.81	1.10	1.22	2.46	61.51
Culicidae	1.87	0.43	1.07	1.03	2.40	63.91
Planorbidae	0.15	1.89	1.06	1.29	2.39	66.30
Glossiphoniidae	2.33	1.86	1.06	1.32	2.39	68.68
Baetidae	1.85	0.25	1.02	0.83	2.29	70.98
Acarina	1.73	1.34	0.87	1.25	1.96	72.94
Stratiomyidae	2.84	1.86	0.82	1.26	1.85	74.78
Corixidae	1.41	0.63	0.81	1.06	1.82	76.60
Sphaeriidae	0.83	1.08	0.78	0.92	1.75	78.35
Ancylidae	0.54	1.18	0.71	1.14	1.58	79.93
Simuliidae	1.21	0.08	0.68	0.94	1.51	81.45
Ceratopogonidae	1.11	0.23	0.62	0.96	1.40	82.85
Tipulidae	1.16	0.73	0.58	1.31	1.30	84.14
Scyphacidae	1.13	0.58	0.57	1.16	1.28	85.42
Dytiscidae	0.81	0.88	0.55	1.18	1.23	86.65
Hydrophilidae	0.34	0.78	0.48	0.85	1.09	87.73
Lymnaeidae	0.97	0.52	0.46	1.25	1.04	88.77
Leptoceridae	0.34	0.76	0.45	0.97	1.02	89.79
Hydraenidae	0.70	0.25	0.42	0.75	0.95	90.74

Groups Buffalo Ck & Porters Ck

Average dissimilarity = 38.50

Species	Buffalo Ck Av.Abund	Porters Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%
Physidae	6.50	4.15	2.03	1.38	5.28	5.28
s-f Chironominae	9.57	9.59	1.84	1.21	4.79	10.07
s-f Orthocladiinae	1.87	3.26	1.62	1.42	4.21	14.28
Hemicorduliidae	3.21	2.61	1.58	1.41	4.10	18.38
Coenagrionidae	3.38	3.65	1.45	1.42	3.78	22.15
Notonectidae	5.09	4.01	1.42	1.53	3.70	25.85
Isostictidae	2.31	4.12	1.38	1.38	3.58	29.42
Hydrobiidae	6.42	8.52	1.36	1.15	3.53	32.95
Hydroptilidae	2.69	2.06	1.34	1.41	3.49	36.45
Sphaeriidae	2.36	1.08	1.24	1.40	3.23	39.67
Planorbidae	3.16	1.89	1.22	1.53	3.16	42.84
Dugesidae	3.54	1.81	1.18	1.48	3.05	45.89
s-f Tanypodinae	2.65	2.52	1.11	1.42	2.89	48.78
Libellulidae	2.36	2.13	1.10	1.34	2.86	51.64
Megapodagrionidae	5.18	4.33	1.08	1.46	2.81	54.45
Glossiphoniidae	0.92	1.86	1.06	1.19	2.75	57.19
Lymnaeidae	2.06	0.52	1.05	1.75	2.72	59.91
Aeshnidae	1.73	0.81	1.04	1.14	2.71	62.62
Oligochaeta	4.09	4.14	0.98	1.09	2.56	65.18
Atyidae	0.12	1.56	0.90	0.89	2.34	67.52
Acarina	0.76	1.34	0.84	1.02	2.18	69.70
Veliidae	1.07	0.98	0.77	1.00	2.01	71.71
Stratiomyidae	1.53	1.86	0.72	1.28	1.88	73.60
Corbiculidae	1.04	0.20	0.71	0.64	1.84	75.44
Culicidae	1.04	0.43	0.65	0.91	1.69	77.13
Ancylidae	0.50	1.18	0.64	1.16	1.67	78.80
Gerridae	1.05	0.33	0.62	1.00	1.62	80.42
Hydrophilidae	0.57	0.78	0.54	1.02	1.41	81.82
Dytiscidae	0.12	0.88	0.54	0.95	1.39	83.22
Baetidae	0.74	0.25	0.49	0.59	1.28	84.50
Scyphacidae	0.67	0.58	0.47	0.93	1.21	85.71
Tipulidae	0.25	0.73	0.46	1.03	1.19	86.90
Corixidae	0.25	0.63	0.45	0.73	1.17	88.07
Leptoceridae	0.08	0.76	0.44	0.87	1.15	89.23
Hydraenidae	0.59	0.25	0.43	0.77	1.12	90.34

Groups Archers Ck & Shrimptons Ck

Average dissimilarity = 47.03

Species	Archers Ck Av.Abund	Shrimptons Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%
s-f Chironominae	1.66	5.69	4.41	1.46	9.37	9.37
Hydrobiidae	4.75	1.74	2.44	1.59	5.19	14.56
Hydroptilidae	3.40	0.73	2.00	1.18	4.25	18.81
Libellulidae	4.62	1.68	1.89	1.55	4.03	22.84
s-f Orthocladiinae	3.46	0.82	1.87	1.01	3.97	26.81
Veliidae	2.96	0.65	1.64	1.46	3.49	30.30
Notonectidae	3.70	2.58	1.54	1.36	3.28	33.58
Physidae	6.45	7.39	1.50	1.28	3.20	36.77
s-f Tanypodinae	3.32	1.10	1.48	1.59	3.15	39.92
Hemicorduliidae	3.98	2.69	1.48	1.42	3.14	43.06
Glossiphoniidae	2.33	3.41	1.48	1.43	3.14	46.20
Dugesidae	5.82	6.95	1.40	1.11	2.98	49.18
Aeshnidae	2.04	0.52	1.24	1.20	2.63	51.81
Megapodagrionidae	2.46	2.26	1.23	1.18	2.61	54.41
Culicidae	1.87	0.32	1.21	0.97	2.57	56.98
Acarina	1.73	3.17	1.19	1.38	2.54	59.52
Coenagrionidae	3.62	3.47	1.16	1.37	2.47	62.00
Stratiomyidae	2.84	1.07	1.13	1.41	2.41	64.41
Baetidae	1.85	0.19	1.11	0.80	2.37	66.77
Oligochaeta	6.28	5.18	1.02	1.17	2.17	68.95
Atyidae	1.50	0.08	0.99	0.77	2.10	71.05
Planorbidae	0.15	1.44	0.93	0.81	1.98	73.03
Corixidae	1.41	1.12	0.86	1.18	1.83	74.86
Corbiculidae	0.27	1.05	0.84	0.65	1.79	76.65
Isostictidae	0.00	1.37	0.80	0.89	1.71	78.36
Simuliidae	1.21	0.00	0.75	0.93	1.60	79.96
Sphaeriidae	0.83	0.78	0.72	0.83	1.53	81.49
Tipulidae	1.16	0.08	0.70	1.25	1.49	82.98
Lymnaeidae	0.97	1.73	0.69	1.13	1.47	84.45
Ceratopogonidae	1.11	0.15	0.67	0.93	1.43	85.88
Scyphacidae	1.13	0.45	0.66	1.12	1.39	87.27
Ancylidae	0.54	0.51	0.52	0.70	1.10	88.37
Parastacidae	0.00	0.82	0.50	1.10	1.07	89.45
Dytiscidae	0.81	0.08	0.50	1.01	1.06	90.51

Groups Buffalo Ck & Shrimptons Ck

Average dissimilarity = 46.62

Species	Buffalo Ck		Shrimptons Ck		Diss/SD	Contrib%	Cum. %
	Av.Abund	Av.Abund	Av.Diss	Av.Diss			
s-f Chironominae	9.57	5.69	3.71	1.37	1.37	7.96	7.96
Hydrobiidae	6.42	1.74	3.46	1.87	1.87	7.42	15.38
Dugesiiidae	3.54	6.95	2.46	1.71	1.71	5.27	20.66
Megapodagrionidae	5.18	2.26	2.24	1.63	1.63	4.80	25.46
Glossiphoniidae	0.92	3.41	2.07	1.44	1.44	4.43	29.89
Notonectidae	5.09	2.58	2.05	1.45	1.45	4.41	34.30
Acarina	0.76	3.17	1.73	1.78	1.78	3.71	38.01
Physidae	6.50	7.39	1.68	1.48	1.48	3.61	41.61
Planorbidae	3.16	1.44	1.67	1.70	1.70	3.59	45.20
Hydroptilidae	2.69	0.73	1.67	1.29	1.29	3.57	48.77
Hemicorduliidae	3.21	2.69	1.55	1.39	1.39	3.32	52.09
Coenagrionidae	3.38	3.47	1.47	1.39	1.39	3.16	55.25
Oligochaeta	4.09	5.18	1.45	1.47	1.47	3.10	58.35
Sphaeriidae	2.36	0.78	1.43	1.32	1.32	3.06	61.41
s-f Tanypodinae	2.65	1.10	1.25	1.62	1.62	2.68	64.10
Isostictidae	2.31	1.37	1.22	1.43	1.43	2.61	66.70
Corbiculidae	1.04	1.05	1.17	0.75	0.75	2.51	69.21
Aeshnidae	1.73	0.52	1.11	1.11	1.11	2.39	71.60
Libellulidae	2.36	1.68	1.07	1.30	1.30	2.29	73.89
s-f Orthoclaadiinae	1.87	0.82	1.04	0.95	0.95	2.23	76.11
Lymnaeidae	2.06	1.73	0.90	1.56	1.56	1.94	78.05
Veliidae	1.07	0.65	0.79	0.91	0.91	1.70	79.75
Culicidae	1.04	0.32	0.79	0.90	0.90	1.70	81.45
Stratiomyidae	1.53	1.07	0.71	1.28	1.28	1.52	82.97
Gerridae	1.05	0.40	0.70	0.99	0.99	1.50	84.46
Corixidae	0.25	1.12	0.69	1.16	1.16	1.47	85.93
Parastacidae	0.00	0.82	0.55	1.10	1.10	1.18	87.11
Ancylidae	0.50	0.51	0.52	0.96	0.96	1.12	88.23
Scyphacidae	0.67	0.45	0.51	0.90	0.90	1.10	89.33
Baetidae	0.74	0.19	0.51	0.54	0.54	1.10	90.43

Groups Porters Ck & Shrimptons Ck

Average dissimilarity = 51.17

Species	Porters Ck		Shrimptons Ck		Diss/SD	Contrib%	Cum. %
	Av.Abund	Av.Abund	Av.Diss	Av.Diss			
Hydrobiidae	8.52	1.74	4.84	2.74	2.74	9.47	9.47
s-f Chironominae	9.59	5.69	3.91	1.31	1.31	7.64	17.11
Dugesiiidae	1.81	6.95	3.69	2.56	2.56	7.22	24.33
Physidae	4.15	7.39	2.45	1.63	1.63	4.78	29.12
Isostictidae	4.12	1.37	2.19	1.65	1.65	4.27	33.39
s-f Orthoclaadiinae	3.26	0.82	1.96	1.48	1.48	3.82	37.21
Megapodagrionidae	4.33	2.26	1.91	1.39	1.39	3.73	40.94
Glossiphoniidae	1.86	3.41	1.79	1.35	1.35	3.51	44.45
Acarina	1.34	3.17	1.66	1.69	1.69	3.25	47.70
Notonectidae	4.01	2.58	1.63	1.31	1.31	3.18	50.87
Hemicorduliidae	2.61	2.69	1.54	1.51	1.51	3.01	53.88
s-f Tanypodinae	2.52	1.10	1.39	1.20	1.20	2.71	56.59
Hydroptilidae	2.06	0.73	1.35	1.29	1.29	2.65	59.24
Planorbidae	1.89	1.44	1.33	1.32	1.32	2.60	61.84
Oligochaeta	4.14	5.18	1.33	1.52	1.52	2.60	64.44
Coenagrionidae	3.65	3.47	1.32	1.49	1.49	2.59	67.03
Atyidae	1.56	0.08	1.01	0.89	0.89	1.97	69.00
Libellulidae	2.13	1.68	0.99	1.34	1.34	1.94	70.94
Lymnaeidae	0.52	1.73	0.93	1.32	1.32	1.82	72.76
Corbiculidae	0.20	1.05	0.91	0.60	0.60	1.77	74.53
Ancylidae	1.18	0.51	0.89	1.14	1.14	1.73	76.26
Sphaeriidae	1.08	0.78	0.83	1.04	1.04	1.61	77.88
Stratiomyidae	1.86	1.07	0.81	1.19	1.19	1.58	79.46
Corixidae	0.63	1.12	0.79	1.19	1.19	1.54	81.00
Veliidae	0.98	0.65	0.78	0.94	0.94	1.53	82.53
Aeshnidae	0.81	0.52	0.69	0.85	0.85	1.35	83.88
Dytiscidae	0.88	0.08	0.60	0.93	0.93	1.17	85.05
Parastacidae	0.00	0.82	0.56	1.11	1.11	1.09	86.14
Hydrophilidae	0.78	0.23	0.55	0.83	0.83	1.07	87.21
Tipulidae	0.73	0.08	0.52	0.96	0.96	1.02	88.23
Leptoceridae	0.76	0.00	0.50	0.86	0.86	0.97	89.21
Scyphacidae	0.58	0.45	0.46	1.03	1.03	0.90	90.10

Groups Archers Ck & Terrys Ck

Average dissimilarity = 43.90

Species	Archers Ck Av.Abund	Terrys Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%
s-f Chironominae	11.66	7.05	2.93	1.47	6.68	6.68
Megapodagrionidae	2.46	6.99	2.63	2.47	5.99	12.67
Hydrobiidae	4.75	8.26	2.33	1.16	5.30	17.97
Isostictidae	0.00	3.27	1.83	3.33	4.18	22.15
Libellulidae	4.62	1.67	1.82	1.55	4.14	26.29
Hydroptilidae	3.40	1.00	1.73	1.30	3.93	30.22
s-f Orthoclaadiinae	3.46	1.61	1.52	1.02	3.47	33.69
Notonectidae	3.70	3.53	1.32	1.32	3.01	36.70
Hemicorduliidae	3.98	3.32	1.31	1.40	2.98	39.68
Dugesiiidae	5.82	4.60	1.29	1.69	2.94	42.62
Coenagrionidae	3.62	1.92	1.27	1.28	2.89	45.50
Sphaeriidae	0.83	2.24	1.24	1.46	2.82	48.32
Physidae	6.45	5.49	1.22	1.49	2.79	51.11
Culicidae	1.87	0.73	1.08	1.08	2.46	53.57
Atyidae	1.50	0.76	1.08	0.84	2.45	56.02
Aeshnidae	2.04	0.64	1.02	1.20	2.32	58.34
Veliidae	2.96	1.59	1.01	1.15	2.29	60.63
Oligochaeta	6.28	4.82	1.00	1.52	2.27	62.90
Baetidae	1.85	0.12	0.99	0.78	2.27	65.17
Planorbidae	0.15	1.86	0.98	1.99	2.23	67.40
Glossiphoniidae	2.33	1.46	0.93	1.30	2.13	69.53
s-f Tanypodinae	3.32	3.86	0.92	1.45	2.09	71.62
Stratiomyidae	2.84	1.49	0.83	1.23	1.88	73.50
Elmidae	0.34	1.75	0.82	1.81	1.88	75.38
Corixidae	1.41	0.19	0.78	0.98	1.78	77.15
Acarina	1.73	1.94	0.77	1.34	1.76	78.91
Simuliidae	1.21	0.87	0.69	1.13	1.57	80.48
Gerridae	0.32	1.34	0.67	1.39	1.54	82.02
Corbiculidae	0.27	1.01	0.64	0.70	1.46	83.48
Ceratopogonidae	1.11	0.50	0.64	1.03	1.45	84.93
Scyphacidae	1.13	0.25	0.60	1.12	1.37	86.30
Tipulidae	1.16	0.77	0.54	1.33	1.23	87.53
Gelastocoridae	0.19	0.88	0.47	1.41	1.08	88.61
Dytiscidae	0.81	0.17	0.44	1.06	1.01	89.62
Lymnaeidae	0.97	0.89	0.44	1.05	0.99	90.61

Groups Buffalo Ck & Terrys Ck

Average dissimilarity = 36.54

Species	Buffalo Ck Av.Abund	Terrys Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%
s-f Chironominae	9.57	7.05	2.19	1.30	6.00	6.00
Megapodagrionidae	5.18	6.99	1.59	1.72	4.36	10.36
Notonectidae	5.09	3.53	1.56	1.53	4.28	14.64
Hydrobiidae	6.42	8.26	1.51	1.31	4.12	18.75
Physidae	6.50	5.49	1.47	1.51	4.01	22.77
Hemicorduliidae	3.21	3.32	1.46	1.37	4.00	26.77
Coenagrionidae	3.38	1.92	1.43	1.47	3.90	30.67
Hydroptilidae	2.69	1.00	1.27	1.19	3.48	34.15
Sphaeriidae	2.36	2.24	1.15	1.40	3.15	37.31
s-f Tanypodinae	2.65	3.86	1.10	1.32	3.01	40.32
Planorbidae	3.16	1.86	1.09	1.68	2.98	43.29
Libellulidae	2.36	1.67	1.09	1.31	2.98	46.27
Elmidae	0.08	1.75	1.02	2.53	2.78	49.05
Dugesiiidae	3.54	4.60	0.98	1.32	2.69	51.74
Acarina	0.76	1.94	0.96	1.42	2.64	54.38
Aeshnidae	1.73	0.64	0.95	1.20	2.61	56.99
Oligochaeta	4.09	4.82	0.95	1.19	2.60	59.58
Isostictidae	2.31	3.27	0.94	1.53	2.57	62.15
Corbiculidae	1.04	1.01	0.94	0.77	2.57	64.72
Lymnaeidae	2.06	0.89	0.93	1.67	2.55	67.27
s-f Orthoclaadiinae	1.87	1.61	0.93	1.06	2.55	69.82
Glossiphoniidae	0.92	1.46	0.84	1.21	2.30	72.11
Veliidae	1.07	1.59	0.82	1.37	2.24	74.36
Culicidae	1.04	0.73	0.76	1.01	2.09	76.45
Gerridae	1.05	1.34	0.75	1.29	2.05	78.50
Stratiomyidae	1.53	1.49	0.58	1.30	1.59	80.09
Atyidae	0.12	0.76	0.53	0.43	1.46	81.55
Simuliidae	0.17	0.87	0.51	1.04	1.40	82.95
Gelastocoridae	0.43	0.88	0.48	1.32	1.32	84.27
Tipulidae	0.25	0.77	0.46	1.10	1.25	85.52
Ceratopogonidae	0.57	0.50	0.44	0.99	1.21	86.73
Baetidae	0.74	0.12	0.44	0.51	1.19	87.92
Scyphacidae	0.67	0.25	0.42	0.81	1.15	89.08
Talitridae	0.25	0.66	0.41	1.05	1.11	90.19

Groups Porters Ck & Terrys Ck

Average dissimilarity = 38.88

Species	Porters Ck Av.Abund	Terrys Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%
s-f Chironominae	9.59	7.05	2.39	1.18	6.14	6.14
Megapodagrionidae	4.33	6.99	2.04	1.77	5.25	11.39
Dugesiiidae	1.81	4.60	1.76	1.80	4.53	15.92
Hemicorduliidae	2.61	3.32	1.50	1.39	3.85	19.77
s-f Orthocladiinae	3.26	1.61	1.49	1.48	3.83	23.61
Physidae	4.15	5.49	1.41	1.45	3.62	27.23
s-f Tanypodinae	2.52	3.86	1.37	1.43	3.53	30.76
Coenagrionidae	3.65	1.92	1.36	1.37	3.49	34.25
Notonectidae	4.01	3.53	1.26	1.32	3.23	37.48
Sphaeriidae	1.08	2.24	1.15	1.45	2.96	40.44
Atyidae	1.56	0.76	1.14	0.91	2.93	43.38
Hydrobiidae	8.52	8.26	1.10	1.43	2.82	46.20
Isostictidae	4.12	3.27	1.09	1.34	2.81	49.01
Elmidae	0.00	1.75	1.08	3.00	2.78	51.79
Hydroptilidae	2.06	1.00	1.08	1.45	2.76	54.56
Acarina	1.34	1.94	1.02	1.37	2.62	57.18
Libellulidae	2.13	1.67	1.02	1.28	2.62	59.80
Glossiphoniidae	1.86	1.46	0.97	1.24	2.49	62.29
Oligochaeta	4.14	4.82	0.86	1.38	2.21	64.50
Planorbidae	1.89	1.86	0.82	1.31	2.12	66.62
Veliidae	0.98	1.59	0.81	1.38	2.09	68.71
Gerridae	0.33	1.34	0.73	1.38	1.87	70.57
Ancyliidae	1.18	0.35	0.72	1.14	1.85	72.42
Corbiculidae	0.20	1.01	0.67	0.64	1.71	74.14
Stratiomyidae	1.86	1.49	0.60	1.16	1.54	75.68
Aeshnidae	0.81	0.64	0.58	0.96	1.49	77.16
Culicidae	0.43	0.73	0.56	0.83	1.43	78.59
Dytiscidae	0.88	0.17	0.53	0.99	1.37	79.96
Simuliidae	0.08	0.87	0.52	1.00	1.34	81.29
Gelastocoridae	0.20	0.88	0.51	1.41	1.32	82.61
Tipulidae	0.73	0.77	0.50	1.23	1.30	83.91
Lymnaeidae	0.52	0.89	0.50	1.12	1.28	85.19
Hydrophilidae	0.78	0.25	0.49	0.85	1.26	86.44
Corixidae	0.63	0.19	0.46	0.68	1.18	87.62
Leptoceridae	0.76	0.00	0.44	0.85	1.14	88.76
Talitridae	0.37	0.66	0.42	1.08	1.08	89.84
Ceratopogonidae	0.23	0.50	0.38	0.73	0.99	90.83

Groups Shrimptons Ck & Terrys Ck

Average dissimilarity = 46.02

Species	Shrimptons Ck Av.Abund	Terrys Ck Av.Abund	Av.Diss	Diss/SD	Contrib%	Cum.%
Hydrobiidae	1.74	8.26	4.62	2.30	10.05	10.05
Megapodagrionidae	2.26	6.99	3.47	1.95	7.54	17.59
s-f Chironominae	5.69	7.05	2.97	1.39	6.45	24.04
s-f Tanypodinae	1.10	3.86	1.95	1.72	4.24	28.28
Dugesiiidae	6.95	4.60	1.78	1.52	3.88	32.16
Glossiphoniidae	3.41	1.46	1.76	1.46	3.82	35.98
Isostictidae	1.37	3.27	1.66	1.81	3.61	39.59
Notonectidae	2.58	3.53	1.60	1.31	3.47	43.06
Physidae	7.39	5.49	1.57	1.49	3.40	46.46
Hemicorduliidae	2.69	3.32	1.41	1.32	3.05	49.51
Coenagrionidae	3.47	1.92	1.33	1.37	2.90	52.41
Sphaeriidae	0.78	2.24	1.33	1.37	2.88	55.29
Acarina	3.17	1.94	1.22	1.32	2.64	57.93
Planorbidae	1.44	1.86	1.18	1.58	2.57	60.50
Elmidae	0.08	1.75	1.15	2.55	2.51	63.01
Corbiculidae	1.05	1.01	1.14	0.75	2.48	65.49
Oligochaeta	5.18	4.82	1.02	1.38	2.22	67.71
Veliidae	0.65	1.59	0.99	1.56	2.15	69.86
Libellulidae	1.68	1.67	0.91	1.22	1.97	71.84
Lymnaeidae	1.73	0.89	0.81	1.19	1.77	73.60
Gerridae	0.40	1.34	0.80	1.37	1.73	75.34
Hydroptilidae	0.73	1.00	0.79	1.28	1.72	77.06
s-f Orthocladiinae	0.82	1.61	0.77	1.23	1.66	78.72
Corixidae	1.12	0.19	0.76	1.25	1.64	80.37
Culicidae	0.32	0.73	0.61	0.71	1.33	81.70
Stratiomyidae	1.07	1.49	0.60	1.20	1.29	83.00
Atyidae	0.08	0.76	0.59	0.42	1.28	84.28
Simuliidae	0.00	0.87	0.57	0.96	1.24	85.52
Parastacidae	0.82	0.00	0.55	1.11	1.19	86.71
Tipulidae	0.08	0.77	0.54	1.09	1.18	87.89
Aeshnidae	0.52	0.64	0.53	1.07	1.14	89.04
Gelastocoridae	0.61	0.88	0.51	1.27	1.12	90.16

SIMPER Archers Creek 2005 – 2011

Data worksheet

Name: Archers Sqrt
 Data type: Abundance
 Sample selection: All
 Variable selection: All

Parameters

Resemblance: S17 Bray Curtis similarity
 Cut off for low contributions: 90.00%

Factor Groups

Sample	Season Year
S5	Autumn 2005
S5	Spring 2005
S5	Autumn 2006
S5	Spring 2006
S5	Autumn 2007
S5	Spring 2007
S5	Autumn 2008
S5	Spring 2008
S5	Autumn 2009
S5	Spring 2009
S5	Autumn 2010
S5	Spring 2010
S5	Autumn 2011

Group Autumn 2005

Average similarity: 68.02

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Megapodagrionidae	3.60	7.56	2.16	11.11	11.11
Atyidae	3.30	7.18	8.38	10.56	21.67
Oligochaeta	3.29	6.80	3.09	9.99	31.67
s-f Chironominae	3.31	6.58	5.68	9.67	41.33
Libellulidae	2.52	5.47	4.54	8.04	49.37
Dugesiiidae	2.75	5.32	5.50	7.82	57.19
Coenagrionidae	3.19	5.20	4.94	7.65	64.83
Veliidae	2.14	5.09	3.65	7.49	72.32
Hemicorduliidae	2.66	4.82	8.37	7.08	79.40
Physidae	1.67	3.65	1.80	5.36	84.77
Stratiomyidae	1.62	2.98	7.13	4.38	89.15
s-f Tanypodinae	1.00	2.65	8.58	3.90	93.04

Group Spring 2005

Average similarity: 58.85

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	8.06	19.91	6.33	33.83	33.83
Oligochaeta	4.19	10.61	6.70	18.04	51.87
Physidae	2.95	7.20	6.86	12.24	64.11
Coenagrionidae	3.08	6.42	6.60	10.90	75.01
Libellulidae	2.87	6.04	1.03	10.27	85.28
Aeshnidae	1.49	2.00	0.58	3.40	88.68
Corbiculidae	1.15	1.97	0.58	3.36	92.04

Group Autumn 2006

Average similarity: 72.08

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	9.12	23.57	21.16	32.71	32.71
Oligochaeta	3.39	8.58	10.99	11.90	44.61
Glossiphoniidae	2.10	4.95	2.66	6.86	51.47
Megapodagrionidae	2.10	4.67	4.46	6.48	57.95
Libellulidae	2.02	4.51	4.25	6.25	64.20
Coenagrionidae	2.03	4.06	1.97	5.63	69.83
Hemicorduliidae	1.96	3.87	2.23	5.38	75.21
Dugesiiidae	1.67	3.67	2.67	5.09	80.29
Veliidae	1.28	3.19	4.05	4.43	84.72
Notonectidae	1.47	3.19	4.25	4.42	89.14
Aeshnidae	2.05	2.48	0.58	3.44	92.58

Group Spring 2006

Average similarity: 60.22

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.35	14.93	4.12	24.80	24.80
Physidae	2.81	10.04	3.55	16.68	41.47
DugesIIDae	2.63	8.66	2.75	14.39	55.86
Oligochaeta	2.43	7.87	2.82	13.07	68.93
Hydrobiidae	1.81	4.51	1.47	7.48	76.41
s-f Tanypodinae	1.07	3.38	1.76	5.62	82.03
Veliidae	0.80	1.96	0.79	3.25	85.28
s-f Orthocladiinae	1.06	1.92	0.79	3.18	88.47
Stratiomyidae	0.87	1.55	0.57	2.58	91.05

Group Autumn 2007

Average similarity: 57.33

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	5.02	17.66	4.21	30.81	30.81
Oligochaeta	2.53	6.59	1.30	11.49	42.30
Physidae	2.35	6.46	3.17	11.27	53.57
DugesIIDae	2.20	4.54	1.22	7.93	61.50
s-f Tanypodinae	1.19	4.00	5.04	6.99	68.48
Libellulidae	1.50	3.89	1.21	6.79	75.27
Veliidae	1.93	3.06	0.75	5.35	80.62
Glossiphoniidae	1.08	2.51	1.28	4.38	85.00
Megapodagrionidae	1.01	1.90	0.77	3.31	88.31
Aeshnidae	0.98	1.76	0.78	3.07	91.38

Group Spring 2007

Average similarity: 60.31

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	5.57	14.73	7.51	24.42	24.42
Physidae	3.77	10.06	6.91	16.67	41.10
Hydrobiidae	2.81	6.59	2.44	10.92	52.02
Oligochaeta	2.95	6.37	3.38	10.56	62.58
DugesIIDae	2.70	4.94	1.22	8.20	70.78
s-f Tanypodinae	1.84	3.90	2.50	6.46	77.24
Sphaeriidae	2.01	3.07	1.01	5.08	82.32
Hemicorduliidae	1.57	2.34	1.14	3.88	86.20
Libellulidae	1.04	2.10	1.29	3.49	89.68
s-f Orthocladiinae	1.33	1.74	0.74	2.88	92.57

Group Autumn 2008

Average similarity: 61.14

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Orthocladiinae	4.42	13.68	5.42	22.37	22.37
s-f Chironominae	3.35	10.70	5.74	17.51	39.88
Hydrobiidae	2.52	6.99	2.75	11.43	51.31
Oligochaeta	1.69	5.02	3.59	8.21	59.52
Veliidae	1.62	4.29	4.33	7.02	66.54
Physidae	2.03	4.28	1.16	7.00	73.54
Notonectidae	1.69	3.38	1.06	5.54	79.07
Hydroptilidae	1.35	2.18	0.74	3.57	82.65
Ceratopogonidae	1.15	2.05	0.78	3.35	86.00
Baetidae	1.26	1.64	0.48	2.68	88.68
Stratiomyidae	0.67	1.42	0.79	2.32	91.00

Group Spring 2008

Average similarity: 69.72

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	3.99	12.17	10.20	17.46	17.46
DugesIIDae	2.87	8.16	7.17	11.70	29.16
Oligochaeta	2.85	8.05	6.36	11.55	40.71
Hydroptilidae	3.13	7.69	3.78	11.04	51.75
Physidae	2.77	6.64	2.38	9.53	61.28
Hydrobiidae	2.46	6.11	5.80	8.77	70.04
s-f Orthocladiinae	2.49	4.74	1.09	6.80	76.84
Notonectidae	1.74	3.20	1.03	4.58	81.43
Ancylidae	1.30	2.59	1.31	3.72	85.15
s-f Tanypodinae	1.24	2.29	1.29	3.29	88.43
Glossiphoniidae	1.08	2.28	1.24	3.27	91.70

Group Autumn 2009

Average similarity: 64.32

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.02	10.17	4.55	15.81	15.81
Libellulidae	3.11	6.61	2.09	10.27	26.08
Dugesiidae	2.69	6.32	2.12	9.83	35.91
Hydroptilidae	2.52	5.85	2.36	9.10	45.00
Oligochaeta	2.33	5.20	3.07	8.08	53.08
s-f Orthoclaadiinae	2.16	4.81	2.62	7.49	60.57
Physidae	2.02	4.29	2.49	6.67	67.24
Simuliidae	1.57	3.61	3.93	5.61	72.85
Coenagrionidae	1.77	3.55	3.75	5.52	78.37
Hydrobiidae	2.04	3.06	1.17	4.76	83.13
Notonectidae	1.78	2.08	0.73	3.23	86.36
Ceratopogonidae	0.97	1.85	1.34	2.87	89.23
Glossiphoniidae	1.22	1.55	0.77	2.41	91.64

Group Spring 2009

Average similarity: 65.01

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.15	11.59	8.44	17.82	17.82
Dugesiidae	3.07	8.94	5.85	13.76	31.58
Physidae	3.07	8.84	5.79	13.60	45.18
Hydrobiidae	3.00	8.47	4.25	13.03	58.22
Oligochaeta	2.40	6.38	3.39	9.81	68.02
Hydroptilidae	2.39	5.15	2.26	7.93	75.95
s-f Tanypodinae	1.98	5.00	3.02	7.68	83.63
Culicidae	1.44	2.40	1.23	3.69	87.33
Stratiomyidae	1.02	2.19	1.33	3.37	90.69

Group Autumn 2010

Average similarity: 65.73

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.77	11.51	6.78	17.51	17.51
Baetidae	2.86	6.16	3.08	9.37	26.88
Hydrobiidae	2.59	5.31	4.67	8.08	34.96
Physidae	2.41	4.83	3.53	7.35	42.31
Notonectidae	2.26	4.75	4.82	7.22	49.54
Hemicorduliidae	1.84	4.05	6.00	6.17	55.71
Oligochaeta	1.91	4.04	7.75	6.14	61.85
Libellulidae	2.62	3.67	1.15	5.59	67.43
Dugesiidae	2.11	3.54	1.34	5.39	72.82
Aeshnidae	1.48	3.24	4.79	4.93	77.75
Hydroptilidae	1.91	3.19	1.25	4.85	82.60
Corixidae	1.60	2.47	1.14	3.75	86.36
Velliidae	1.93	2.31	0.78	3.52	89.87
s-f Orthoclaadiinae	1.35	1.60	0.69	2.44	92.31

Group Spring 2010

Average similarity: 63.78

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	5.75	13.41	6.83	21.03	21.03
Physidae	3.75	9.75	7.66	15.28	36.31
Notonectidae	2.97	7.16	4.95	11.22	47.53
Hydrobiidae	3.19	7.00	2.73	10.98	58.51
Oligochaeta	2.38	4.92	2.16	7.71	66.22
Dugesiidae	2.55	4.84	4.06	7.59	73.81
s-f Tanypodinae	1.94	3.18	1.32	4.98	78.80
Hydroptilidae	2.24	2.92	0.97	4.58	83.38
Libellulidae	1.83	2.76	1.17	4.32	87.70
Coenagrionidae	1.52	2.11	1.16	3.30	91.01

Group Autumn 2011

Average similarity: 52.28

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.36	8.97	3.04	17.16	17.16
Physidae	3.07	6.55	3.87	12.53	29.69
Oligochaeta	2.21	4.63	3.36	8.86	38.55
Dugesiiidae	2.40	4.45	1.27	8.52	47.06
Hemicorduliidae	2.81	3.99	1.25	7.62	54.68
Libellulidae	2.50	3.64	1.21	6.96	61.65
Glossiphoniidae	2.08	3.04	1.20	5.81	67.46
Aeshnidae	1.26	2.87	7.19	5.49	72.95
Coenagrionidae	2.13	2.81	1.16	5.37	78.32
Stratiomyidae	2.11	2.43	0.70	4.65	82.97
Hydrobiidae	1.52	2.10	0.65	4.02	86.99
Velliidae	1.37	1.62	0.77	3.10	90.09

SIMPER Shrimptons Creek 2005 – 2011

Data worksheet

Name: Shrimpton Ck Sqrt
 Data type: Abundance
 Sample selection: All
 Variable selection: All

Parameters

Resemblance: S17 Bray Curtis similarity
 Cut off for low contributions: 90.00%

Factor Groups

Sample	Season Year
S2	Autumn 2005
S2	Spring 2005
S2	Autumn 2006
S2	Spring 2006
S2	Autumn 2007
S2	Spring 2007
S2	Autumn 2008
S2	Spring 2008
S2	Autumn 2009
S2	Spring 2009
S2	Autumn 2010
S2	Spring 2010
S2	Autumn 2011

Group Autumn 2005

Average similarity: 75.89

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	3.90	16.31	7.41	21.49	21.49
DugesIIDae	3.81	15.30	9.53	20.16	41.65
Oligochaeta	3.43	13.48	44.44	17.77	59.41
Glossiphoniidae	3.04	10.94	8.30	14.42	73.83
Corbiculidae	2.63	9.41	3.56	12.40	86.23
Planorbidae	2.39	7.68	3.56	10.12	96.35

Group Spring 2005

Average similarity: 76.54

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	4.03	13.28	19.85	17.35	17.35
Oligochaeta	3.91	13.08	46.28	17.09	34.44
DugesIIDae	3.46	11.45	11.43	14.97	49.41
Glossiphoniidae	3.04	9.70	10.63	12.67	62.08
s-f Chironominae	3.09	8.94	4.43	11.68	73.76
Planorbidae	2.88	8.57	3.06	11.20	84.96
Corbiculidae	2.64	7.51	12.72	9.82	94.78

Group Autumn 2006

Average similarity: 76.70

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Oligochaeta	3.68	16.90	13.74	22.03	22.03
DugesIIDae	2.82	13.43	9.18	17.51	39.55
Physidae	2.96	13.00	3.19	16.95	56.50
Acarina	2.08	9.91	14.34	12.92	69.42
Corbiculidae	2.39	9.70	6.21	12.64	82.06
Hemicorduliidae	1.88	6.51	2.65	8.49	90.55

Group Spring 2006

Average similarity: 62.17

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.59	20.77	7.14	33.41	33.41
Physidae	3.41	15.57	10.74	25.04	58.46
Oligochaeta	2.05	7.05	1.41	11.35	69.80
DugesIIDae	1.31	3.75	1.10	6.03	75.83
Notonectidae	1.03	3.23	1.14	5.19	81.03
Acarina	1.12	3.02	1.10	4.86	85.89
Hemicorduliidae	1.12	2.85	0.79	4.58	90.47

Group Autumn 2007

Average similarity: 60.39

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	2.95	8.39	2.58	13.89	13.89
Megapodagrionidae	2.10	6.97	5.43	11.55	25.44
DugesIIDae	2.16	6.71	3.12	11.10	36.54
Acarina	2.02	5.61	3.42	9.28	45.83
Coenagrionidae	1.80	5.41	2.78	8.96	54.79
Isostictidae	1.72	5.19	3.30	8.59	63.38
Hemicorduliidae	2.14	4.74	1.11	7.85	71.23
Oligochaeta	1.72	4.72	1.08	7.81	79.04
Physidae	2.28	4.63	1.08	7.67	86.71
Notonectidae	1.01	2.01	0.75	3.33	90.04

Group Spring 2007

Average similarity: 63.13

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.53	12.18	4.26	19.29	19.29
Physidae	3.79	10.55	5.00	16.72	36.01
Oligochaeta	2.22	6.38	4.93	10.10	46.12
DugesIIDae	2.25	5.37	2.58	8.51	54.62
Coenagrionidae	2.01	4.99	3.48	7.90	62.53
Isostictidae	1.88	4.87	3.27	7.71	70.23
Megapodagrionidae	1.95	3.26	0.78	5.17	75.41
Ancylidae	1.37	3.05	1.34	4.83	80.24
Corixidae	1.28	2.94	1.28	4.65	84.89
Hemicorduliidae	1.25	2.90	1.35	4.59	89.48
Notonectidae	0.67	1.56	0.78	2.48	91.96

Group Autumn 2008

Average similarity: 57.63

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
DugesIIDae	3.55	20.83	4.34	36.15	36.15
Physidae	2.91	16.00	4.67	27.76	63.91
Oligochaeta	1.52	6.57	1.29	11.39	75.30
Megapodagrionidae	1.05	3.47	0.77	6.02	81.32
Glossiphoniidae	1.22	2.81	0.76	4.87	86.19
Acarina	0.98	2.63	0.78	4.57	90.76

Group Spring 2008

Average similarity: 62.97

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	3.46	15.55	5.33	24.69	24.69
DugesIIDae	2.86	12.16	5.11	19.31	44.00
s-f Chironominae	2.37	9.80	3.96	15.56	59.56
Oligochaeta	2.02	7.51	2.08	11.93	71.48
Coenagrionidae	1.95	6.57	2.85	10.43	81.91
Acarina	1.41	2.94	0.78	4.66	86.58
Glossiphoniidae	0.98	2.04	0.77	3.24	89.82
Sphaeriidae	0.79	1.84	0.78	2.92	92.73

Group Autumn 2009

Average similarity: 48.10

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
DugesIIDae	2.87	15.22	5.70	31.64	31.64
Glossiphoniidae	1.62	7.41	1.34	15.41	47.05
Notonectidae	1.50	5.22	1.18	10.86	57.91
Lymnaeidae	1.09	4.86	1.30	10.10	68.01
Physidae	1.45	4.82	0.72	10.02	78.03
Coenagrionidae	1.16	3.69	0.76	7.68	85.71
s-f Chironominae	1.07	3.50	0.69	7.29	92.99

Group Spring 2009

Average similarity: 61.92

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	3.60	11.95	3.70	19.30	19.30
DugesIIDae	3.09	10.92	5.93	17.63	36.93
s-f Chironominae	3.18	10.78	6.17	17.41	54.34
Glossiphoniidae	2.04	4.82	1.22	7.79	62.13
Hemicorduliidae	1.51	4.32	3.66	6.98	69.10
Oligochaeta	1.68	4.26	1.16	6.88	75.98
Lymnaeidae	1.37	3.56	1.25	5.74	81.73
Coenagrionidae	1.47	3.48	1.29	5.63	87.35
Acarina	1.13	1.91	0.77	3.08	90.43

Group Autumn 2010

Average similarity: 57.66

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
DugesIIDae	3.48	19.10	6.71	33.12	33.12
Physidae	2.47	10.75	2.60	18.64	51.77
Oligochaeta	2.13	9.50	5.06	16.48	68.25
Glossiphoniidae	1.57	4.69	1.25	8.13	76.38
Hemicorduliidae	1.21	4.67	1.30	8.10	84.48
s-f Chironominae	1.48	3.05	0.73	5.28	89.76
Coenagrionidae	0.97	2.71	0.76	4.69	94.45

Group Spring 2010

Average similarity: 65.17

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	3.73	11.90	10.18	18.27	18.27
DugesIIDae	3.41	11.54	6.05	17.71	35.98
s-f Chironominae	3.16	8.22	1.99	12.61	48.58
Acarina	1.88	5.33	3.24	8.18	56.76
Coenagrionidae	2.01	4.86	1.31	7.46	64.22
Hydrobiidae	2.23	4.83	1.07	7.41	71.64
Hydroptilidae	1.33	4.39	6.28	6.74	78.38
Megapodagrionidae	1.40	3.91	6.32	6.00	84.37
Oligochaeta	1.14	2.85	1.32	4.38	88.75
Lymnaeidae	1.06	1.88	0.78	2.88	91.63

Group Autumn 2011

Average similarity: 60.20

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
DugesIIDae	3.46	18.21	3.73	30.24	30.24
Oligochaeta	2.51	12.10	3.22	20.11	50.35
Glossiphoniidae	2.20	9.62	2.83	15.98	66.33
Physidae	1.96	7.85	1.33	13.04	79.37
Acarina	1.24	5.12	1.14	8.50	87.86
Coenagrionidae	1.15	2.19	0.77	3.64	91.50

SIMPER Buffalo Creek 2005 – 2011

Data worksheet

Name: Buffalo Ck Sqrt
 Data type: Abundance
 Sample selection: All
 Variable selection: All

Parameters

Resemblance: S17 Bray Curtis similarity
 Cut off for low contributions: 90.00%

Factor Groups

Sample	Season Year
S4	Autumn 2005
S4	Spring 2005
S4	Autumn 2006
S4	Autumn 2007
S4	Spring 2007
S4	Autumn 2008
S4	Spring 2008
S4	Autumn 2009
S4	Spring 2009
S4	Autumn 2010
S4	Spring 2010
S4	Autumn 2011

Group Autumn 2005

Average similarity: 75.60

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Megapodagrionidae	3.98	7.74	6.17	10.23	10.23
s-f Chironominae	4.20	7.21	13.30	9.54	19.77
Notonectidae	3.21	7.16	10.08	9.47	29.24
Oligochaeta	3.21	7.06	6.27	9.34	38.58
Coenagrionidae	2.98	5.47	10.81	7.24	45.82
Hydrobiidae	2.90	4.86	7.76	6.42	52.24
Dugesidae	2.23	4.69	14.61	6.21	58.45
Corbiculidae	2.40	4.56	5.24	6.03	64.48
Hemicorduliidae	3.12	4.35	1.26	5.75	70.22
Planorbidae	1.52	3.20	10.08	4.24	74.46
s-f Tanypodinae	1.82	3.20	10.08	4.24	78.69
Physidae	1.82	3.00	2.44	3.96	82.66
Acarina	1.28	2.61	3.18	3.45	86.11
Stratiomyidae	1.38	2.57	5.00	3.40	89.51
Glossiphoniidae	1.28	2.57	5.00	3.40	92.90

Group Spring 2005

Average similarity: 66.33

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	6.42	14.47	9.77	21.81	21.81
Oligochaeta	4.67	11.44	24.58	17.24	39.05
Physidae	3.67	8.06	4.71	12.15	51.20
Hydrobiidae	3.19	7.89	6.77	11.89	63.10
Scyphacidae	1.72	4.18	7.44	6.30	69.40
Dugesidae	1.87	4.16	13.86	6.27	75.67
Corbiculidae	2.18	3.82	2.22	5.75	81.42
Notonectidae	1.67	3.67	2.28	5.53	86.95
Libellulidae	2.01	3.11	6.34	4.68	91.64

Group Autumn 2006

Average similarity: 74.85

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	7.70	15.83	20.12	21.15	21.15
Notonectidae	3.55	7.57	11.10	10.11	31.26
Libellulidae	2.92	5.38	3.70	7.19	38.44
Physidae	2.57	5.35	10.99	7.15	45.59
Coenagrionidae	2.72	5.21	13.45	6.96	52.55
Corbiculidae	2.37	4.86	7.03	6.49	59.04
Oligochaeta	2.57	4.72	3.44	6.31	65.36
Megapodagrionidae	2.41	4.24	2.37	5.67	71.02
Dugesiidae	1.82	3.91	49.03	5.22	76.25
Aeshnidae	1.97	3.91	49.03	5.22	81.47
Hemicorduliidae	2.34	3.63	5.00	4.85	86.31
s-f Orthocladiinae	1.61	3.19	49.03	4.26	90.58

Group Autumn 2007

Average similarity: 69.52

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.26	9.71	5.15	13.97	13.97
Notonectidae	3.23	7.79	6.96	11.21	25.18
Physidae	3.23	6.36	2.24	9.14	34.32
Hydrobiidae	2.51	5.25	2.69	7.55	41.88
Hemicorduliidae	2.47	5.09	2.72	7.33	49.20
Megapodagrionidae	2.06	4.40	6.84	6.33	55.54
Hydroptilidae	1.93	4.09	4.75	5.88	61.42
s-f Tanypodinae	1.71	3.53	3.54	5.07	66.49
Isostictidae	1.64	3.19	4.05	4.59	71.08
Lymnaeidae	1.60	3.15	4.78	4.53	75.61
Aeshnidae	1.64	2.85	1.35	4.10	79.71
Coenagrionidae	1.57	2.28	1.24	3.28	83.00
Dugesiidae	1.43	1.76	0.79	2.53	85.53
Baetidae	1.70	1.71	0.48	2.46	87.99
Stratiomyidae	1.02	1.68	1.33	2.42	90.41

Group Spring 2007

Average similarity: 65.17

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.63	14.76	4.83	22.65	22.65
Physidae	3.92	14.38	11.29	22.07	44.72
Hydrobiidae	2.54	8.68	5.00	13.33	58.05
Megapodagrionidae	1.97	5.33	2.53	8.17	66.22
Oligochaeta	1.68	5.09	2.75	7.81	74.03
Notonectidae	1.43	4.64	4.77	7.12	81.15
Isostictidae	1.51	2.99	0.78	4.58	85.73
Coenagrionidae	1.01	1.86	0.77	2.85	88.58
s-f Tanypodinae	0.97	1.81	0.77	2.78	91.36

Group Autumn 2008

Average similarity: 63.54

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	3.63	9.91	6.68	15.60	15.60
Notonectidae	3.12	9.02	3.62	14.19	29.79
Physidae	3.11	6.90	2.31	10.86	40.65
Megapodagrionidae	2.49	6.89	3.97	10.85	51.50
Dugesiidae	2.10	6.11	4.67	9.62	61.12
Hydrobiidae	2.61	5.21	1.25	8.20	69.32
Hydroptilidae	2.31	4.66	1.15	7.34	76.66
s-f Orthocladiinae	2.39	4.48	1.24	7.04	83.70
Planorbidae	1.33	1.85	0.71	2.91	86.61
Aeshnidae	0.93	1.51	0.75	2.38	88.99
Oligochaeta	0.96	1.37	0.77	2.16	91.14

Group Spring 2008

Average similarity: 65.89

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	3.74	19.20	4.53	29.13	29.13
s-f Chironominae	3.71	17.49	4.38	26.54	55.67
Hydrobiidae	3.12	15.80	3.82	23.98	79.65
Megapodagrionidae	1.19	4.54	1.29	6.89	86.54
Oligochaeta	1.26	4.07	1.28	6.18	92.72

Group Autumn 2009

Average similarity: 68.72

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	3.01	9.79	5.02	14.24	14.24
s-f Chironominae	2.86	8.96	3.23	13.04	27.28
Hydrobiidae	2.97	8.61	3.50	12.53	39.81
Notonectidae	2.35	7.03	3.56	10.23	50.04
Megapodagrionidae	2.08	6.23	5.89	9.06	59.11
s-f Orthoclaadiinae	1.54	4.90	3.79	7.14	66.24
Coenagrionidae	1.63	4.57	4.41	6.65	72.90
Dugesidae	1.67	3.69	1.31	5.36	78.26
Hydroptilidae	1.50	3.25	1.24	4.72	82.98
Planorbidae	1.55	3.02	0.78	4.39	87.38
Isostictidae	0.90	2.48	1.34	3.61	90.98

Group Spring 2009

Average similarity: 68.93

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	3.41	11.88	3.46	17.24	17.24
Hydrobiidae	3.23	11.39	6.09	16.53	33.77
Megapodagrionidae	2.52	7.62	2.47	11.05	44.82
Physidae	2.36	7.44	2.02	10.79	55.61
Sphaeriidae	1.82	5.66	4.05	8.21	63.82
Planorbidae	1.55	5.46	4.11	7.92	71.74
Oligochaeta	1.24	4.24	5.85	6.15	77.89
Lymnaeidae	1.14	4.15	10.80	6.02	83.91
Notonectidae	1.09	3.07	1.31	4.46	88.37
Hydroptilidae	1.74	2.90	0.75	4.21	92.57

Group Autumn 2010

Average similarity: 56.43

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	2.88	9.63	4.34	17.07	17.07
Notonectidae	2.69	8.86	3.26	15.69	32.76
Hydrobiidae	3.06	8.34	2.42	14.79	47.55
Planorbidae	1.58	4.46	4.16	7.90	55.45
Physidae	1.45	3.19	1.24	5.66	61.11
Megapodagrionidae	1.48	2.67	0.77	4.73	65.84
Oligochaeta	1.14	2.34	1.34	4.14	69.98
Coenagrionidae	1.91	2.26	0.70	4.00	73.98
Dugesidae	1.44	2.10	0.74	3.73	77.71
Veliidae	1.47	1.73	0.73	3.07	80.78
s-f Tanypodinae	1.18	1.61	0.72	2.85	83.62
Aeshnidae	1.41	1.54	0.77	2.73	86.36
Hemicorduliidae	1.09	1.38	0.76	2.45	88.80
Sphaeriidae	1.29	1.37	0.48	2.43	91.23

Group Spring 2010

Average similarity: 60.60

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.90	21.28	3.90	35.12	35.12
Hydrobiidae	2.49	11.38	3.60	18.77	53.89
Physidae	2.78	8.81	3.59	14.54	68.43
Megapodagrionidae	1.59	4.41	1.32	7.28	75.71
Oligochaeta	1.38	3.46	1.29	5.72	81.42
Notonectidae	1.04	2.95	0.76	4.86	86.29
s-f Tanypodinae	0.97	2.07	0.74	3.42	89.71
Dugesidae	0.67	1.69	0.78	2.79	92.50

Group Autumn 2011

Average similarity: 62.15

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Coenagrionidae	2.54	6.82	3.65	10.97	10.97
Notonectidae	2.55	6.57	2.46	10.57	21.54
Hemicorduliidae	2.69	6.43	2.34	10.35	31.89
Hydrobiidae	2.65	6.17	1.28	9.92	41.81
s-f Chironominae	2.37	5.38	2.46	8.66	50.47
Planorbidae	1.89	4.80	2.46	7.73	58.20
Megapodagrionidae	1.89	4.56	4.57	7.34	65.54
Gerridae	1.56	4.01	3.84	6.46	72.00
Dugesiiidae	1.66	3.40	1.16	5.47	77.47
Sphaeriidae	1.61	2.94	1.16	4.73	82.20
Oligochaeta	1.50	2.92	1.26	4.69	86.90
Lymnaeidae	1.04	2.38	1.30	3.83	90.73

SIMPER Porters Creek 2005 – 2011

Data worksheet

Name: Porters Ck Sqrt

Data type: Abundance

Sample selection: All

Variable selection: All

Parameters

Resemblance: S17 Bray Curtis similarity

Cut off for low contributions: 90.00%

Factor Groups

Sample	Season Year
S3	Autumn 2005
S3	Spring 2005
S3	Autumn 2006
S3	Autumn 2007
S3	Spring 2007
S3	Autumn 2008
S3	Spring 2008
S3	Autumn 2009
S3	Spring 2009
S3	Autumn 2010
S3	Spring 2010
S3	Autumn 2011

Group Autumn 2005

Average similarity: 76.82

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	6.51	12.89	10.12	16.78	16.78
Hydrobiidae	4.59	8.30	7.22	10.80	27.58
Isostictidae	4.18	8.07	10.67	10.50	38.08
Hemicorduliidae	2.89	5.93	55.42	7.71	45.80
Physidae	3.09	5.90	11.52	7.68	53.47
Megapodagrionidae	3.01	5.40	10.58	7.03	60.50
Coenagrionidae	2.83	4.64	2.61	6.04	66.54
Planorbidae	2.30	4.51	7.69	5.87	72.40
Oligochaeta	2.45	4.13	4.56	5.38	77.79
Glossiphoniidae	2.10	3.54	3.45	4.61	82.40
s-f Tanypodinae	2.39	3.38	4.51	4.40	86.79
Aeshnidae	1.41	2.96	55.42	3.86	90.65

Group Spring 2005

Average similarity: 72.69

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	10.09	23.35	7.12	32.12	32.12
Hydrobiidae	4.74	10.08	8.73	13.86	45.98
Oligochaeta	2.68	5.99	19.38	8.24	54.22
Isostictidae	2.63	5.99	19.38	8.24	62.46
Physidae	2.49	5.65	4.31	7.77	70.24
Glossiphoniidae	1.99	4.63	6.74	6.37	76.61
Libellulidae	2.22	4.33	2.89	5.95	82.56
Corixidae	1.80	2.91	3.64	4.00	86.56
Eropodellidae	1.28	2.88	4.62	3.97	90.53

Group Autumn 2006

Average similarity: 71.92

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	4.07	8.77	4.85	12.20	12.20
Coenagrionidae	3.33	7.27	5.23	10.10	22.30
Megapodagrionidae	3.64	7.01	7.28	9.75	32.05
Isostictidae	3.18	6.57	18.65	9.14	41.19
Oligochaeta	2.58	6.08	18.65	8.46	49.65
Hemicorduliidae	2.69	5.55	18.65	7.72	57.37
Atyidae	2.74	5.55	18.65	7.72	65.09
Glossiphoniidae	2.85	5.24	2.96	7.29	72.38
Aeshnidae	2.20	4.69	10.46	6.53	78.91
Physidae	1.93	3.76	15.62	5.23	84.14
Libellulidae	1.66	3.11	2.72	4.32	88.46
s-f Tanypodinae	2.52	2.58	0.58	3.58	92.04

Group Autumn 2007

Average similarity: 71.28

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	4.72	10.58	6.69	14.84	14.84
Physidae	2.61	5.88	5.12	8.24	23.09
Notonectidae	2.63	5.79	5.28	8.12	31.21
Isostictidae	2.79	5.76	3.27	8.08	39.29
s-f Chironominae	2.78	5.51	4.23	7.73	47.02
Coenagrionidae	2.63	5.44	3.69	7.64	54.66
Megapodagrionidae	2.45	5.12	4.50	7.18	61.84
Hemicorduliidae	2.37	4.88	3.60	6.85	68.68
Libellulidae	2.15	4.36	3.65	6.11	74.80
Hydroptilidae	1.89	3.85	4.08	5.41	80.20
Atyidae	2.15	3.77	2.18	5.29	85.49
s-f Orthocladiinae	1.72	2.70	1.17	3.79	89.28
s-f Tanypodinae	1.33	1.79	0.78	2.51	91.80

Group Spring 2007

Average similarity: 67.64

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.72	18.60	7.09	27.50	27.50
Hydrobiidae	3.74	12.08	4.54	17.86	45.36
Physidae	2.81	9.67	4.32	14.29	59.65
Oligochaeta	2.70	8.17	3.30	12.08	71.73
Megapodagrionidae	2.43	7.43	3.03	10.98	82.71
Isostictidae	1.45	3.67	1.28	5.42	88.13
Planorbidae	0.79	1.59	0.78	2.35	90.48

Group Autumn 2008

Average similarity: 59.84

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	3.77	14.45	6.36	24.15	24.15
Hydrobiidae	3.11	11.65	5.95	19.47	43.62
Megapodagrionidae	2.24	6.95	2.96	11.62	55.23
s-f Orthocladiinae	2.30	5.72	2.27	9.57	64.80
Notonectidae	1.87	5.31	3.06	8.88	73.68
Stratiomyidae	1.45	3.70	1.25	6.19	79.87
Oligochaeta	1.34	2.46	0.78	4.12	83.99
Physidae	1.20	2.10	0.73	3.51	87.50
Dugesidae	1.15	2.00	0.70	3.33	90.84

Group Spring 2008

Average similarity: 52.26

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Physidae	2.92	12.47	4.82	23.87	23.87
Oligochaeta	2.24	9.31	2.70	17.81	41.68
s-f Chironominae	2.57	7.72	0.77	14.77	56.44
Hydrobiidae	2.22	6.22	1.09	11.90	68.35
Notonectidae	1.26	4.09	1.34	7.82	76.17
Megapodagrionidae	1.09	4.04	1.19	7.72	83.89
Planorbidae	1.00	3.25	1.33	6.23	90.12

Group Autumn 2009

Average similarity: 58.24

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	3.36	13.95	5.78	23.95	23.95
s-f Chironominae	3.22	12.90	5.48	22.15	46.10
s-f Orthocladiinae	2.18	7.88	2.89	13.53	59.63
Megapodagrionidae	1.59	5.08	1.32	8.72	68.36
Coenagrionidae	1.30	3.40	1.31	5.83	74.19
Oligochaeta	1.07	3.25	1.32	5.58	79.77
Notonectidae	1.21	2.64	0.79	4.53	84.30
Isostictidae	1.13	2.26	0.77	3.89	88.18
Antipodoecidae	0.87	2.18	0.78	3.75	91.93

Group Spring 2009

Average similarity: 55.42

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	3.68	14.22	2.90	25.66	25.66
Hydrobiidae	3.16	13.04	3.67	23.52	49.18
Notonectidae	1.77	5.30	0.92	9.57	58.75
Megapodagrionidae	1.38	4.07	1.28	7.35	66.10
Planorbidae	1.60	4.00	1.27	7.22	73.32
Physidae	1.54	3.96	1.26	7.15	80.47
Coenagrionidae	1.09	3.28	1.33	5.92	86.39
s-f Orthocladiinae	0.90	1.42	0.48	2.56	88.95
s-f Tanypodinae	0.81	1.23	0.48	2.22	91.17

Group Autumn 2010

Average similarity: 54.37

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.57	19.58	4.29	36.01	36.01
Hydrobiidae	3.34	13.16	2.30	24.20	60.21
Notonectidae	1.79	4.67	1.23	8.59	68.80
s-f Orthocladiinae	1.33	4.30	1.26	7.91	76.71
Physidae	0.90	3.05	1.28	5.60	82.31
Hydroptilidae	0.93	2.32	0.72	4.28	86.59
Acarina	0.97	2.13	0.76	3.91	90.50

Group Spring 2010

Average similarity: 66.54

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	3.76	16.37	4.47	24.60	24.60
Hydrobiidae	3.56	16.26	10.13	24.43	49.03
Notonectidae	2.02	7.40	2.94	11.12	60.15
Oligochaeta	1.61	7.24	9.07	10.88	71.03
s-f Orthocladiinae	1.78	5.65	1.28	8.48	79.51
Isostictidae	1.19	3.92	1.30	5.89	85.40
Hydroptilidae	1.09	3.74	1.29	5.62	91.02

Group Autumn 2011

Average similarity: 50.56

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	3.90	11.94	3.56	23.61	23.61
Notonectidae	2.21	6.96	4.51	13.76	37.37
s-f Chironominae	2.82	4.95	1.26	9.78	47.15
Coenagrionidae	2.14	3.79	1.22	7.50	54.64
Hemicorduliidae	1.95	3.77	1.26	7.46	62.10
Hydroptilidae	1.49	3.34	1.34	6.60	68.70
Acarina	1.97	3.21	0.96	6.36	75.06
Megapodagrionidae	1.25	2.46	1.30	4.87	79.93
Isostictidae	1.57	2.12	0.75	4.20	84.12
Stratiomyidae	1.12	1.63	0.75	3.23	87.35
Tipulidae	0.74	1.22	0.79	2.41	89.77
Veliidae	1.26	1.17	0.48	2.32	92.08

SIMPER Terrys Creek 2005 – 2011

Data worksheet

Name: Terrys Ck Sqrt
 Data type: Abundance
 Sample selection: All
 Variable selection: All

Parameters

Resemblance: S17 Bray Curtis similarity
 Cut off for low contributions: 90.00%

Factor Groups

Sample	Season Year
S1	Autumn 2005
S1	Spring 2005
S1	Autumn 2006
S1	Autumn 2007
S1	Spring 2007
S1	Autumn 2008
S1	Spring 2008
S1	Autumn 2009
S1	Spring 2009
S1	Autumn 2010
S1	Spring 2010
S1	Autumn 2011

Group Autumn 2005

Average similarity: 69.53

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Megapodagrionidae	4.28	8.68	8.63	12.48	12.48
Hydrobiidae	3.36	7.27	14.80	10.45	22.93
s-f Chironominae	3.72	5.63	2.01	8.10	31.03
Isostictidae	2.58	5.54	13.25	7.97	39.00
Oligochaeta	2.90	5.12	5.83	7.36	46.36
Dugesidae	2.73	4.89	5.00	7.04	53.39
Physidae	2.46	4.70	3.87	6.76	60.16
Corbiculidae	2.38	4.28	8.30	6.15	66.31
s-f Tanypodinae	2.77	4.11	14.09	5.91	72.22
Notonectidae	2.46	4.09	2.80	5.89	78.11
Hemicorduliidae	2.78	3.94	3.39	5.67	83.78
Planorbidae	1.80	3.62	5.83	5.20	88.98
Glossiphoniidae	1.38	2.60	3.43	3.74	92.72

Group Spring 2005

Average similarity: 64.98

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	5.63	13.05	19.49	20.08	20.08
Physidae	3.14	6.76	7.02	10.41	30.49
Oligochaeta	3.17	6.51	11.44	10.02	40.52
Megapodagrionidae	2.93	6.38	16.31	9.82	50.33
Isostictidae	2.57	5.61	6.89	8.63	58.96
Corbiculidae	2.05	3.57	12.60	5.50	64.46
s-f Tanypodinae	2.26	3.50	1.77	5.38	69.84
Dugesidae	1.52	3.33	20.76	5.13	74.97
Acarina	1.88	3.10	2.59	4.78	79.74
Notonectidae	1.47	2.70	3.90	4.15	83.90
Libellulidae	2.45	2.70	0.58	4.15	88.04
Hydrobiidae	2.35	2.48	0.58	3.82	91.86

Group Autumn 2006

Average similarity: 72.76

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Megapodagrionidae	4.95	8.98	18.42	12.34	12.34
Hemicorduliidae	4.41	8.71	18.55	11.98	24.31
Oligochaeta	4.04	8.33	18.95	11.45	35.77
Hydrobiidae	3.58	5.51	2.85	7.58	43.34
Notonectidae	2.23	4.48	14.83	6.16	49.50
Dugesiidae	2.63	4.25	2.15	5.85	55.35
Gerridae	1.73	3.74	15.97	5.14	60.48
Physidae	2.33	3.70	1.32	5.08	65.57
s-f Tanypodinae	2.44	3.70	1.32	5.08	70.65
s-f Chironominae	2.67	3.68	2.86	5.06	75.71
Coenagrionidae	2.10	3.62	3.89	4.98	80.69
Isostictidae	1.80	3.45	5.69	4.75	85.44
Acarina	1.52	3.05	15.97	4.19	89.63
Libellulidae	1.47	2.44	5.69	3.36	92.99

Group Autumn 2007

Average similarity: 65.81

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	4.10	9.26	5.36	14.08	14.08
Megapodagrionidae	3.47	8.29	5.44	12.60	26.68
s-f Chironominae	3.19	7.79	4.33	11.84	38.52
Dugesiidae	2.60	5.83	2.72	8.85	47.37
Physidae	2.59	5.46	2.48	8.29	55.66
Notonectidae	2.19	5.03	6.23	7.64	63.30
Oligochaeta	1.93	4.37	3.50	6.64	69.94
s-f Tanypodinae	2.09	3.92	2.41	5.96	75.90
Hemicorduliidae	1.68	3.07	1.30	4.66	80.56
Isostictidae	1.38	2.07	1.31	3.15	83.71
s-f Orthoclaadiinae	1.29	1.75	0.77	2.66	86.36
Libellulidae	0.97	1.40	0.76	2.13	88.50
Coenagrionidae	1.06	1.28	0.76	1.95	90.45

Group Spring 2007

Average similarity: 64.85

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.77	14.86	7.37	22.92	22.92
Hydrobiidae	3.55	10.82	3.75	16.68	39.60
Megapodagrionidae	2.98	8.85	4.52	13.66	53.26
Physidae	2.57	7.67	4.19	11.83	65.08
Dugesiidae	2.43	6.32	2.15	9.75	74.83
Oligochaeta	1.85	4.55	1.27	7.01	81.85
Hemicorduliidae	1.52	2.87	1.21	4.42	86.27
s-f Tanypodinae	1.00	2.33	1.35	3.60	89.87
Sphaeriidae	1.12	1.98	0.73	3.06	92.93

Group Autumn 2008

Average similarity: 66.65

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	3.50	14.68	4.25	22.02	22.02
Megapodagrionidae	3.03	12.52	7.16	18.78	40.81
Notonectidae	2.29	9.65	6.24	14.48	55.29
Physidae	2.35	8.98	5.21	13.48	68.77
Dugesiidae	1.66	6.76	7.68	10.14	78.90
Oligochaeta	1.37	4.27	1.31	6.40	85.30
s-f Chironominae	1.35	3.72	1.29	5.59	90.89

Group Spring 2008

Average similarity: 61.90

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	3.61	13.31	7.64	21.51	21.51
Physidae	3.19	11.33	6.91	18.30	39.81
Megapodagrionidae	3.06	10.70	4.47	17.29	57.10
s-f Chironominae	2.90	8.20	1.27	13.25	70.35
Oligochaeta	2.07	6.47	3.13	10.46	80.81
Dugesiidae	1.34	3.47	1.32	5.61	86.42
Sphaeriidae	1.44	2.93	1.29	4.74	91.15

Group Autumn 2009

Average similarity: 61.96

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Megapodagrionidae	3.52	12.06	7.66	19.46	19.46
Hydrobiidae	3.60	11.81	5.04	19.05	38.51
Notonectidae	2.00	5.78	2.89	9.33	47.84
Isostictidae	1.71	5.05	3.97	8.15	55.99
Oligochaeta	1.55	4.68	4.73	7.56	63.55
Physidae	1.80	4.17	1.12	6.73	70.28
Dugesidae	1.60	3.93	1.31	6.34	76.62
s-f Tanypodinae	1.29	2.92	1.24	4.71	81.33
Gerridae	0.74	1.53	0.78	2.46	83.79
Coenagrionidae	0.94	1.44	0.78	2.32	86.12
Hemicorduliidae	0.86	1.43	0.77	2.31	88.43
s-f Chironominae	0.71	0.99	0.48	1.59	90.02

Group Spring 2009

Average similarity: 66.93

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	3.58	11.58	4.65	17.30	17.30
Megapodagrionidae	3.49	11.30	5.22	16.89	34.19
s-f Chironominae	3.03	10.22	5.42	15.27	49.46
s-f Tanypodinae	2.28	6.67	4.35	9.97	59.43
Dugesidae	1.88	5.70	3.74	8.52	67.95
Physidae	1.73	5.51	3.75	8.23	76.18
Oligochaeta	1.87	5.40	2.58	8.06	84.24
Isostictidae	1.38	4.58	3.88	6.85	91.09

Group Autumn 2010

Average similarity: 59.25

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
Hydrobiidae	4.61	18.14	7.36	30.61	30.61
Atyidae	2.68	9.50	2.43	16.04	46.64
Notonectidae	1.97	6.14	2.41	10.36	57.00
Dugesidae	2.06	4.92	1.11	8.31	65.31
Megapodagrionidae	1.96	4.78	1.32	8.07	73.38
Oligochaeta	1.47	4.14	1.33	7.00	80.37
Physidae	1.21	2.87	1.28	4.84	85.21
Elmidae	0.86	1.70	0.77	2.87	88.09
s-f Chironominae	0.98	1.66	0.78	2.81	90.90

Group Spring 2010

Average similarity: 64.47

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.18	14.38	6.02	22.31	22.31
Hydrobiidae	3.62	13.08	7.40	20.29	42.60
Megapodagrionidae	2.73	9.32	3.53	14.46	57.06
Physidae	2.62	9.30	10.25	14.42	71.48
Oligochaeta	1.55	3.72	1.35	5.78	77.26
s-f Tanypodinae	1.44	3.10	1.27	4.81	82.07
Dugesidae	1.02	2.76	1.33	4.28	86.35
Acarina	1.21	2.11	0.73	3.27	89.62
Isostictidae	0.96	1.87	0.73	2.90	92.52

Group Autumn 2011

Average similarity: 66.06

Species	Av.Abund	Av.Sim	Sim/SD	Contrib%	Cum.%
s-f Chironominae	4.43	13.43	5.80	20.33	20.33
Hydrobiidae	3.67	9.24	4.16	13.99	34.32
Notonectidae	2.97	9.03	3.91	13.67	47.99
Physidae	2.94	8.19	10.99	12.40	60.39
Dugesidae	2.28	6.60	7.67	10.00	70.38
Glossiphoniidae	1.96	4.64	2.51	7.02	77.41
Oligochaeta	1.94	4.49	2.43	6.80	84.20
Stratiomyidae	1.14	3.40	7.55	5.14	89.34
Hemicorduliidae	1.22	1.42	0.58	2.15	91.49

Appendix 6 BIOENV output

ALL FIVE CREEKS Spring 2010

Data worksheet

Name: Data1
 Data type: Environmental
 Sample selection: All
 Variable selection: All

Resemblance worksheet

Name: All five Cks(2)
 Data type: Similarity
 Selection: All

Parameters

Rank correlation method: Spearman
 Method: BIOENV
 Maximum number of variables: 5
 Resemblance:
 Analyse between: Samples
 Resemblance measure: D1 Euclidean distance

Variables

- 1 Log10 Faecal Coliforms
- 2 Log10 Ammonia
- 3 Log10 Oxidised Nitrogen
- 4 Log10 Total Phosphorus
- 5 Log10 Total Kjeldahl Nitrogen
- 6 Alkalinity mg
- 7 Log10 Turbidity
- 8 Log10 Conductivity
- 9 Log10 Total Dissolved Solids
- 10 pH
- 11 Dissolved Oxygen DO mg/L
- 12 Temperature OC
- 13 Rainfall
- 14 Altitude
- 15 Bedrock
- 16 Boulder
- 17 Cobble
- 18 Total Length Pipe
- 19 No. Outlets
- 20 Catchment Area
- 21 Ratio TLP/CA
- 22 Ratio NO/CA

Best results

No. Vars	Corr.	Selections
5	0.396	3,8,11,21,22
5	0.392	3,8,11,17,22
5	0.389	3,9,11,21,22
5	0.387	3,9,11,17,22
5	0.384	3,11,17,21,22
5	0.384	3,8,10,21,22
5	0.384	3,6,11,21,22
5	0.382	3,8,10,17,22
5	0.381	3,8,17,21,22
5	0.381	8,10,11,17,22

ARCHERS CK Spring 2010

Data worksheet

Name: Data1

Data type: Environmental

Sample selection: All

Variable selection: All

Resemblance worksheet

Name: Archers Ck(2)

Data type: Similarity

Selection: All

Parameters

Rank correlation method: Spearman

Method: BIOENV

Maximum number of variables: 5

Resemblance:

Analyse between: Samples

Resemblance measure: D1 Euclidean distance

Variables

- 1 Log10 Faecal Coliforms
- 2 Log10 Ammonia
- 3 Log10 Oxidised Nitrogen
- 4 Log10 Total Phosphorus
- 5 Log10 Total Kjeldahl Nitrogen
- 6 Log10 Alkalinity
- 7 Log10 Turbidity
- 8 Log10 Conductivity
- 9 Log10 Total Dissolved Solids
- 10 pH
- 11 Dissolved Oxygen
- 12 Temperature
- 13 Rainfall

Best results

No.Vars Corr. Selections

- | | | |
|---|-------|------------|
| 5 | 0.237 | 3,4,6,7,12 |
| 4 | 0.232 | 3,4,6,7 |
| 5 | 0.220 | 3,4,6-8 |
| 4 | 0.218 | 3,4,7,12 |
| 5 | 0.218 | 4-7,12 |
| 3 | 0.218 | 4,5,7 |
| 4 | 0.218 | 4,5,7,12 |
| 2 | 0.217 | 4,7 |
| 5 | 0.217 | 3-7 |
| 5 | 0.215 | 3,4,6,7,13 |

BUFFALO CK Spring 2010

Data worksheet

Name: Data1
 Data type: Environmental
 Sample selection: All
 Variable selection: All

Resemblance worksheet

Name: Buffalo Ck(2)
 Data type: Similarity
 Selection: All

Parameters

Rank correlation method: Spearman
 Method: BIOENV
 Maximum number of variables: 5
 Resemblance:
 Analyse between: Samples
 Resemblance measure: D1 Euclidean distance

Variables

- 1 Log10 Faecal Coliforms
- 2 Log10 Ammonia
- 3 Log10 Oxidised Nitrogen
- 4 Log10 Total Phosphorus
- 5 Log10 Total Kjeldahl Nitrogen
- 6 Alkalinity
- 7 Log10 Turbidity
- 8 Log10 Conductivity
- 9 Log10 Total Dissolved Solids
- 10 pH
- 11 Dissolved Oxygen
- 12 Temperature
- 13 Rainfall

Best results

No. Vars	Corr.	Selections
5	0.243	3-5,7,13
5	0.242	3-5,7,9
5	0.241	3-5,9,11
5	0.240	3-5,7,8
5	0.239	3-5,11,13
5	0.238	3-5,7,11
5	0.236	3-5,8,11
5	0.234	3,5,7,11,13
5	0.234	1,3-5,11
5	0.234	3,5,7,9,13

PORTERS CK Spring 2010

Data worksheet

Name: Data1

Data type: Environmental

Sample selection: All

Variable selection: All

Resemblance worksheet

Name: Porters Ck(2)

Data type: Similarity

Selection: All

Parameters

Rank correlation method: Spearman

Method: BIOENV

Maximum number of variables: 5

Resemblance:

Analyse between: Samples

Resemblance measure: D1 Euclidean distance

Variables

- 1 Log10 Faecal Coliforms
- 2 Log10 Ammonia
- 3 Log10 Oxidised Nitrogen
- 4 Log10 Total Phosphorus
- 5 Log10 Total Kjeldahl Nitrogen
- 6 Alkalinity
- 7 Log10 Turbidity
- 8 Log10 Conductivity
- 9 Log10 Total Dissolved Solids
- 10 pH
- 11 Dissolved Oxygen
- 12 Temperature
- 13 Rainfall

Best results

No. Vars Corr. Selections

- | | | |
|---|-------|----------|
| 5 | 0.364 | 5,7-9,13 |
| 4 | 0.361 | 5,7-9 |
| 5 | 0.361 | 1,5,7-9 |
| 5 | 0.360 | 2,7-9,13 |
| 4 | 0.359 | 2,7-9 |
| 3 | 0.357 | 5,8,9 |
| 4 | 0.357 | 5,8,9,13 |
| 5 | 0.356 | 5-9 |
| 5 | 0.354 | 1,2,7-9 |
| 5 | 0.352 | 2,6-9 |

SHRIMPTONS CK Spring 2011

Data worksheet

Name: Data1
 Data type: Environmental
 Sample selection: All
 Variable selection: All

Resemblance worksheet

Name: Shrimptons Ck(2)
 Data type: Similarity
 Selection: All

Parameters

Rank correlation method: Spearman
 Method: BIOENV
 Maximum number of variables: 5
 Resemblance:
 Analyse between: Samples
 Resemblance measure: D1 Euclidean distance

Variables

- 1 Log10 Faecal Coliforms
- 2 Log10 Ammonia
- 3 Log10 Oxidised Nitrogen
- 4 Log10 Total Phosphorus
- 5 Log10 Total Kjeldahl Nitrogen
- 6 Alkalinity
- 7 Log10 Turbidity
- 8 Log10 Conductivity
- 9 Log10 Total Dissolved Solids
- 10 pH
- 11 Dissolved Oxygen
- 12 Temperature
- 13 Rainfall

Best results

No. Vars	Corr.	Selections
5	0.280	1,7,9,11,12
4	0.275	1,9,11,12
3	0.274	1,9,11
4	0.269	1,7,9,11
5	0.266	1,2,9,11,12
5	0.263	1,8,9,11,12
2	0.262	9,11
5	0.261	1,7-9,11
5	0.258	1,2,7,9,11
5	0.255	1,7,8,11,12

TERRYS CK Spring 2011

Data worksheet

Name: Data1
 Data type: Environmental
 Sample selection: All
 Variable selection: All

Resemblance worksheet

Name: Terrys Ck(2)
 Data type: Similarity
 Selection: All

Parameters

Rank correlation method: Spearman
 Method: BIOENV
 Maximum number of variables: 5
 Resemblance:
 Analyse between: Samples
 Resemblance measure: D1 Euclidean distance

Variables

- 1 Log10 Faecal Coliforms
- 2 Log10 Ammonia
- 3 Log10 Oxidised Nitrogen
- 4 Log10 Total Phosphorus
- 5 Log10 Total Kjeldahl Nitrogen
- 6 Alkalinity
- 7 Log10 Turbidity
- 8 Log10 Conductivity
- 9 Log10 Total Dissolved Solids
- 10 pH
- 11 Dissolved Oxygen
- 12 Temperature
- 13 Rainfall

Best results

No. Vars	Corr.	Selections
2	0.260	6,13
3	0.258	3,6,13
4	0.255	3,6,8,13
2	0.255	3,13
5	0.252	4,6,8,11,13
5	0.251	3,6,8,11,13
3	0.250	3,8,13
2	0.250	4,13
3	0.249	6,8,13
3	0.248	3,4,13